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(54) Title: ANTIBACTERIAL TARGETS IN ALLOIOCOCCUS OTITIDIS

(57) Abstract: The present invention relates to the identification of polynucleotide sequences encoding polypeptides of Alloiococcus otitidis that are essential for the growth and survival of the bacteria. In particular, the invention relates to polypeptides encoded by the Alloiococcus otitidis open reading frames (ORFs), and to their use in pharmaceutical compositions, therapeutics, diagnostics and the like. The present invention also relates to methods for identifying pharmaceutical compounds that inhibit the activity of the polypeptides that are essential for the growth of Alloiococcus otitidis. to pharmaceutical compositions containing these compounds and to their use in treatment and amelioration of diseases caused by Alloiococcus otitidis

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# ANTIBACTERIAL TARGETS IN ALLOIOCOCCUS OTITIDIS FIELD OF THE INVENTION

The present invention relates to the genomic sequence of *Alloiococcus otitidis* and polynucleotide sequences encoding polypeptides of the Gram-positive bacterium, *Alloiococcus otitidis*. The invention also relates to polynucleotides and polynucleotides encoding polypeptides, preferably antigenic polypeptides, encoded by the *Alloiococcus otitidis* open reading frames and the uses thereof.

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### BACKGROUND OF THE INVENTION

Since the discovery of penicillin, the use of antibiotics to treat the ravages of bacterial infections has saved millions of lives. With the advent of these "miracle drugs," for a time it was popularly believed that humanity might, once and for all, be saved from the scourge of bacterial infections. In fact, during the 1980s and early 1990s, many large pharmaceutical companies cut back or eliminated antibiotics research and development. They believed that infectious disease caused by bacteria finally had been conquered and that markets for new drugs were limited. Unfortunately, this belief was overly optimistic. The tide is beginning to turn in favor of the bacteria, as reports of drug resistant bacteria become more frequent. The United States Centers for Disease Control and Prevention announced that one of the most powerful known antibiotics, vancomycin, was unable to treat an infection of the common bacterial pathogen, Staphylococcus aureus. This organism, commonly found in our environment, is responsible for many nosocomial infections. The import of this announcement becomes clear when one considers that vancomycin was used for years to treat infections caused by Staphylococcus species as well as other stubborn strains of bacteria. In short, bacteria are becoming resistant to our most powerful antibiotics. If this trend continues, it is conceivable that we will return to a time when what are presently considered minor bacterial infections are fatal diseases.

Over-prescription and improper prescription habits by some physicians have caused an indiscriminate increase in the availability of antibiotics to the public. The patients are also partly responsible, since they will often improperly use the drug,

thereby generating yet another population of bacteria that is resistant, in whole or in part, to traditional antibiotics.

The bacterial pathogens that have haunted humanity remain, in spite of the development of modern scientific practices to deal with the diseases that they cause. Drug resistant bacteria are now an increasing threat to the health of humanity. A new generation of antibiotics is needed to once again deal with the pending health threats that bacteria present.

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As more and more bacterial strains become resistant to the panel of available antibiotics, new antibiotics are required to treat infections. In the past, practitioners of pharmacology relied upon traditional methods of drug discovery to generate novel, safe and efficacious compounds for the treatment of disease. Traditional drug discovery methods involve blindly testing potential drug candidate- molecules, often selected at random, in the hope that one might prove to be an effective treatment for some disease. The process is painstaking and laborious, with no guarantee of success.

Newly emerging practices in drug discovery utilize a number of biochemical techniques to provide for directed approaches to creating new drugs, rather than discovering them at random. For example, gene sequences and proteins encoded thereby that are required for the proliferation of a cell or microorganism make excellent targets since exposure of bacteria to compounds active against these targets would result in the inactivation of the cell or microorganism. Once a target is identified, biochemical analysis of that target can be used to discover or to design molecules that interact with and alter the functions of the target. Use of physical and computational techniques to analyze structural and biochemical properties of targets in order to derive compounds that interact with such targets is called rational drug design and offers great potential. Thus, emerging drug discovery practices use molecular modeling techniques, combinatorial chemistry approaches, and other means to produce and screen and/or design large numbers of candidate compounds.

Nevertheless, while this approach to drug discovery is clearly the way of the future, problems remain. For example, the initial step of identifying molecular targets for investigation can be an extremely time consuming task. It may also be difficult to design molecules that interact with the target by using computer modeling techniques. Furthermore, in cases where the function of the target is not known or is

poorly understood, it may be difficult to design assays to detect molecules that interact with and alter the functions of the target. To improve the rate of novel drug discovery and development, methods of identifying important molecular targets in pathogenic cells or microorganisms and methods for identifying molecules that interact with and alter the functions of such molecular targets are urgently required.

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The present invention is directed to identifying important molecular targets in a recently identified bacteria, *Alloiococcus otitidis*, which has been implicated in otitis media with effusion (OME). Otitis media, an inflammatory disease of the middle ear, is the most frequent cause of visits to pediatricians' offices in the United States (Schappert, 1991). Approximately 80% of all children experience at least one episode of otitis media by the age of three (Klein, 1994). There are three main types of otitis media: Acute otitis media (AOM), otorrhea, and otitis media with effusion (OME). *Alloiococcus otitidis* has only been associated with otitis media with effusion (OME), but this may be due to the difficulty of its detection by standard bacterial culturing methods. Its detection in the effusions is likely due to the fact that the effusions are normally sterile and few or no competing bacterial species are isolated from them. Without the interference of faster growing nasophryngeal species, the culture plates can be incubated for the longer duration needed to detect *Alloiococcus otitidis* colonies.

Three other bacterial species are commonly isolated from middle ear effusions. These are nontypeable *Haemophilus influenzae*, *Moraxella catarrhalis*, and *Streptococcus pneumoniae*. One or more of these species have been found in one study to be associated with about 77% of all cases of OME using a PCR detection method (Post, 2000). This study did not include assaying for *Alloiococcus otitidis*, so a portion of the unaccounted cases may be due to this organism.

The bacterium *Alloiococcus otitidis* was first isolated from the middle ear fluids of 10 children in the Buffalo, NY area with persistent OME and characterized as a large catalase negative, Gram-positive cocci that tend to occur in clumps, often in tetrads. It is slow growing and requires 2 to 5 days at 37°C before colonies can be seen on sheep blood agar plates. The bacterium was named *Alloiococcus otitis* by Aguirre and Collins (1992), who showed that it was different from other known Grampositive species based on its 16S rRNA sequence. The bacterium's name has been

changed from *Alloiococcus otitis* to *Alloiococcus otitidis*. (Hendolin, et al., (1999), and Hendolin et al., (2000)).

Several studies of the epidemiology *Alloiococcus otitidis* indicate it is associated with otitis media with effusion. These are summarized in Table 1. These studies have been done using both culture and PCR techniques. The number of cases detected by culture, as might be expected from the fastidious growth requirements of the bacterium, was less than the number detected by PCR. Assuming that the bacterium is detected more accurately by the PCR method, the bacterium is detected in between 10 and 50% of patients with OME. This frequency suggests that this organism represents a significant public health problem. Consequently, there is a need for identifying gene targets in *Alloiococcus otitidis* for the development of anti-infectives. There is also a need for compositions for diagnosing *Alloiococcus otitidis* infection.

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TABLE 1: SUMMARY OF STUDIES INDICATING AN ASSOCIATION OF ALLOIOCOCCUS
OTITIDIS WITH OTITIS MEDIA WITH EFFUSION (OME).

% detected	Nª	Method	Reference
8	200	Culture	Faden & Dryja, J. Clin. Microbiol. 27:2488 (1989)
3	100	Culture	Sih et al., ICAAC (1992)
20	25	PCR	Hendolin et al., J. Clin. Microbiol. 35:2854 (1997)
50	12	PCR	Beswick, et al., Lancet 345:386 (1999)
42	67	PCR	Hendolin, et al., Pediatr. Infect. Dis. J. 18:860 (1999)
10	49	PCR	Hendolin et al., J. Clin. Microbiol. 38:125 (2000)

<sup>&</sup>lt;sup>a</sup> Number of persons in study.

### **SUMMARY OF INVENTION**

The present invention broadly relates to *Alloiococcus otitidis* genomic sequence. Particularly, the invention relates to newly identified polynucleotide open reading frames (ORFs) comprised within the genomic nucleotide sequence of *Alloiococcus otitidis*, and to polypeptides encoded by the ORFs. More particularly, the ORFs encode polypeptides that are essential for the growth and survivability of *Alloiococcus otitidis*.

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Thus, in certain aspects, the invention relates to Alloiococcus otitidis ORFs that encode Alloiococcus otitidis polypeptides that function as enzymes in various biosynthetic pathways in the bacterium. In one embodiment, the invention relates to a purified or isolated Alloiococcus otitidis nucleic acid sequence comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, wherein expression of said nucleic acid is essential for the proliferation of a cell. In a preferred embodiment the ORF selected from one of the odd numbered sequence listings set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105 encodes an essential gene. The essential gene and the polypeptide encoded by them include ACPS (holo-(acyl carrier protein) synthase), murF (UDP-Nacetylmuramoylalanyl-D-glutamyl-2,6-diamino pimelate-D-alanyl-D-alanyl ligase) murA-2 (UDP-N-acetylglucosamine 1-carboxyvinyltransferase), RpoE (DNA-directed RNA polymerase, delta subunit), rpoA (DNA-directed RNA polymerase alpha subunit), rpoC (RNA polymerase beta' subunit), rpoB (DNA-dependent RNA polymerase subunit beta), dnaB/C (DNA polymerase III delta prime subunit), gyrA (DNA gyrase A subunit), gyrB (DNA gyrase B subunit), dnaN (DNA polymerase III beta chain, folC-2 (folyl-polyglutamate synthetase), murE (UDP-N-acetylmuramoyl-Lalanyl-D-glutamyl-L-lysine Ligase), srtA (sortase), folC-1 (folyl-polyglutamate synthetase), folB (dihydroneopterin aldolase), folK (7,8-dihydro-6hydroxymethylpterin-pyrophosphokinase), mvaS (hydroxymethylglutaryl-CoA synthase), mvaA (3-hydroxy-3-methylglutaryl-coenzyme a reductase), murB (UDP-Nacetylglucosaminyl-3-enolpyruvate reductase), mvaK2 (phosphomevalonate kinase), mvaD (mevalonate diphosphate decarboxylase), mvaK1 (mevalonate kinase), coaA (pantothenate kinase), nadE (NAD+ synthase), murl, Glutamate racemase), folP (Dihydropteroate synthase), folA (dihydrofolate reductase), grlB (topoisomerase IV B

subunit), grIA (topoisomerase IV A subunit), rpoD (transcription initiation factor sigma), dnaG (DNA primase), era (GTP-binding protein), norA (drug-export protein), polC (DNA polymerase III, alpha subunit), obg (GTP-binding protein), yphC (similar to Escherichia coli GTP-binding protein Era), dnaE (DNA polymerase III, alpha subunit), coaBC (phosphopantothenoylcysteine synthetase/decarboxylase), holA (DNA polymerase III delta subunit), coaD (phosphopantetheine adenylyltransferase) ftsZ (Cell division protein ftsZ), ftsA (Cell division protein ftsA), murG (phospho-N-acetylmuramoyl-pentapeptide-transferase), murD (UDP-N-acetylmuramoylalanine D-glutamate ligase), nadD (nicotinic acid mononucleotide adenylyltransferase), coaE (dephospho-CoA kinase), murC (UDP-N-acetyl muramate-alanine ligase), fmhB FemX (factor essential for methicillin resistance), pcrA (ATP-dependent DNA helicase), murA-1 (UDP-N-acetylglucosamine 1-carboxyvinyltransferase), holB (DNA polymerase III delta' subunit) and dnaX (DNA polymerase III -gamma and tau subunits).

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In another embodiment, the invention relates to purified or isolated nucleic acid of *Alloiococcus otitidis* comprising a fragment of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, wherein said fragment is selected from the group consisting of fragments comprising at least 10, at least 20, at least 25, at least 30, at least 50 and more than 50 consecutive nucleotides of one of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

In yet another embodiment, the invention relates to a purified or isolated antisense nucleic acid comprising a nucleotide sequence complementary to at least a portion of an intragenic sequence, intergenic sequence, sequences spanning at least a portion of two or more genes, 5' noncoding region, or 3' noneoding region within an operon comprising a proliferation-required gene of Alloiococcus otitidis whose activity or expression is inhibited by an antisense nucleic acid and selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

In a nother embodiment, the invention relates to a purified or isolated nucleic acid comprising a nucleotide sequence having at least 70% identity to a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, fragments comprising at least 25 consecutive nucleotides selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, the nucleotide sequences complementary to one of odd numbered

sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, and the sequences complementary to fragments comprising at least 25 consecutive nucleotides of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

In another embodiment, the invention relates to a vector comprising a promoter operably linked to a nucleic acid encoding a polypeptide whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence of any one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

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In another embodiment, the invention relates to purified or isolated polypeptide of Alloiococcus otitidis comprising a polypeptide whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or a fragment selected from the group consisting of fragments comprising at least 5, at least 10, at least 20, at least 30, at least 40, at least 50, at least 60 or more than 60 consecutive amino acids of one of the said polypeptides.

In yet another embodiment, the invention relates to purified or isolated *Alloiococcus otitidis* polypeptide comprising a amino acid sequence having at least 25% amino acid identity to a polypeptide whose expression is inhibited by a nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or at least 25% amino acid identity to a fragment comprising at least 10, at least 20, at least 30, at least 40, at least 50, at least 60 or more than 60 consecutive amino acids of a polypeptide whose expression is inhibited by a nucleic acid comprising a nucleotide sequence selected from the group consisting of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

In one embodiment, the invention relates to a purified or isolated *Alloiococcus* otitidis polypeptide comprising selected from one of the even numbered sequences set forth in Seq. ID Nos: 2 to Seq. ID Nos: 106, wherein the polypeptide is essential for the proliferation of a cell..

In yet another embodiment, the invention relates to a method of producing an Alloiococcus otitidis polypeptide comprising introducing into a cell a vector comprising a promoter operably linked to a nucleic acid comprising a nucleotide sequence encoding a polypeptide whose expression is essential for the proliferation and viability of Alloiococcus otitidis, and which is inhibited by an antisense nucleic

acid, and which is selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

In yet another embodiment, the invention relates to a method of inhibiting the proliferation of *Alloiococcus otitidis* in an individual comprising inhibiting the activity or reducing the amount of a gene product whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105 or inhibiting the activity or reducing the amount of a nucleic acid encoding said gene product.

In a preferred embodiment, the invention relates to method for identifying a compound which influences the activity of an *Alloiococcus otitidis* gene product, which is required for proliferation, said gene product comprising a gene product whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, said method comprising: (a) contacting said gene product with a candidate compound; and (b) determining whether said compound influences the activity of said gene product.

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In a preferred embodiment, the invention relates to method for identifying a compound or an antisense nucleic acid having the ability to reduce activity or level of a *Alloiococcus otitidis* gene product, which is required for proliferation, said gene product comprising a gene product whose activity or expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, said method comprising the steps of: (a) contacting a target gene or RNA encoding said gene product with a candidate compound or antisense nucleic acid; and(b) measuring the activity of said target.

In yet another preferred embodiment, the invention relates to method for inhibiting cellular proliferation of *Alloiococcus otitidis* comprising introducing an effective amount of a compound with activity against a gene whose activity or expression is essential for cellular proliferation, and which is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or a compound with activity against the product of said gene into a population of *Alloiococcus otitidis* cells expressing said gene.

In a preferred embodiment, the invention relates to a composition comprising an effective concentration of an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or a proliferation-inhibiting portion thereof in a pharmaceutically acceptable carrier.

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In a preferred embodiment, the invention relates to method for identifying a compound having the ability to inhibit proliferation of *Alloiococcus otitidis* cell comprising: (a) identifying a homologue of a gene or gene product whose activity or level is inhibited by a nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, in a test cell, wherein said test cell is not *Alloiococcus otitidis*; (b) identifying an inhibitory nucleic acid sequence which inhibits the activity of said homologue in said test cell; (c) contacting said test cell with a sublethal level of said inhibitory nucleic acid, thus sensitizing said cell; (d) contacting the sensitized cell of step (c) with a compound; and (e) determining the degree to which said compound inhibits proliferation of said sensitized cell relative to a cell which does not contain said inhibitory nucleic acid.

In a preferred embodiment, the invention relates to a method for identifying a compound having activity against a biological pathway required for proliferation comprising: (a) sensitizing a cell by providing a sublethal level of an antisense nucleic acid complementary to a nucleic acid encoding a gene product required for proliferation, wherein the activity or expression of said gene product is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, in said cell to reduce the activity or amount of said gene product; (b) contacting the sensitized cell with a compound; and (c) determining the degree to which said compound inhibits the growth of said sensitized cell relative to a cell which does not contain said antisense nucleic acid.

In a preferred embodiment, the invention relates to a method for identifying a compound having the ability to inhibit one of the *Alloiococcus otitidis* polypeptides encoded by a polynucleotide selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, and which is essential for cellular proliferation comprising: (a) contacting a cell which expresses the polypeptide with the compound;

and (b) determining whether said compound reduces proliferation of said contacted cell by acting on said gene product.

In a preferred embodiment, the invention relates to a method for identifying a compound having the ability to inhibit one of the purified and isolated *Alloiococcus* otitidis polypeptides selected from one of the even numbered sequences set forth in Seq. ID No.: 2 to Seq. ID No.: 106, and which is essential for cellular proliferation comprising: (a) contacting the purified and isolated polypeptide with the compound *in* vitro in the presence or absence of a substrate, which is essential for the activity of the polypeptide; and (b) determining the effect of the compound on the polypeptide by measuring the effect of the polypeptide on the substrate.

In a preferred embodiment, the invention relates to a compound which interacts with an *Alloiococcus otitidis* polypeptide selected from one of the even numbered sequences set forth in Seq. ID No.: 2 to Seq. ID No.: 106 and inhibits its activity.

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In a preferred embodiment, the invention relates to a method for manufacturing an antimicrobial compound comprising the steps of screening one or more candidate compounds to identify a compound that reduces the activity or level of an *Alloiococcus otitidis* polypeptide selected from one of the even numbered sequences set forth in Seq. ID No.: 2 to Seq. ID No.: 106, said polypeptide comprising a gene product whose activity or expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105; and manufacturing the compound so identified.

In a preferred embodiment, the invention relates to a compound which inhibits proliferation of *Alloiococcus otitidis* by interacting with a gene encoding a polypeptide that is required for proliferation or with a polypeptide required for proliferation, wherein said polypeptide is selected from the group consisting of a gene product having at least 70% nucleotide sequence identity from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105, polypeptide encoded by a nucleic acid having at least 70% nucleotide sequence identity to a nucleic acid encoding a polypeptide whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105, a polypeptide having at

least 25% amino acid identity to a gene product whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105, a polypeptide encoded by a nucleic acid comprising a nucleotide sequence which hybridizes to a nucleic acid selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105 under stringent conditions, a gene product encoded by a nucleic acid comprising a nucleotide sequence which hybridizes to a nucleic acid selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105 under moderate conditions, and a gene product whose activity may be complemented by the gene product whose activity is inhibited by a nucleic acid selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105.

### **DETAILED DESCRIPTION OF THE INVENTION**

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### A. Definitions:

By "biological pathway" is meant any discrete cell function or process that is carried out by a gene product or a subset of gene products. Biological pathways include anabolic, catabolic, enzymatic, biochemical and metabolic pathways as well as pathways involved in the production of cellular structures such as cell walls. Biological pathways that are usually required for proliferation of cells or microorganisms include, but are not limited to, cell division, DNA synthesis and replication, RNA synthesis (transcription), protein synthesis (translation), protein processing, protein transport, fatty acid biosynthesis, electron transport chains, cell wall synthesis, cell membrane production, synthesis and maintenance, and the like.

By "inhibit activity of a gene or gene product" is meant having the ability to interfere with the function of a gene or gene product in such a way as to decrease expression of the gene, in such a way as to reduce the level or activity of a product of the gene or in such a way as to inhibit the interaction of the gene or gene product with other biological molecules required for its activity.

Agents which inhibit the activity of a gene include agents that inhibit transcription of the gene, agents that inhibit processing of the transcript of the gene, agents that reduce the stability of the transcript of the gene, and agents that inhibit translation of the mRNA transcribed from the gene. In microorganisms, agents which

inhibit the activity of a gene can act to decrease expression of the operon in which the gene resides or alter the folding or processing of operon RNA so as to reduce the level or activity of the gene product. The gene product can be a non- translated RNA such as ribosomal RNA, a translated RNA (mRNA) or the protein product resulting from translation of the gene mRNA. Of particular utility to the present invention are antisense RNAs that have activities against the operons or genes to which they specifically hybridze.

By "activity against a gene product" is meant having the ability to inhibit the function or to reduce the level or activity of the gene product in a cell. This includes, but is not limited to, inhibiting the enzymatic activity of the gene product or the ability of the gene product to interact with other biological molecules required for its activity, including inhibiting the gene product's assembly into a multimeric structure.

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By "activity against a protein" is meant having the ability to inhibit the function or to reduce the level or activity of the protein in a cell. This includes, but is not limited to, inhibiting the enzymatic activity of the protein or the ability of the protein to interact with other biological molecules required for its activity, including inhibiting the protein's assembly into a multimeric structure.

By "activity against a nucleic acid" is meant having the ability to inhibit the function or to reduce the level or activity of the nucleic acid in a cell. This includes, but is not limited to, inhibiting the ability of the nucleic acid interact with other biological molecules required for its activity, including inhibiting the nucleic acid's assembly into a multimeric structure.

By "activity against a gene" is meant having the ability to inhibit the function or expression of the gene in a cell. This includes, but is not limited to, inhibiting the ability of the gene to interact with other biological molecules required for its activity. By "activity against an operon" is meant having the ability to inhibit the function or reduce the level of one or more products of the operon in a cell. This includes, but is not limited to, inhibiting the enzymatic activity of one or more products of the operon or the ability of one or more products of the operon to interact with other biological molecules required for its activity.

By "antibiotic" is meant an agent which inhibits the proliferation of a cell or microorganism.

By "homologous coding nucleic acid" is meant a nucleic acid homologous to a nucleic acid encoding a gene product whose activity or level is inhibited by a nucleic acid selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105 or a portion thereof. In some embodiments, the homologous coding nucleic acid may have at least 97%, at least 95%, at least 90%, at least 85%, at least 80%, or at least 70% nucleotide sequence identity to a nucleotide sequence selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and fragments comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides thereof. In other embodiments the homologous coding nucleic acids may have at least 97%, at least 5 95%, at least 90%, at least 85%, at least 80%, or at least 70% nucleotide sequence identity to a nucleotide sequence selected from the group consisting of the nucleotide sequences complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and fragments comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides thereof. Identity may be measured using BLASTN version 2.0 with the default parameters or tBLASTX with the default parameters. (Altschul, S.F. et al. Gapped BLAST and PSI-BLAST: A New Generation of Protein Database Search Programs, Nucleic Acid Res. 25: 3389-3402 (1997)) Alternatively a "homologuous coding nucleic acid" could be identified by membership of the gene of interest to a functional orthologue cluster. All other members of that orthologue cluster would be considered homologues. Such a library of functional orthologue clusters can be found at hltp://www.nebi.nlm.nib.gov/COG. A gene can be classified into a cluster of orthologous groups or COG by using the COGNITOR program available at the above web site, or by direct BLASTP comparison of the gene of interest to the members of the COGs and analysis of these results as described by Tatusov, R.L., Galperin, M.Y., Natale, D. A. and Koonin, E.V. (2000) The COG database: a tool for genome- scale analysis of protein functions and evolution. Nucleic Acids Research v. 2 8 n. 1, pp3 3 -3 6.

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The term "homologous coding nucleic acid" also includes nucleic acids comprising nucleotide sequences which encode polypeptides having at least 99%, 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, at least 50%, at least 40% or at least 25% amino acid identity or similarity to a polypeptide comprising the amino acid sequence of one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 or to a polypeptide whose expression is inhibited by a nucleic acid comprising a

nucleotide sequence of one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 or fragments comprising at least 5, 10, 15, 20, 25, 30,35, 40, 50, 75, 100, or 150 consecutive amino acids thereof as determined using the FASTA version 3.0t78 algorithm with the default parameters. Alternatively, protein identity or similarity may be identified using BLASTP with the default parameters, BLASTX with the default parameters, TBLASTN with the default parameters, or tBLASTX with the default parameters. (Altschul, S.F. et al. Gapped BLAST and PSI-BLAST: A New Generation of Protein Database Search Programs, Nucleic Acid Res. 25: 3389-3402 (1997)).

The term "homologous coding nucleic acid" also includes coding nucleic acids which hybridize under stringent conditions to a nucleic acid selected from the group consisting of the nucleotide sequences complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and coding nucleic acids comprising nucleotide sequences which hybridize under stringent conditions to a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides of the sequences complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105.

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As used herein, "stringent conditions" means hybridization to filter-bound nucleic acid in 6xSSC at about 45'C followed by one or more washes in 0. lxSSC/0.2/SDS at about 680C. Other exemplary stringent conditions may refer, e.g., to washing in 6xSSC/0.05% sodium pyrophosphate at 37C, 48'C, 55'C, and 60'C as appropriate for the 5 particular probe being used.

The term "homologous coding nucleic acid" also includes coding nucleic acids comprising nucleotide sequences which hybridize under moderate conditions to a nucleotide sequence selected from the group consisting of the sequences complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and coding nucleic acids comprising nucleotide sequences which hybridize under moderate conditions to a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150,200,300,400, or 500 consecutive nucleotides of the sequences complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105. As used herein, "moderate conditions" means hybridization to filter-bound DNA in 6x sodium chloride/sodium citrate (SSC) at about 45'C followed by one or more washes in 0.2xSSC/0. 1 % SDS at about 42-65'C.

The term "homologous coding nucleic acids" also includes nucleic acids comprising nucleotide sequences which encode a gene product whose activity may

be complemented by a gene encoding a gene product whose activity is inhibited by a nucleic acid comprising a nucleotide sequence selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105. In some embodiments, the homologous coding nucleic acids may encode a gene product whose activity is complemented by the gene product encoded by a nucleic acid comprising a nucleotide sequence selected from the group consisting Seq ID Nos.: 1 to Seq. ID Nos.: 105. In other embodiments, the homologous coding nucleic acids may comprise a nucleotide sequence encodes a gene product whose activity is complemented by one of the polypeptides of Seq ID Nos.: 1 to Seq. ID Nos.: 105.

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The term "homologous antisense nucleic acid" includes nucleic acids comprising a nucleotide sequence having at least 97%, at least 95%, at least 90%, at least 85%, at least 80%, or at least 70% nucleotide sequence identity to a nucleotide sequence selected from the group consisting of one of the sequences of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and fragments comprising at least 10, 15, 20, 25, 30,35,40, 50, 75, 100, 150, 200,300,400, or 500 consecutive nucleotides thereof. Homologous antisense nucleic acids may also comprising nucleotide sequences which have at least 97%, at least 95%, at least 90%, at least 85%, at least 80%, or at least 70% nucleotide sequence identity to a nucleotide sequence selected from the group consisting of the sequences complementary to one of sequences of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and fragments comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides thereof.

Nucleic acid identity may be determined as described above.

The term "homologous antisense nucleic acid" also includes antisense nucleic acids comprising nucleotide sequences which hybridize under stringent conditions to a nucleotide sequence complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and antisense nucleic acids comprising nucleotide sequences which hybridize under stringent conditions to a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150,200, 300, 400, or 500 consecutive nucleotides of the sequence complementary to one Seq ID Nos.: 1 to Seq. ID Nos.: 105. Homologous antisense nucleic acids also include antisense nucleic acids comprising nucleotide sequences which hybridize under stringent conditions to a nucleotide sequence selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105, and antisense nucleic acids comprising nucleotide sequences which hybridize under stringent conditions to

a fragment comprising at least 10, 15, 20,25, 30, 35, 40, 50, 75, 100,150,200,300,400, or 500 consecutive nucleotides of one of Seq ID Nos.: 1 to Seq. ID Nos.: 105.

The term "homologous antisense nucleic acid" also includes antisense nucleic acids comprising nucleotide sequences which hybridize under moderate conditions to a nucleotide sequence complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and antisense nucleic acids comprising nucleotide sequences which hybridize under moderate conditions to a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides of the sequence complementary to one of Seq ID Nos.: 1 to Seq. ID Nos.: 105.

Homologous antisense nucleic acids also include antisense nucleic acids comprising nucleotide sequences which hybridize under moderate conditions to a nucleotide sequence selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105 and antisense nucleic acids which comprising nucleotide sequences hybridize under moderate conditions to a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides of one of Seq ID Nos.: 1 to Seq. ID Nos.: 1

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By "homologous polypeptide" is meant a polypeptide homologous to a polypeptide whose activity or level is inhibited by a nucleic acid comprising a nucleotide sequence selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105 by a homologous antisense nucleic acid. The term "homologous polypeptide" includes polypeptides having at least 99%, 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, at least 50%, at least 40% or at least 25% amino acid identity or similarity to a polypeptide whose activity or level is inhibited by a nucleic acid selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105 or by a homologous antisense nucleic acid, or polypeptides having at least 99%, 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, at least 50%, at least 40% or at least 25% amino acid identity or similarity to a polypeptide to a fragment comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids of a polypeptide whose activity or level is inhibited by a nucleic acid selected from the group consisting of Seq ID Nos.: 1 to Seq. ID Nos.: 105 or by a homologous antisense nucleic acid. Identity or similarity may be determined using the FASTA version 3. Ot78 algorithm with the default

parameters. Alternatively, protein identity or similarity may be identified using BLASTP with the default parameters, BLASTX with the default parameters, or TBLASTN with the default parameters. (Altschul, S.F. et al. Gapped BLAST and PSI-BLAST: A New Generation of Protein Database Search Programs, Nucleic Acid Res. 25: 3389-3402 (1997).

The term homologous polypeptide also includes polypeptides having at least 99%, 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, at least 50%, at least 40% or at least 25% amino acid identity or similarity to a polypeptide selected from the group consisting of Seq ID Nos.: 2 to Seq. ID Nos.: 106 and polypeptides having at least 99%, 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, at least 50%, at least 40% or at least 25% amino acid identity or similarity to a fragment comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids of a polypeptide selected from the group consisting of Seq ID Nos.: 2 to Seq. ID Nos.: 106.

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The invention also includes polynucleotides, preferably DNA molecules, that hybridize to one of the nucleic acids of Seq ID Nos.: 2 to Seq. ID Nos.: 106 or the complements of any of the preceding nucleic acids. Such hybridization may be under stringent or moderate conditions as defined above or under other conditions which permit specific hybridization. The nucleic acid molecules of the invention that hybridize to these DNA sequences include oligodeoxynucleotides ("oligos") which hybridize to the target gene under highly stringent or stringent conditions. In general, for oligos between 14 and 70 nucleotides in length the melting temperature (Tm) is calculated using the formula:

Tm ff) = 81.5 + 16.6(log[monovalent cations (molar)] + 0.41 (% G+Q - (500N)) where N is the length of the probe. If the hybridization is carried out in a solution containing formamide, the melting temperature may be calculated using the equation:

Tm('C) = 81.5 + 16.6(log[monovalent cations (niolar)] + 0.4 1 (% G+C) - (0.6 1) (% formamide) - (500N) where N is the length of the probe. In general, hybridization is carried out at about 20-25 degrees below Tin (for DNA-DNA hybrids) or about 10- 15 degrees below Tin (for RNA-DNA hybrids).

Other hybridization conditions are apparent to those of skill in the art (see, for example, Ausubel, F.M. et al., eds., 1989, Current Protocols in Molecular Biology,

Vol. 1, Green Publishing Associates, Inc. and John Wiley & Sons, Inc., New York, at pp. 6.3.1-6.3.6 and 2.10.3.

By "identifying a compound" is meant to screen one or more compounds in a collection of compounds such as a combinatorial chemical library or other library of chemical compounds or to characterize a single compound by testing the compound in a given assay and determining whether it exhibits the desired activity.

By "inducer" is meant an agent or solution which, when placed in contact with a cell or microorganism, increases transcription, or inhibitor and/or promoter clearance/fidelity, from a desired promoter.

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As used herein, "nucleic acid" means DNA, RNA, or modified nucleic acids. Thus, the terminology "the nucleic acid of SEQ ID NO: V or "the nucleic acid comprising the nucleotide sequence" includes both the DNA sequence of SEQ ID NO: X and an RNA sequence in which the thymidines in the DNA sequence have been substituted with uridines in the RNA sequence and in which the deoxyribose backbone 'of the DNA sequence has been substituted with a ribose backbone in the RNA sequence. Modified nucleic acids are nucleic acids having nucleotides or structures which do not occur in nature, such as nucleic acids in which the internucleotide phosphate residues with methylphosphonates, phosphorothioates, phosphoramidates, and phosphate esters. Nonphosphate internucleotide analogs such as siloxane bridges, carbonate bridges, thioester bridges, as well as many others known in the art may also be used in modified nucleic acids. Modified nucleic acids may also comprise, (x-anomeric nucleotide units and modified micleotides such as 1 2 dideoxy-d-ribofuranose, 1,2-dideoxy- I -phenylribofuranose, and N4, N4ethano-5 -methyl-cytosine are contemplated for use in the present invention. Modified nucleic acids may also be peptide nucleic acids in which the entire deoxyribose-phosphate backbone has been exchanged with a chemically completely different, but structurally homologous, polyamide (peptide) backbone containing 2aminoethyl glycogen units.

As used herein, "sub-lethal" means a concentration of an agent below the concentration required to inhibit all cell growth.

A proliferation-required gene or gene family is one where, in the absence or substantial reduction of a gene transcript and/or gene product, growth or viability of the cell or microorganism is reduced or eliminated. Thus, as used herein, the

terminology "proliferation- required" or "required for proliferation" encompasses instances where the absence or substantial reduction of a gene transcript and/or gene product completely eliminates cell growth as well as instances where the absence of a gene transcript and/or gene product merely reduces cell growth. These proliferation-required genes can be used as potential targets for the generation of new antimicrobial agents. To achieve that goal, the present invention also encompasses assays for analyzing proliferation- required genes and for identifying compounds which interact with the gene and/or gene products of the proliferation-required genes. In addition, the present invention contemplates the expression of genes and the purification of the proteins encoded by the nucleic acid sequences identified as required proliferation genes and reported herein. The purified proteins can be used to generate reagents and screen small molecule libraries or other candidate compound libraries for compounds that can be further developed to yield novel antimicrobial compounds.

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The invention described herein addresses the need for identifying Alloiococcus otitidis proliferation-required gene or gene family that may be used to identify compounds, which are effective in preventing or treating most or all of the disease caused by Alloiococcus otitidis. The invention further addresses the need for methods of diagnosing Alloiococcus otitidis infection using the genes and the polypeptides identified herein. The inventors have identified novel Alloiococcus otitidis open reading frames (Ors), which encode proteins/polypeptides that are essential for the growth and proliferation of the bacteria. More particularly, the newly identified Ors encode polypeptides that are essential for proliferation of Alloiococcus otitidis, and thus serve as potential targets for antimicrobial compounds. Thus, in certain embodiments, the invention comprises Alloiococcus otitidis Ors encoding polypeptides that are essential for cellular proliferation, transcription gene products of Alloiococcus otitidis Ors, including, but not limited to mRNA, antisense RNA, antisense oligonucleotides, and ribozyme molecules, which can be used to inhibit or control growth of the microorganism. The invention relates also to methods of detecting Alloiococcus otitidis nucleic acids or polypeptides and kits for diagnosing Alloiococcus otitidis infection. The invention also relates to pharmaceutical compositions, in particular antimicrobial compounds in pharmaceutical compositions, for the prevention and/or treatment of bacterial infection, in particular infection

caused by or exacerbated by Alloiococcus otitidis.

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# B. ALLOIOCOCCUS OTITIDIS ORF POLYNUCLEOTIDES ENCODING POLYPEPTIDES ESSENTIAL FOR PROLIFERATION

Isolated and purified Alloiococcus otitidis ORF polynucleotides of the present invention are contemplated for use in the production of Alloiococcus otitidis polypeptides. More specifically, in certain embodiments, the ORFs encode Alloiococcus otitidis polypeptides that are essential for cell proliferation. Thus, in one aspect, the present invention provides isolated and purified polynucleotides (ORFs) that encode Alloiococcus otitidis essential for cell proliferation. In particular embodiments, a polynucleotide of the present invention is a DNA molecule, wherein the DNA may be genomic DNA, plasmid DNA or cDNA. In a preferred embodiment, a polynucleotide of the present invention is a recombinant polynucleotide, which encodes an Alloiococcus otitidis polypeptide comprising an amino acid sequence that has at least 25% identity to an amino acid sequence of one of even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106 or a fragment thereof. In another embodiment, an isolated and purified ORF polynucleotide comprises a nucleotide sequence that has at least 70% identity to one of the ORF polynucleotide nucleotide sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105, a degenerate variant thereof, or a complement thereof. In yet another embodiment, an ORF polynucleotide of one of SEQ ID NO: 1 through SEQ ID NO: 105 is comprised in a plasmid vector and expressed in a host cell. In a preferred embodiment, the host cell is a prokaryotic host cell.

As used herein, the term "polynucleotide" means a sequence of nucleotides connected by phosphodiester linkages. Polynucleotides are presented herein in the direction from the 5' to the 3' direction. A polynucleotide of the present invention can comprise from about 10 to about several hundred thousand base pairs. Preferably, a polynucleotide comprises from about 10 to about 3,000 base pairs. Preferred lengths of particular polynucleotide are set forth hereinafter.

A polynucleotide of the present invention can be a deoxyribonucleic acid (DNA) molecule, a ribonucleic acid (RNA) molecule, or analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA. Where a

polynucleotide is a DNA molecule, that molecule can be a gene, a cDNA molecule or a genomic DNA molecule. Nucleotide bases are indicated herein by a single letter code: adenine (A), guanine (G), thymine (T) and cytosine (C).

"Isolated" means altered "by the hand of man" from the natural state. An "isolated" composition or substance is one that has been changed or removed from its original environment, or both. For example, a polynucleotide or a polypeptide naturally present in a living animal is not "isolated," but the same polynucleotide or polypeptide separated from the coexisting materials of its natural state is "isolated," as the term is employed herein.

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Preferably, an "isolated" polynucleotide is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated *Alloiococcus otitidis* nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0. 5 kb or 0. 1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. However, the *Alloiococcus otitidis* nucleic acid molecule can also be fused to heterologous protein encoding or regulatory sequences and still be considered isolated.

ORF polynucleotides of the present invention may also be obtained using standard cloning and screening techniques from a cDNA library derived from mRNA. Polynucleotides of the invention can also be obtained from natural sources such as genomic DNA libraries (*e.g.*, an *Alloiococcus otitidis* library) or can be synthesized using well-known and commercially available techniques. As contemplated in the present invention, ORF polynucleotides are obtained using *Alloiococcus otitidis* chromosomal DNA as the template.

The invention further encompasses nucleic acid molecules that differ from the nucleotide sequences set forth in the odd numbered sequences listed in ID NO: 1 through SEQ ID NO: 105 (and fragments thereof) due to degeneracy of the genetic code, and thus encode the same *Alloiococcus otitidis* polypeptides as those encoded by the amino acid sequences shown in even numbered sequences set forth in SEQ ID NO:2 through SEQ ID NO: 106

Orthologs and allelic variants of the *Alloiococcus otitidis* polynucleotides are readily identified using methods well known in the art. An allelic variant or an

orthologue of the polynucleotides comprises a nucleotide sequence that is typically at least about 70-75%, more typically at least about 80-85%, and most typically at least about 90-95% or more homologous to the nucleotide sequence shown in one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO: 105, or a fragment of these nucleotide sequences. Such nucleic acid molecules are readily identified as being able to hybridize, preferably under stringent conditions, to the nucleotide sequence shown in one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO: 105, or a fragment of these nucleotide sequences.

Moreover, the polynucleotides of the invention can comprise only a fragment of the coding region of an *Alloiococcus otitidis* polynucleotide or gene, such as a fragment of one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO: 105.

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When the ORF polynucleotides of the invention are used for the recombinant production of *Alloiococcus otitidis* polypeptides of the present invention, the polynucleotide may include the coding sequence for the mature polypeptide, by itself, or the coding sequence for the mature polypeptide in reading frame with other coding sequences, such as those encoding a leader or secretory sequence, a pre-, or pro- or prepro- protein sequence, or other fusion peptide portions. For example, a marker sequence which facilitates purification of the fused polypeptide can be linked to the coding sequence (*see* Gentz *et al.*, 1989, incorporated herein by reference). Thus, contemplated in the present invention is the preparation of polynucleotides encoding fusion polypeptides permitting His-tag purification of expression products. The polynucleotide may also contain non-coding 5' and 3' sequences, such as transcribed, non-translated sequences, splicing and polyadenylation signals.

Thus, a polynucleotide encoding a polypeptide of the present invention, including homologs and orthologs from species other than *Alloiococcus otitidis*, may be obtained by a process which comprises the steps of screening an appropriate library under stringent hybridization conditions with a labeled probe having the sequence of one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO: 105 or a fragment thereof; and isolating full-length cDNA and genomic clones containing the polynucleotide sequence. Such hybridization techniques are well known to the skilled artisan. The skilled artisan will appreciate that, in many cases, an isolated cDNA sequence will be incomplete, in that the region coding for

the polypeptide is cut short at the 5' end of the cDNA. This is a consequence of reverse transcriptase, an enzyme with inherently low "processivity" (a measure of the ability of the enzyme to remain attached to the template during the polymerization reaction), failing to complete a DNA copy of the mRNA template during the first-strand cDNA synthesis.

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Thus, in certain embodiments, the polynucleotide sequence information provided by the present invention allows for the preparation of relatively short DNA (or RNA) oligonucleotide sequences having the ability to specifically hybridize to gene sequences of the selected polynucleotides disclosed herein. The term "oligonucleotide" as used herein is defined as a molecule comprised of two or more deoxyribonucleotides or ribonucleotides, usually more than three (3), and typically more than ten (10) and up to one hundred (100) or more (although preferably between twenty and thirty). The exact size will depend on many factors, which in turn depends on the ultimate function or use of the oligonucleotide. Thus, in particular embodiments of the invention, nucleic acid probes of an appropriate length are prepared based on a consideration of a selected nucleotide sequence, e.g., a sequence such as that shown in one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO: 105. The ability of such nucleic acid probes to specifically hybridize to a polynucleotide encoding an Alloiococcus otitidis polypeptide lends them particular utility in a variety of embodiments. Most importantly, the probes can be used in a variety of assays for detecting the presence of complementary sequences in a given sample.

In certain embodiments, it is advantageous to use oligonucleotide primers. These primers are generated in any manner, including chemical synthesis, DNA replication, reverse transcription, or a combination thereof. The sequence of such primers is designed using a polynucleotide of the present invention for use in detecting, amplifying or mutating a defined segment of an ORF polynucleotide that encodes an *Alloiococcus otitidis* polypeptide from prokaryotic cells using polymerase chain reaction (PCR) technology.

In certain embodiments, it is advantageous to employ a polynucleotide of the present invention in combination with an appropriate label for detecting hybrid formation. A wide variety of appropriate labels are known in the art, including

radioactive, enzymatic or other ligands, such as avidin/biotin, which are capable of giving a detectable signal.

Polynucleotides which are identical or sufficiently identical to a nucleotide sequence contained in one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO: 105, or a fragment thereof, may be used as hybridization probes for cDNA and genomic DNA or as primers for a nucleic acid amplification (PCR) reaction, to isolate full-length cDNAs and genomic clones encoding polypeptides of the present invention and to isolate cDNA and genomic clones of other genes (including genes encoding homologs and orthologs from species other than *Alloiococcus otitidis*) that have a high sequence similarity to polynucleotide sequences set forth in one of the odd numbered sequences set forth in SEQ ID NO:1 through SEQ ID NO:105, or a fragment thereof. Typically these nucleotide sequences are from at least 70% identical to at least about 95% identical to that of the reference polynucleotide sequence. The probes or primers will generally comprise at least 15 nucleotides, preferably, at least 30 nucleotides and may have at least 50 nucleotides. Particularly preferred probes will have between 30 and 50 nucleotides.

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There are several methods available and well known to those skilled in the art to obtain full-length cDNAs, or extend short cDNAs, for example those based on the method of Rapid Amplification of cDNA ends (RACE) (see, Frohman et al., 1988). Recent modifications of the technique, exemplified by the Marathon™ technology [Promega, Madison, WI], for example, have significantly simplified the search for longer cDNAs. In the Marathon™ technology, cDNAs have been prepared from mRNA extracted from a chosen tissue and an "adaptor" sequence ligated onto each end. Nucleic acid amplification (PCR) is then carried out to amplify the "missing" 5′ end of the cDNA using a combination of gene specific and adaptor specific oligonucleotide primers. The PCR reaction is then repeated using "nested" primers, that is, primers designed to anneal within the amplified product (typically an adaptor specific primer that anneals further 3' in the adaptor sequence and a gene specific primer that anneals further 5' in the known gene sequence). The products of this reaction are then analyzed by DNA sequencing and a full-length cDNA constructed either by joining the product directly to the existing cDNA to give a complete

sequence, or carrying out a separate full-length PCR using the new sequence information for the design of the 5' primer.

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To provide certain of the advantages in accordance with the present invention, a preferred nucleic acid sequence employed for hybridization studies or assays includes probe molecules that are complementary to at least a 10 to about 70 nucleotides long stretch of a polynucleotide that encodes an Alloiococcus otitidis polypeptide, such as that shown in one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106. A size of at least 10 nucleotides in length helps to ensure that the fragment will be of sufficient length to form a duplex molecule that is both stable and selective. Molecules having complementary sequences over stretches greater than 10 bases in length are generally preferred in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. It is generally preferable to design nucleic acid molecules with gene-complementary stretches of 25 to 40 nucleotides, 55 to 70 nucleotides, or even longer where desired. For example, such fragments are readily prepared by directly synthesizing the fragment by chemical means, by application of nucleic acid reproduction technology, such as the PCR technology (U.S. Patent 4,683,202, incorporated herein by reference), or by excising selected DNA fragments from recombinant plasmids containing appropriate inserts and suitable restriction enzyme sites.

In another aspect, the present invention contemplates an isolated and purified polynucleotide comprising a nucleotide sequence that is identical or complementary to a segment of at least 10 contiguous bases of one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105, wherein the polynucleotide hybridizes to a polynucleotide that encodes an *Alloiococcus otitidis* polypeptide. Preferably, the isolated and purified polynucleotide comprises a base sequence that is identical or complementary to a segment of at least 25 to 70 contiguous bases of one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105. For example, the polynucleotide of the invention can comprise a segment of bases identical or complementary to from 40 to 55 contiguous bases of the disclosed nucleotide sequences.

Accordingly, a polynucleotide probe molecule of the invention can be used for its ability to selectively form duplex molecules with complementary stretches of the

gene. Depending on the application envisioned, varying conditions of hybridization are employed to achieve varying degrees of selectivity of the probe toward the target sequence. For applications requiring a high degree of selectivity, relatively stringent conditions are employed to form the hybrids. Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate an *Alloiococcus otitidis* homologous polypeptide coding sequence from other cells, functional equivalents, or the like, less stringent hybridization conditions are typically needed to allow formation of the heteroduplex (see Table 2). Cross-hybridizing species are thereby readily identified as positively hybridizing signals with respect to control hybridizations. Thus, hybridization conditions are readily manipulated, and thus will generally be a method of choice depending on the desired results.

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Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate a homologous polypeptide coding sequence from other cells, functional equivalents, or the like, less stringent hybridization conditions are typically needed to allow formation of the heteroduplex. Cross-hybridizing species are thereby readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions are readily manipulated, and thus are generally a method of choice depending on the desired results.

The present invention also includes polynucleotides capable of hybridizing under reduced stringency conditions, more preferably stringent conditions, and most preferably highly stringent conditions, to polynucleotides described herein. Examples of stringency conditions are shown in the table below: highly stringent conditions are those that are at least as stringent as, for example, conditions A-F; stringent conditions are at least as stringent as, for example, conditions G-L; and reduced stringency conditions are at least as stringent as, for example, conditions M-R.

TABLE 2
STRINGENCY CONDITIONS

Stringency Condition	Polynucleotide Hybrid	Hybrid Length (bp) <sup>l</sup>	Hybridization Temperature and Buffer <sup>H</sup>	Wash Temperature and BufferH
A	DNA:DNA	> 50	65°C; 1xSSC -or- 42 °C; 1xSSC, 50% formamide	65 °C; 0.3xSSC
В	DNA:DNA	< 50	T <sub>B</sub> : 1xSSC	T <sub>B</sub> ; 1xSSC
C	DNA:RNA	> 50	67 °C; 1xSSC -or- 45 °C; 1xSSC, 50% formamide	67 °C; 0.3xSSC
D	DNA:RNA	< 50	T <sub>D</sub> ; 1xSSC	T <sub>D</sub> ; 1xSSC
E	RNA:RNA	> 50	70°C; 1xSSC -or- 50°C; 1xSSC, 50% formamide	70°C; 0.3xSSC
F	RNA:RNA	< 50	T <sub>F</sub> ; 1xSSC	T <sub>F</sub> ; 1xSSC
G	DNA:DNA	> 50	65°C; 4xSSC -or- 42°C; 4xSSC, 50% formamide	65°C; 1xSSC
H.	DNA:DNA	< 50	T <sub>H</sub> ; 4xSSC	T <sub>H</sub> ; 4xSSC
Ĭ	DNA:RNA	> 50	67 °C; 4xSSC -or- 45 °C; 4xSSC, 50% formamide	67°C; 1xSSC
J	DNA:RNA	< 50	T <sub>J</sub> ; 4xSSC	T <sub>J</sub> ; 4xSSC
К	RNA:RNA	> 50	70°C; 4xSSC -or-	67°C; 1xSSC
			50EC; 4xSSC, 50% formamide	
L	RNA:RNA	< 50	T <sub>L</sub> ; 2xSSC	T <sub>L</sub> ; 2xSSC
	DNA:DNA	> 50	50 °C; 4xSSC -or-	50 °C; 2xSSC
			40 °C; 6xSSC, 50% formamide	
N	DNA:DNA	< 50	T <sub>N</sub> ; 6xSSC	T <sub>N</sub> ; 6xSSC
<del></del>	DNA:RNA	> 50	55 °C; 4xSSC -or-	55 °C; 2xSSC
_			42 °C; 6xSSC, 50%	
			formamide	
Р	DNA:RNA	< 50	T <sub>P</sub> ; 6xSSC	T <sub>P</sub> ; 6xSSC
Q	RNA:RNA	> 50	60°C; 4xSSC -or-	60°C; 2xSSC
			45 °C; 6xSSC, 50% formamide	
		- 50		Te: 4xSSC
R	RNA:RNA	< 50	formamide T <sub>R</sub> ; 4xSSC	T <sub>R</sub> ; 4xSSC

(bp)<sup>1</sup>: The hybrid length is that anticipated for the hybridized region(s) of the hybridizing polynucleotides. When hybridizing a polynucleotide to a target

polynucleotide of unknown sequence, the hybrid length is assumed to be that of the hybridizing polynucleotide. When polynucleotides of known sequence are hybridized, the hybrid length can be determined by aligning the sequences of the polynucleotides and identifying the region or regions of optimal sequence complementarity.

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Buffer<sup>H</sup>: SSPE (1xSSPE is 0.15M NaCl, 10mM NaH<sub>2</sub>PO<sub>4</sub>, and 1.25mM EDTA, pH 7.4), can be substituted for SSC (1xSSC is 0.15M NaCl and 15mM sodium citrate) in the hybridization and wash buffers; washes are performed for 15 minutes after hybridization is complete.

 $T_B$  through  $T_R$ : The hybridization temperature for hybrids anticipated to be less than 50 base pairs in length should be 5-10EC less than the melting temperature  $(T_m)$  of the hybrid, where  $T_m$  is determined according to the following equations. For hybrids less than 18 base pairs in length,  $T_m(EC) = 2(\# \text{ of A} + T \text{ bases}) + 4(\# \text{ of G} + C \text{ bases})$ . For hybrids between 18 and 49 base pairs in length,  $T_m(EC) = 81.5 + 16.6(log_{10}[Na^+]) + 0.41(\%G+C) - (600/N)$ , where N is the number of bases in the hybrid, and  $[Na^+]$  is the concentration of sodium ions in the hybridization buffer ( $[Na^+]$  for 1xSSC = 0.165 M).

Additional examples of stringency conditions for polynucleotide hybridization are provided in Sambrook *et al.*, 1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, chapters 9 and 11, and Ausubel *et al.*, 1995, Current Protocols in Molecular Biology, Eds., John Wiley & Sons, Inc., sections 2.10 and 6.3-6.4, incorporated herein by reference.

In addition to the nucleic acid molecules encoding *Alloiococcus otitidis* polypeptides described above, another aspect of the invention pertains to isolated nucleic acid molecules that are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence that is complementary to a "sense" nucleic acid encoding a protein, *e.g.*, complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire *Alloiococcus otitidis* coding strand, or to only a fragment thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding an *Alloiococcus otitidis* polypeptide.

The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues, *e.g.*, the entire coding region of each of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105. In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding an *Alloiococcus otitidis* polypeptide. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (*i.e.*, also referred to as 5' and 3' untranslated regions).

Given the coding strand sequence encoding the *Alloiococcus otitidis* polypeptides disclosed herein antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of *Alloiococcus otitidis* mRNA, but more preferably is an oligonucleotide which is antisense to only a fragment of the coding or noncoding region of *Alloiococcus otitidis* mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of *Alloiococcus otitidis* mRNA.

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An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxylmethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, I-methylguanine, I-methylinosine, 2,2-dimethylguanine, 2methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-

methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine.

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Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated in situ such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding an Alloiococcus otitidis polypeptide to thereby inhibit expression of the polypeptide, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of an antisense nucleic acid molecule of the invention includes direct injection at a tissue site. Alternatively, an antisense nucleic acid molecule can be modified to target selected cells and then administered systemically. For example, for systemic administration, an antisense molecule can be modified such that it specifically binds to a receptor or an antigen expressed on a selected cell surface, e.g., by linking the antisense nucleic acid molecule to a peptide or an antibody which binds to a cell surface receptor or antigen. The antisense nucleic acid molecule can also be delivered to cells using the vectors described herein.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an α-anomeric nucleic acid molecule. An α-anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual γ-units, the strands run parallel to each other (Gaultier *et al.*, 1987). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue *et al.*, 1987) or a chimeric RNA-DNA analogue (Inoue *et al.*, 1987).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity that are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes described in Haselhoff and Gerlach, 1988) can be used to catalytically cleave Alloiococcus otitidis mRNA transcripts to thereby inhibit translation of Alloiococcus otitidis mRNA. A ribozyme having specificity for an Alloiococcus otitidis-encoding nucleic acid can be designed based upon the nucleotide sequence of an Alloiococcus otitidis cDNA disclosed herein. For example, a derivative of a Tetrahymena L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an Alloiococcus otitidis-encoding mRNA. See, e.g., Cech et al. U.S. 4,987,071 and Cech et al. U.S. 5,116,742 both incorporated herein in their entirety by reference. Alternatively, Alloiococcus otitidis mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, e.g., Bartel and Szostak, 1993.

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Alternatively Alloiococcus otitidis gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the Alloiococcus otitidis gene (e.g., the Alloiococcus otitidis gene promoter and/or enhancers) to form triple helical structures that prevent transcription of the Alloiococcus otitidis gene in target cells. See generally, Helene, 1991; Helene et al., 1992; and Maher, 1992.

Alloiococcus otitidis gene expression can also be inhibited using RNA interference (RNAi). This is a technique for post-transcriptional gene silencing (PTGS), in which target gene activity is specifically abolished with cognate double-stranded RNA (dsRNA). RNAi resembles in many aspects PTGS in plants and has been detected in many invertebrates including trypanosome, hydra, planaria, nematode and fruit fly (*Drosophila melangnoster*). It may be involved in the modulation of transposable element mobilization and antiviral state formation. RNAi in mammalian systems is disclosed in WO 00/63364, which is incorporated by reference herein in its entirety. Basically, dsRNA of at least about 600 nucleotides, homologous to the target is introduced into the cell and a sequence specific reduction in gene activity is observed.

### C. ALLOIOCOCCUS OTITIDIS POLYPEPTIDES

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In particular embodiments, the present invention provides isolated and purified *Alloiococcus otitidis* polypeptides. Preferably, an *Alloiococcus otitidis* polypeptide of the invention is a recombinant polypeptide. In certain embodiments, an *Alloiococcus otitidis* polypeptide of the present invention comprises the amino acid sequence that has at least 25% identity to the amino acid sequence of one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106, a biological equivalent thereof, or a fragment thereof.

An *Alloiococcus otitidis* polypeptide according to the present invention encompasses a polypeptide that comprises: 1) the amino acid sequence shown in one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106) functional and non-functional naturally occurring variants or biological equivalents of *Alloiococcus otitidis* polypeptides of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106 and recombinantly produced variants or biological equivalents of *Alloiococcus otitidis* polypeptides set out in SEQ ID NO: 2 through SEQ ID NO: 106) polypeptides isolated from organisms other than *Alloiococcus otitidis* (orthologs of *Alloiococcus otitidis* polypeptides.)

A biological equivalent or variant of an *Alloiococcus otitidis* polypeptide according to the present invention encompasses 1) a polypeptide isolated from *Alloiococcus otitidis*; and 2) a polypeptide that contains substantial homology to an *Alloiococcus otitidis* polypeptide.

Biological equivalents or variants of *Alloiococcus otitidis* include both functional and non-functional *Alloiococcus otitidis* polypeptides. Functional biological equivalents or variants are naturally occurring amino acid sequence variants of an *Alloiococcus otitidis* polypeptide that maintain the ability to elicit an immunological or antigenic response in a subject. Functional variants will typically contain only conservative substitutions of one or more amino acids in any one of even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106 or substitution, deletion or insertion of non-critical residues in non-critical regions of the polypeptide.

The present invention further provides non-Alloiococcus otitidis orthologues of Alloiococcus otitidis polypeptides. Orthologues of Alloiococcus otitidis polypeptides

are polypeptides that are isolated from non-Alloiococcus otitidis organisms and possess antigenic capabilities of the Alloiococcus otitidis polypeptide. Orthologues of an Alloiococcus otitidis polypeptide can readily be identified as comprising an amino acid sequence that is substantially homologous to one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106.

Modifications and changes can be made in the structure of a polypeptide of the present invention and still obtain a molecule having *Alloiococcus otitidis* antigenicity. For example, certain amino acids can be substituted for other amino acids in a sequence without appreciable loss of antigenicity. Because it is the interactive capacity and nature of a polypeptide that defines that polypeptide's biological functional activity, certain amino acid sequence substitutions can be made in a polypeptide sequence (or, of course, its underlying DNA coding sequence) and nevertheless obtain a polypeptide with like properties.

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In making such changes, the hydropathic index of amino acids can be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a polypeptide is generally understood in the art (Kyte & Doolittle, 1982). It is known that certain amino acids can be substituted for other amino acids having a similar hydropathic index or score and still result in a polypeptide with similar biological activity. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics. Those indices are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5); glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

It is believed that the relative hydropathic character of the amino acid residue determines the secondary and tertiary structure of the resultant polypeptide, which in turn defines the interaction of the polypeptide with other molecules, such as enzymes, substrates, receptors, antibodies, antigens, and the like. It is known in the art that an amino acid can be substituted by another amino acid having a similar hydropathic index and still obtain a functionally equivalent polypeptide. In such changes, the substitution of amino acids whose hydropathic indices are within +/-2 is

preferred, those within +/-1 are particularly preferred, and those within +/-0.5 are even more particularly preferred.

Substitution of like amino acids can also be made on the basis of hydrophilicity, particularly where the biologically functional equivalent polypeptide or peptide thereby created is intended for use in immunological embodiments. U.S. Pat. No. 4,554,101, incorporated herein by reference, states that the greatest local average hydrophilicity of a polypeptide, as governed by the hydrophilicity of its adjacent amino acids, correlates with its immunogenicity and antigenicity, *i.e.* with a biological property of the polypeptide.

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As detailed in U.S. Pat. No. 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine ( $\pm$ 3.0); lysine ( $\pm$ 3.0); aspartate ( $\pm$ 3.0  $\pm$ 1); glutamate ( $\pm$ 3.0  $\pm$ 1); serine ( $\pm$ 0.3); asparagine ( $\pm$ 0.2); glutamine ( $\pm$ 0.2); glycine (0); proline ( $\pm$ 0.5  $\pm$ 1); threonine ( $\pm$ 0.4); alanine ( $\pm$ 0.5); histidine ( $\pm$ 0.5); cysteine ( $\pm$ 1.0); methionine ( $\pm$ 1.3); valine ( $\pm$ 1.5); leucine ( $\pm$ 1.8); isoleucine ( $\pm$ 1.8); tyrosine ( $\pm$ 2.3); phenylalanine ( $\pm$ 2.5); tryptophan ( $\pm$ 3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent polypeptide. In such changes, the substitution of amino acids whose hydrophilicity values are within  $\pm$ 2 is preferred, those which are within  $\pm$ 1 are particularly preferred, and those within  $\pm$ 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions which take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine (See Table 3, below). The present invention thus contemplates functional or biological equivalents of an *Alloiococcus otitidis* polypeptide as set forth above.

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TABLE 3:
AMINO ACID SUBSTITUTIONS

Original Residue	Exemplary Residue Substitution
Ala	Gly; Ser
Arg	Lys
Asn	Gln; His
Asp	Glu
Cys	Ser
Gln	Asn
Glu	Asp
Gly	Ala
His	Asn; Gln
lle	Leu; Val
Leu	lle; Val
Lys	Arg
Met	Met; Leu; Tyr
Ser	Thr
Thr	Ser
Ттр	Tyr
Tyr	Trp; Phe
Val	lle; Leu

Biological or functional equivalents of a polypeptide are also prepared using site-specific mutagenesis. Site-specific mutagenesis is a technique useful in the preparation of second generation polypeptides, or biologically functional equivalent polypeptides or peptides, derived from the sequences thereof, through specific mutagenesis of the underlying DNA. As noted above, such changes can be desirable where amino acid substitutions are desirable. The technique further provides a capacity to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the DNA. Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Typically, a primer of about 17 to 25 nucleotides in length is preferred, with about 5 to 10 residues on both sides of the site of the alteration of the sequence.

In general, the technique of site-specific mutagenesis is well known in the art.

As will be appreciated, the technique typically employs a phage vector, that can exist

in both a single stranded and double stranded form. Typically, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector which includes within its sequence a DNA sequence which encodes all or a portion of the *Alloiococcus otitidis* polypeptide sequence selected. An oligonucleotide primer bearing the desired mutated sequence is prepared (*e.g.*, synthetically). This primer is then annealed to the singled-stranded vector, and extended by the use of enzymes such as *Escherichia coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells such as *Escherichia coli* cells and clones are selected which include recombinant vectors bearing the mutation. Commercially available kits come with all the reagents necessary, except the oligonucleotide primers.

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An *Alloiococcus otitidis* polypeptide or polypeptide antigen of the present invention is understood to be any *Alloiococcus otitidis* polypeptide comprising substantial sequence similarity, structural similarity and/or functional similarity to an *Alloiococcus otitidis* polypeptide comprising the amino acid sequence of one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106. In addition, an *Alloiococcus otitidis* polypeptide or polypeptide antigen of the invention is not limited to a particular source. Thus, the invention provides for the general detection and isolation of the polypeptides from a variety of sources.

It is contemplated in the present invention, that an *Alloiococcus otitidis* polypeptide may advantageously be cleaved into fragments for use in further structural or functional analysis, or in the generation of reagents such as *Alloiococcus otitidis*-related polypeptides and *Alloiococcus otitidis*-specific antibodies. This can be accomplished by treating purified or unpurified *Alloiococcus otitidis* polypeptides with a peptidase such as endoproteinase glu-C (Boehringer, Indianapolis, IN). Treatment with CNBr is another method by which peptide fragments may be produced from natural *Alloiococcus otitidis* polypeptides. Recombinant techniques also can be used to produce specific fragments of an *Alloiococcus otitidis* polypeptide.

In addition, the inventors also contemplate that compounds sterically similar to a particular *Alloiococcus otitidis* polypeptide antigen, called peptidomimetics, may

be formulated to mimic the key portions of the peptide structure. Peptidemimetics are peptide-containing molecules that mimic elements of protein secondary structure. (See, for example, Johnson et al., 1993.) The underlying rationale behind the use of peptide mimetics is that the peptide backbone of proteins exists chiefly to orient amino acid side chains in such a way as to facilitate molecular interactions, such as those of receptor and ligand.

Successful applications of the peptide mimetic concept have thus far focused on mimetics of  $\beta$ -turns within proteins. Likely  $\beta$ -turn structures, within *Alloiococcus otitidis*, can be predicted by computer-based algorithms as discussed above. Once the component amino acids of the turn are determined, mimetics can be constructed to achieve a similar spatial orientation of the essential elements of the amino acid side chains, as discussed in Johnson *et al.*, 1993.

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Fragments of the *Alloiococcus otitidis* polypeptides are also included in the invention. A fragment is a polypeptide having an amino acid sequence that entirely is the same as a part, but not all, of the amino acid sequence. The fragment can comprise, for example, at least 7 or more (e.g., 8, 10 12, 14, 16, 18, 20 or more) contiguous amino acids of an one of amino acid sequence selected from one of the even numbered sequences set forth in SEQ ID NO.: 2 through SEQ ID NO.: 106. Fragments may be "freestanding" or comprised within a larger polypeptide of which they form a part or region, most preferably as a single, continuous region. In one embodiment, the fragments include at least one epitope of the mature polypeptide sequence.

"Fusion protein" refers to a protein encoded by two, often unrelated, fused genes or fragments thereof. For example, fusion proteins comprising various portions of constant region of immunoglobulin molecules together with another human protein or part thereof have been described. In many cases, employing an immunoglobulin Fc region as a part of a fusion protein is advantageous for use in therapy and diagnosis resulting in, for example, improved pharmacokinetic properties (see, e.g., EP-A 0232 2621). On the other hand, for some uses it would be desirable to be able to delete the Fc part after the fusion protein has been expressed, detected and purified.

## D. ALLOIOCOCCUS OTITIDIS POLYNUCLEOTIDE AND POLYPEPTIDE VARIANTS

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"Variant" as the term is used herein, is a polynucleotide or polypeptide that differs from a reference polynucleotide or polypeptide respectively, but retains essential properties. A typical variant of a polynucleotide differs in nucleotide sequence from another, reference polynucleotide. Changes in the nucleotide sequence of the variant may or may not alter the amino acid sequence of a polypeptide encoded by the reference polynucleotide. Nucleotide changes may result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptide encoded by the reference sequence, as discussed below. A typical variant of a polypeptide differs in amino acid sequence from another, reference polypeptide. Generally, differences are limited so that the sequences of the reference polypeptide and the variant are closely similar overall and, in many regions, identical. A variant and reference polypeptide may differ in amino acid sequence by one or more substitutions, additions and deletions in any combination. A substituted or inserted amino acid residue may or may not be one encoded by the genetic code. A variant of a polynucleotide or polypeptide may be a naturally occurring variant such as an allelic variant, or it may be a variant that is not known to occur naturally. Non-naturally occurring variants of polynucleotides and polypeptides may be made by mutagenesis techniques or by direct synthesis.

"Identity," as known in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as the case may be, as determined by the match between strings of such sequences. "Identity" can be readily calculated by known methods, including but not limited to those described in (Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Computer Analysis of Sequence Data, Part I, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; and Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991; and Carillo, H., and Lipman, D., SIAM J. Applied Math., 48: 1073 (1988). Preferred methods to determine identity are designed to give the largest match

between the sequences tested. Methods to determine identity are codified in publicly available computer programs. Preferred computer program methods to determine identity between two sequences include, but are not limited to; the GCG program package (Devereux, J., et al 1984), BLASTP, BLASTN, and FASTA (Altschul, S. F., et al., 1990. The BLASTX program is publicly available from NCBI and other sources (BLAST Manual, Altschul, S., et al., NCBI NLM NIH Bethesda, Md. 20894; Altschul, S., et al., 1990). The well known Smith-Waterman algorithm may also be used to determine identity.

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By way of example, a polynucleotide sequence of the present invention may be identical to the reference sequence of one of SEQ ID NO:1 through SEQ ID NO: 105, that is be 100% identical, or it may include up to a certain integer number of nucleotide alterations as compared to the reference sequence. Such alterations are selected from the group consisting of at least one nucleotide deletion, substitution, including transition and transversion, or insertion, and wherein said alterations may occur at the 5' or 3' terminal positions of the reference nucleotide sequence or anywhere between those terminal positions, interspersed either individually among the nucleotides in the reference sequence or in one or more contiguous groups within the reference sequence. The number of nucleotide alterations is determined by multiplying the total number of nucleotides in one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105 by the numerical percent of the respective percent identity (divided by 100) and subtracting that product from said total number of nucleotides in one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105.

For example, the alterations in an isolated *Alloiococcus otitidis* polynucleotide comprise a polynucleotide sequence that has at least 70% identity to the nucleic acid sequence of one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105; a degenerate variant thereof or a fragment thereof, wherein the polynucleotide sequence may include up to  $n_n$  nucleic acid alterations over the entire polynucleotide region of the nucleic acid sequence of any on of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105, wherein  $n_n$  is the maximum number of alterations and is calculated by the formula:

$$n_n \leq x_n - (x_n \cdot y),$$

in which  $x_n$  is the total number of nucleic acids of one of SEQ ID NO:1 through SEQ ID NO:105 and y has a value of 0.70, wherein any non-integer product of  $x_n$  and y is rounded down to the nearest integer prior to subtracting such product from  $x_n$ . Of course, y may also have a value of 0.80 for 80%, 0.85 for 85%, 0.90 for 90% 0.95 for 95%, etc.

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Similarly, a polypeptide sequence of the present invention may be identical to the reference sequence of any one of even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106, that is 100% identical, or it may include up to a certain integer number of amino acid alterations as compared to the reference sequence such that the percentage identity is less than 100%. Such alterations are selected from the group consisting of at least one amino acid deletion, substitution, including conservative and non-conservative substitution, or insertion, and wherein said alterations may occur at the amino- or carboxy-terminal positions of the reference polypeptide sequence or anywhere between those terminal positions, interspersed either individually among the amino acids in the reference sequence or in one or more contiguous groups within the reference sequence. The number of amino acid alterations for a given % identity is determined by multiplying the total number of amino acids in one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106 by the numerical percent of the respective percent identity (divided by 100) and then subtracting that product from said total number of amino acids in one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106, or:

 $n_a \leq x_a - (x_a \cdot y),$ 

wherein  $n_a$  is the number of amino acid alterations,  $x_a$  is the total number of amino acids in one of SEQ ID NO: 2 through SEQ ID NO: 106, and y is, for instance 0.70 for 70%, 0.80 for 80%, 0.85 for 85% etc., and wherein any non-integer product of x.sub.a and y is rounded down to the nearest integer prior to subtracting it from  $x_a$ .

## E. VECTORS, HOST CELLS AND RECOMBINANT ALLOIOCOCCUS OTITIDIS POLYPEPTIDES

In a preferred embodiment, the present invention provides expression vectors comprising ORF polynucleotides that encode *Alloiococcus otitidis* polypeptides. Preferably, the expression vectors of the present invention comprise ORF

polynucleotides that encode *Alloiococcus otitidis* polypeptides comprising the amino acid residue sequence of one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106. More preferably, the expression vectors of the present invention comprise a polynucleotide comprising the nucleotide base sequence of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105. Even more preferably, the expression vectors of the invention comprise a polynucleotide operatively linked to promoter. Still more preferably, the expression vectors of the invention comprise a polynucleotide operatively linked to a prokaryotic promoter. Alternatively, the expression vectors of the present invention comprise a polynucleotide operatively linked to an enhancer-promoter, that is, an eukaryotic promoter. The expression vectors further comprise a polyadenylation signal that is positioned 3' of the carboxy-terminal amino acid and within a transcriptional unit of the encoded polypeptide.

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Expression of proteins in prokaryotes is most often carried out in *Escherichia coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith and Johnson, 1988), pMAL (New England Biolabs, Beverly; MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S- transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

In one embodiment, the coding sequence of the *Alloiococcus otitidis* polynucleotide is cloned into a pGEX expression vector to create a vector encoding a fusion protein comprising, from the N-terminus to the C-terminus, GST-thrombin cleavage site-*Alloiococcus otitidis* polypeptide. The fusion protein can be purified by

affinity chromatography using glutathione-agarose resin. Recombinant *Alloiococcus* otitidis polypeptide unfused to GST can be recovered by cleavage of the fusion protein with thrombin.

Examples of suitable inducible non-fusion *Escherichia coli* expression vectors include pTrc (Amann *et al.*, 1988) and pET I I d (Studier *et al.*, 1990). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET I I d vector relies on transcription from a T7 gn1 0-lac fusion promoter mediated by a coexpressed viral RNA polymerase T7 gnl. This viral polymerase is supplied by host strains BL21 (DE3) or HMS I 74(DE3) from a resident prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter.

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One strategy to maximize recombinant protein expression in *Escherichia coli* is to express the protein in a host bacterium with an impaired capacity to proteolytically cleave the recombinant protein. Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *Escherichia coli*. Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA mutagenesis or synthesis techniques.

In another embodiment, the *Alloiococcus otitidis* polynucleotide expression vector is a yeast expression vector. Examples of vectors for expression in a yeast such as *S. cerevisiae* include pYepSec I (Baldari, *et al.*, 1987), pMFa (Kurjan and Herskowitz, 1982), pJRY88 (Schultz *et al.*, 1987), and pYES2 (Invitrogen Corporation, San Diego, CA).

Alternatively, an *Alloiococcus otitidis* polynucleotide is expressed in insect cells using, for example, baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (*e.g.*, Sf 9 or Sf 21 cells) include the pAc series (Smith *et al.*, 1983) and the pVL series (Lucklow and Summers, 1989).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed, 1987) and pMT2PC (Kaufman *et al.*, 1987). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements.

As used herein, a promoter is a region of a DNA molecule typically within about 100 nucleotide pairs in front of (upstream of) the point at which transcription begins (*i.e.*, a transcription start site). That region typically contains several types of DNA sequence elements that are located in similar relative positions in different genes. As used herein, the term "promoter" includes what is referred to in the art as an upstream promoter region, a promoter region or a promoter of a generalized eukaryotic RNA Polymerase II transcription unit.

Another type of discrete transcription regulatory sequence element is an enhancer. An enhancer provides specificity of time, location and expression level for a particular encoding region (e.g., gene). A major function of an enhancer is to increase the level of transcription of a coding sequence in a cell that contains one or more transcription factors that bind to that enhancer. Unlike a promoter, an enhancer can function when located at variable distances from transcription start sites so long as a promoter is present.

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As used herein, the phrase "enhancer-promoter" means a composite unit that contains both enhancer and promoter elements. An enhancer-promoter is operatively linked to a coding sequence that encodes at least one gene product. As used herein, the phrase "operatively linked" means that an enhancer-promoter is connected to a coding sequence in such a way that the transcription of that coding sequence is controlled and regulated by that enhancer-promoter. Means for operatively linking an enhancer-promoter to a coding sequence are well known in the art. As is also well known in the art, the precise orientation and location relative to a coding sequence whose transcription is controlled, is dependent *inter alia* upon the specific nature of the enhancer-promoter. Thus, a TATA box minimal promoter is typically located from about 25 to about 30 base pairs upstream of a transcription initiation site and an upstream promoter element is typically located from about 100 to about 200 base pairs upstream of a transcription initiation site. In contrast, an enhancer can be located downstream from the initiation site and can be at a considerable distance from that site.

An enhancer-promoter used in a vector construct of the present invention can be any enhancer-promoter that drives expression in a cell to be transfected. By employing an enhancer-promoter with well-known properties, the level and pattern of gene product expression can be optimized.

For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus (CMV) and Simian Virus 40 (SV40). For other suitable expression systems for both prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook *et al.*, "Molecular Cloning: A Laboratory Manual" 2nd, ed, Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989, incorporated herein by reference.

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In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (*e.g.*, tissue-specific regulatory elements are used to express the nucleic acid). Tissue- specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert *et al.*, 1987), lymphoid-specific promoters (Calame and Eaton, 1988), in particular promoters of T cell receptors (Winoto and Baltimore, 1989) and immunoglobulins (Banerji *et al.*, 1983), Queen and Baltimore (1983), neuron-specific promoters (*e.g.*, the neurofilament promoter; Byrne and Ruddle, 1989), pancreas-specific promoters (Edlund *et al.*, 1985), and mammary gland-specific promoters (*e.g.*, milk whey promoter; U.S. 4, 873,316 and EP 264,166). Developmentally-regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss, 1990) and the α-fetoprotein promoter (Campes and Tilghman, 1989).

The invention further provides a recombinant expression vector comprising a DNA molecule encoding an *Alloiococcus otitidis* polypeptide cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to *Alloiococcus otitidis* mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced.

Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but also to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein. A host cell can be any prokaryotic or eukaryotic cell. For example, an *Alloiococcus otitidis* polypeptide can be expressed in bacterial cells such as *Escherichia coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO), NIH3T3, PER C6, NSO, VERO or COS cells). Other suitable host cells are known to those skilled in the art.

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Vector DNA is can be introduced into prokaryotic or eukaryotic cells *via* conventional transformation, infection or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, protoplast fusion, direct microinfection. Another recognized technique for introducing DNA into a host cell is "infection", such as by adenovirus infection or electroporation. Suitable methods for transforming, infecting or transfecting host cells can be found in Sambrook, *et al.* ("Molecular Cloning: A Laboratory Manual" 2nd ed, Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory manuals.

The most widely used method is transfection mediated by either calcium phosphate or DEAE-dextran. Although the mechanism remains unclear, it is believed that the transfected DNA enters the cytoplasm of the cell by endocytosis and is transported to the nucleus. Depending on the cell type, up to 90% of a population of cultured cells can be transfected at any one time. Because of its high efficiency, transfection mediated by calcium phosphate or DEAE-dextran is the method of choice for experiments that require transient expression of the foreign DNA in large numbers of cells. Calcium phosphate-mediated transfection is also used to establish cell lines that integrate copies of the foreign DNA, which are usually arranged in head-to-tail tandem arrays into the host cell genome.

In the protoplast fusion method, protoplasts derived from bacteria carrying high numbers of copies of plasmid of interest are mixed directly with cultured mammalian cells. After fusion of the cell membranes (usually with polyethylene glycol), the contents of the bacteria are delivered into the cytoplasm of the mammalian cells and the plasmid DNA is transported to the nucleus. Protoplast fusion is not as efficient as transfection for many of the cell lines that are commonly used for transient expression assays, but it is useful for cell lines in which endocytosis of DNA occurs inefficiently. Protoplast fusion frequently yields multiple copies of the plasmid DNA tandemly integrated into the host chromosome.

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The application of brief, high-voltage electric pulses (electroporation) to a variety of mammalian and plant cells leads to the formation of nanometer-sized pores in the plasma membrane. DNA is taken directly into the cell cytoplasm either through these pores or as a consequence of the redistribution of membrane components that accompanies closure of the pores. Electroporation can be extremely efficient and can be used both for transient expression of cloned genes and for establishment of cell lines that carry integrated copies of the gene of interest. Electroporation, in contrast to calcium phosphate-mediated transfection and protoplast fusion, frequently gives rise to cell lines that carry one, or at most a few, integrated copies of the foreign DNA.

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Liposome transfection involves encapsulation of DNA and RNA within liposomes, followed by fusion of the liposomes with the cell membrane. The mechanism of how DNA is delivered into the cell is unclear, but transfection efficiencies can be as high as 90%.

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Direct microinjection of a DNA molecule into nuclei has the advantage of not exposing DNA to cellular compartments such as low-pH endosomes. Microinjection therefore used primarily as a method to establish lines of cells that carry integrated copies of the DNA of interest.

The use of adenovirus as a vector for cell transfection is well known in the art. Adenovirus vector-mediated cell transfection has been reported for various cells (Stratford-Perricaudet, *et al.* 1992).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, is used to produce (i.e., express) an Alloiococcus otitidis polypeptide.

Accordingly, the invention further provides methods for producing an Alloiococcus

otitidis polypeptide using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding an *Alloiococcus otitidis* polypeptide has been introduced) in a suitable medium until the *Alloiococcus otitidis* polypeptide is produced. In another embodiment, the method further comprises isolating the *Alloiococcus otitidis* polypeptide from the medium or the host cell.

A coding sequence of an expression vector is operatively linked to a transcription-terminating region. RNA polymerase transcribes an encoding DNA sequence through a site where polyadenylation occurs. Typically, DNA sequences located a few hundred base pairs downstream of the polyadenylation site serve to terminate transcription. Those DNA sequences are referred to herein as transcription-termination regions. Those regions are required for efficient polyadenylation of transcribed messenger RNA (mRNA). Transcription-terminating regions are well known in the art. A preferred transcription-terminating region used in an adenovirus vector construct of the present invention comprises a polyadenylation signal of SV40 or the protamine gene.

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An expression vector comprises a polynucleotide that encodes an *Alloiococcus otitidis* polypeptide. Such a polypeptide is meant to include a sequence of nucleotide bases encoding an *Alloiococcus otitidis* polypeptide sufficient in length to distinguish the segment from a polynucleotide segment encoding a non-*Alloiococcus otitidis* polypeptide. A polypeptide of the invention can also encode biologically functional polypeptides or peptides which have variant amino acid sequences, such as with changes selected based on considerations such as the relative hydropathic score of the amino acids being exchanged. These variant sequences are those isolated from natural sources or induced in the sequences disclosed herein using a mutagenic procedure such as site-directed mutagenesis.

Preferably, an expression vector of the present invention comprises a polynucleotide that encodes a polypeptide comprising the amino acid residue sequence of one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO:.4036 An expression vector can include an *Alloiococcus otitidis* polypeptide coding region itself of any of the *Alloiococcus otitidis* polypeptides noted above or it can contain coding regions bearing selected alterations or modifications in the basic coding region of such an *Alloiococcus otitidis* polypeptide. Alternatively,

such vectors or fragments can also encode larger polypeptides or polypeptides which nevertheless include the basic coding region. In any event, it should be appreciated that due to codon redundancy as well as biological functional equivalence, this aspect of the invention is not limited to the particular DNA molecules corresponding to the polypeptide sequences noted above.

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Exemplary vectors include the mammalian expression vectors of the pCMV family including pCMV6b and pCMV6c (Chiron Corp., Emeryville CA.). In certain cases, and specifically in the case of these individual mammalian expression vectors, the resulting constructs can require co-transfection with a vector containing a selectable marker such as pSV2neo. *Via* co-transfection into a dihydrofolate reductase-deficient Chinese hamster ovary cell line, such as DG44, clones expressing *Alloiococcus otitidis* polypeptides by virtue of DNA incorporated into such expression vectors can be detected.

A DNA molecule of the present invention can be incorporated into a vector by a number of techniques that are well known in the art. For instance, the vector pUC18 has been demonstrated to be of particular value in cloning and expression of genes. Likewise, the related vectors M13mp18 and M13mp19 can also be used in certain embodiments of the invention, in particular, in performing dideoxy sequencing.

An expression vector of the present invention is useful both as a means for preparing quantities of the *Alloiococcus otitidis* polypeptide-encoding DNA itself, and as a means for preparing the encoded polypeptide and peptides. It is contemplated that where *Alloiococcus otitidis* polypeptides of the invention are made by recombinant means, one can employ either prokaryotic or eukaryotic expression vectors as shuttle systems. In another aspect, the recombinant host cells of the present invention are prokaryotic host cells. Preferably, the recombinant host cells of the invention are bacterial cells of the DH5α strain of *Escherichia coli*. In general, prokaryotes are preferred for the initial cloning of DNA sequences and constructing the vectors useful in the invention. For example, *Escherichia coli* K12 strains can be particularly useful. Other microbial strains that can be used include *Escherichia coli* B, *Escherichia coli* W3110 (ATCC No. 273325) and *Escherichia. coli*<sub>X</sub>1976 (ATCC No. 31537). *Bacilli* such as *Bacillus subtilis*, or other enterobacteriaceae such as *Salmonella typhimurium* or other *Salmonella* species or *Serratia marcesans*, and

various pseudomonas species can be used. These examples are, of course, intended to be illustrative rather than limiting.

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In general, plasmid vectors containing replicon and control sequences that are derived from species compatible with the host cell are used in connection with these hosts. The vector ordinarily carries a replication site, as well as marking sequences that are capable of providing phenotypic selection in transformed cells. For example, *Escherichia coli* can be transformed using pBR322, a plasmid derived from an *Escherichia coli* species (Bolivar, *et al.* 1977). pBR322 contains genes for ampicillin and tetracycline resistance and thus provides easy means for identifying transformed cells. The pBR322 plasmid, or other microbial plasmid or phage, must also contain, or be modified to contain, promoters which can be used by the microbial organism for expression of its own polypeptides.

Those promoters most commonly used in recombinant DNA construction include the β-lactamase (penicillinase) and lactose promoter systems (Chang, *et al.* 1978; Itakura., *et al.* 1977, Goeddel, *et al.* 1979; Goeddel, *et al.* 1980) and a tryptophan (TRP) promoter system (EP 0036776; Siebwenlist *et al.* 1980). While these are the most commonly used, other microbial promoters have been discovered and utilized, and details concerning their nucleotide sequences have been published, enabling a skilled worker to introduce functional promoters into plasmid vectors (Siebwenlist, *et al.* 1980).

In addition to prokaryotes, eukaryotic microbes such as yeast can also be used. Saccharomyces cerevisiase or common baker's yeast is the most commonly used among eukaryotic microorganisms, although a number of other strains are commonly available. For expression in Saccharomyces, the plasmid YRp7, for example, is commonly used (Stinchcomb, et al. 1979; Kingsman, et al. 1979; Tschemper, et al. 1980). This plasmid already contains the trpl gene that provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example ATCC No. 44076 or PEP4-1 (Jones, 1977). The presence of the trpl lesion as a characteristic of the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan.

Suitable promoter sequences in yeast vectors include the promoters for 3-phosphoglycerate kinase (PGK) (Hitzeman, et al. 1980) or other glycolytic enzymes (Hess, et al. 1968; Holland, et al. 1978) such as enolase, glyceraldehyde-3-

phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase. In constructing suitable expression plasmids, the termination sequences associated with these genes are also introduced into the expression vector downstream from the sequences to be expressed to provide polyadenylation of the mRNA and termination. Other promoters, which have the additional advantage of transcription controlled by growth conditions are the promoter region for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, and the aforementioned glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Any plasmid vector containing a yeast-compatible promoter, origin of replication, and termination sequences is suitable.

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In addition to microorganisms, cultures of cells derived from multicellular organisms can also be used as hosts. In principle, any such cell culture is workable, whether from vertebrate or invertebrate culture. However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years. Examples of such useful host cell lines are AtT-20, VERO, HeLa, NSO, PER C6, Chinese hamster ovary (CHO) cell lines, W138, BHK, COSM6, COS-7, 293, VERO and MDCK cell lines. Expression vectors for such cells ordinarily include (if necessary) an origin of replication, a promoter located upstream of the gene to be expressed, along with any necessary ribosome binding sites, RNA splice sites, polyadenylation site, and transcriptional terminator sequences.

Where expression of recombinant *Alloiococcus otitidis* polypeptides is desired and a eukaryotic host is contemplated, it is most desirable to employ a vector, such as a plasmid, that incorporates a eukaryotic origin of replication. Additionally, for the purposes of expression in eukaryotic systems, one desires to position the *Alloiococcus otitidis* encoding sequence adjacent to and under the control of an effective eukaryotic promoter such as promoters used in combination with Chinese hamster ovary cells (CHO). To bring a coding sequence under control of a promoter, whether it is eukaryotic or prokaryotic, what is generally needed is to position the 5' end of the translation initiation side of the proper translational reading frame of the

polypeptide between about 1 and about 50 nucleotides 3' of or downstream with respect to the promoter chosen. Furthermore, where eukaryotic expression is anticipated, one would typically desire to incorporate an appropriate polyadenylation site into the transcriptional unit that includes the *Alloiococcus otitidis* polypeptide.

A transfected cell can be prokaryotic or eukaryotic. Preferably, the host cells of the invention are prokaryotic host cells. Where it is of interest to produce an *Alloiococcus otitidis* polypeptide, cultured prokaryotic host cells are of particular interest.

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In yet another embodiment, the present invention contemplates a process or method of preparing *Alloiococcus otitidis* polypeptides comprising transfecting, transforming or infecting cells with a polynucleotide that encodes an *Alloiococcus otitidis* polypeptide to produce transformed host cells; and maintaining the transformed host cells under biological conditions sufficient for expression of the polypeptide. Preferably, the transformed host cells are prokaryotic cells. Alternatively, the host cells are eukaryotic cells. More preferably, the prokaryotic cells are bacterial cells of the DH5 $\alpha$  strain of *Escherichia coli*. Even more preferably, the polynucleotide transfected into the transformed cells comprises the nucleic acid sequence of one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105. Additionally, transfection is accomplished using an expression vector disclosed above. A host cell used in the process is capable of expressing a functional, recombinant *Alloiococcus otitidis* polypeptide.

Following transfection, the cell is maintained under culture conditions for a period of time sufficient for expression of an *Alloiococcus otitidis* polypeptide. Culture conditions are well known in the art and include ionic composition and concentration, temperature, pH and the like. Typically, transfected cells are maintained under culture conditions in a culture medium. Suitable media for various cell types are well known in the art. In a preferred embodiment, temperature is from about 20°C to about 50°C, more preferably from about 30°C to about 40°C and, even more preferably about 37°C.

The pH is preferably from about a value of 6.0 to a value of about 8.0, more preferably from about a value of about 6.8 to a value of about 7.8 and, most preferably about 7.4. Osmolality is preferably from about 200 milliosmols per liter (mosm/L) to about 400 mosm/l and, more preferably from about 290 mosm/L to

about 310 mosm/L. Other biological conditions needed for transfection and expression of an encoded protein are well known in the art.

Transfected cells are maintained for a period of time sufficient for expression of an *Alloiococcus otitidis* polypeptide. A suitable time depends *inter alia* upon the cell type used and is readily determinable by a skilled artisan. Typically, maintenance time is from about 2 to about 14 days.

Recombinant *Alloiococcus otitidis* polypeptide is recovered or collected either from the transfected cells or the medium in which those cells are cultured. Recovery comprises isolating and purifying the *Alloiococcus otitidis* polypeptide. Isolation and purification techniques for polypeptides are well known in the art and include such procedures as precipitation, filtration, chromatography, electrophoresis and the like.

## F. ANTIBODIES IMMUNOREACTIVE WITH ALLOIOCOCCUS OTITIDIS POLYPEPTIDES

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In still another embodiment, the present invention provides antibodies immunoreactive with *Alloiococcus otitidis* polypeptides. Preferably, the antibodies of the invention are monoclonal antibodies. Additionally, the *Alloiococcus otitidis* polypeptides comprise the amino acid residue sequence of one of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106. Means for preparing and characterizing antibodies are well known in the art (*See, e.g.*, Antibodies "A Laboratory Manual", E. Howell and D. Lane, Cold Spring Harbor Laboratory, 1988). Polyclonal antisera is obtained by bleeding an immunized animal into a glass or plastic container, incubating the blood at 25°C for one hour, followed by incubating at 4°C for 2-18 hours. The serum is then recovered by centrifugation.

Briefly, a polyclonal antibody is prepared by immunizing an animal with an immunogen comprising a polypeptide or polynucleotide of the present invention, and collecting antisera from that immunized animal. A wide range of animal species can be used for the production of antisera. Typically an animal used for production of anti-antisera is a rabbit, a mouse, a rat, a hamster or a guinea pig. Because of the relatively large blood volume of rabbits, a rabbit is a preferred choice for production of polyclonal antibodies.

As is well known in the art, a given polypeptide or polynucleotide may vary in its immunogenicity. It is often necessary therefore to couple the immunogen (e.g., a polypeptide or polynucleotide) of the present invention with a carrier. Exemplary and

preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin or rabbit serum albumin can also be used as carriers.

Means for conjugating a polypeptide or a polynucleotide to a carrier protein are well known in the art and include glutaraldehyde, m-maleimidobencoyl-N-hydroxysuccinimide ester, carbodiimide and bis-biazotized benzidine.

As is also well known in the art, immunogencity to a particular immunogen can be enhanced by the use of non-specific stimulators of the immune response known as adjuvants. Exemplary and preferred adjuvants include complete Freund's adjuvant, incomplete Freund's adjuvants, cholera toxin (e.g. mutant cholera toxin E29H; see published International Patent Application WO 00/18434), and aluminum hydroxide adjuvant.

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The amount of immunogen used for the production of polyclonal antibodies depends upon the nature of the immunogen as well as the animal used for immunization. A variety of routes can be used to administer the immunogen (subcutaneous, intramuscular, intradermal, intravenous and intraperitoneal). The production of polyclonal antibodies is monitored by sampling blood from the immunized animal at various points following immunization. When a desired level of immunogenicity is obtained, the immunized animal can be bled and the serum isolated and stored.

In another aspect, the present invention contemplates a process of producing an antibody immunoreactive with an *Alloiococcus otitidis* polypeptide comprising the steps of (a) transfecting recombinant host cells with a polynucleotide that encodes an *Alloiococcus otitidis* polypeptide; (b) culturing the host cells under conditions sufficient for expression of the polypeptide; (c) recovering the polypeptides; and (d) preparing the antibodies to the polypeptides. Preferably, the host cell is transfected with the polynucleotide of one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 4035. Even more preferably, the present invention provides antibodies prepared according to the process described above.

A monoclonal antibody of the present invention can be readily prepared through use of well-known techniques such as those exemplified in U.S. Pat. No. 4,196,265, herein incorporated by reference. Typically, a technique involves first immunizing a suitable animal with a selected antigen (e.g., a polypeptide or

polynucleotide of the present invention) in a manner sufficient to provide an immune response. Rodents such as mice and rats are preferred animals. Spleen cells from the immunized animal are then fused with cells of an immortal myeloma cell. Where the immunized animal is a mouse, a preferred myeloma cell is a murine NS-1 myeloma cell.

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The fused spleen/myeloma cells are cultured in a selective medium to select fused spleen/myeloma cells from the parental cells. Fused cells are separated from the mixture of non-fused parental cells, *e.g.*, by the addition of agents that block the *de novo* synthesis of nucleotides in the tissue culture media. Exemplary and preferred agents are aminopterin, methotrexate, and azaserine. Aminopterin and methotrexate block *de novo* synthesis of both purines and pyrimidines, whereas azaserine blocks only purine synthesis. Where aminopterin or methotrexate is used, the media is supplemented with hypoxanthine and thymidine as a source of nucleotides. Where azaserine is used, the media is supplemented with hypoxanthine.

This culturing provides a population of hybridomas from which specific hybridomas are selected. Typically, selection of hybridomas is performed by culturing the cells by single-clone dilution in microtiter plates, followed by testing the individual clonal supernatants for reactivity with an antigen-polypeptide. The selected clones can then be propagated indefinitely to provide the monoclonal antibody.

By way of specific example, to produce an antibody of the present invention, mice are injected intraperitoneally with between about 1-200 µg of an antigen comprising a polypeptide of the present invention. B lymphocyte cells are stimulated to grow by injecting the antigen in association with an adjuvant such as complete Freund's adjuvant (CFA; a non-specific stimulator of the immune response containing killed Mycobacterium tuberculosis). At some time (e.g., at least two weeks) after the first injection, mice are boosted by injection with a second dose of the antigen mixed with incomplete Freund's adjuvant (IFA; lacks the killed mycobacterium of CFA).

A few weeks after the second injection, mice are tail bled and the sera titered by immunoprecipitation against radiolabeled antigen. Preferably, the process of boosting and titering is repeated until a suitable titer is achieved. The spleen of the mouse with the highest titer is removed and the spleen lymphocytes are obtained by

homogenizing the spleen with a syringe. Typically, a spleen from an immunized mouse contains approximately  $5x10^7$  to  $2x10^8$  lymphocytes.

Mutant lymphocyte cells known as myeloma cells are obtained from laboratory animals in which such cells have been induced to grow by a variety of well-known methods. Myeloma cells lack the salvage pathway of nucleotide biosynthesis. Because myeloma cells are tumor cells, they can be propagated indefinitely in tissue culture, and are thus denominated immortal. Numerous cultured cell lines of myeloma cells from mice and rats, such as murine NS-1 myeloma cells, have been established.

Myeloma cells are combined under conditions appropriate to foster fusion with the normal antibody-producing cells from the spleen of the mouse or rat injected with the antigen/polypeptide of the present invention. Fusion conditions include, for example, the presence of polyethylene glycol. The resulting fused cells are hybridoma cells. Like myeloma cells, hybridoma cells grow indefinitely in culture.

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Hybridoma cells are separated from unfused myeloma cells by culturing in a selection medium such as HAT media (hypoxanthine, aminopterin, thymidine). Unfused myeloma cells lack the enzymes necessary to synthesize nucleotides from the salvage pathway because they are killed in the presence of aminopterin, methotrexate, or azaserine. Unfused lymphocytes also do not continue to grow in tissue culture. Thus, only cells that have successfully fused (hybridoma cells) can grow in the selection media.

Each of the surviving hybridoma cells produces a single antibody. These cells are then screened for the production of the specific antibody immunoreactive with an antigen/polypeptide of the present invention. Single cell hybridomas are isolated by limiting dilutions of the hybridomas. The hybridomas are serially diluted many times and, after the dilutions are allowed to grow, the supernatant is tested for the presence of the monoclonal antibody. The clones producing that antibody are then cultured in large amounts to produce an antibody of the present invention in convenient quantity.

By use of a monoclonal antibody of the present invention, specific polypeptides and polynucleotide of the invention are identified as antigens. Once identified, those polypeptides and polynucleotide are isolated and purified by techniques such as antibody-affinity chromatography. In antibody-affinity

chromatography, a monoclonal antibody is bound to a solid substrate and exposed to a solution containing the desired antigen. The antigen is removed from the solution through an immunospecific reaction with the bound antibody. The polypeptide or polynucleotide is then easily removed from the substrate and purified.

Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, U.S. 5,223,409; WO 92/18619; WO 91/17271; WO 92/20791; WO 92/15679; WO 93/01288; WO 92/01047; WO 92/09690; WO 90/02809, which are incorporated herein in their entirety by reference.

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Additionally, recombinant anti-*Alloiococcus otitidis* antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human fragments, which are made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies are produced by recombinant DNA techniques known in the art, for example using methods described in PCT/US86/02269; EP 184,187; EP 171,496; EP 173,494; WO 86/01533; U.S. 4,816,567; and EP 125,023.

An anti-Alloiococcus otitidis antibody (e.g., monoclonal antibody) is used to isolate Alloiococcus otitidis polypeptides by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-Alloiococcus otitidis antibody facilitates the purification of a natural Alloiococcus otitidis polypeptide from cells and recombinantly produced Alloiococcus otitidis polypeptides expressed in host cells. Moreover, an anti-Alloiococcus otitidis antibody is used to detect Alloiococcus otitidis polypeptide (e.g., in a cellular lysate or cell supernatant) in order to evaluate the abundance of the Alloiococcus otitidis polypeptide. The detection of circulating fragments of an Alloiococcus otitidis polypeptide is used to identify Alloiococcus otitidis polypeptide turnover in a subject. Anti-Alloiococcus otitidis antibodies are used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, e.g., to, for example, determine the efficacy of a given treatment regimen. Detection is facilitated by coupling (i.e., physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, P-galactosidase, or

acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylarnine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and acquorin, and examples of suitable radioactive material include 125 l, 131 l, 15 or 3 H.

## G. PHARMACEUTICAL COMPOSITIONS

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In certain embodiments, the present invention provides pharmaceutical compositions comprising compounds that inhibit the activities of *Alloiococcus otitidis* polypeptides, and physiologically acceptable carriers. Compounds that inhibit the activities of *Alloiococcus otitidis* polypeptides polypeptides, which are essential for the proliferation of the bacteria, are identified using one or more assay systems set forth in Examples 5-38. More preferably, the pharmaceutical compositions comprise one or more compounds that inhibit the activities of *Alloiococcus otitidis* polypeptides comprising the amino acid residue sequence of one or more of the even numbered sequences set forth in SEQ ID NO: 2 through SEQ ID NO: 106. In other embodiments, the pharmaceutical compositions of the invention comprise antisense polynucleotides of polynucleotides selected from one of the odd numbered sequences set forth in Seq. ID NO: 1 to Seq. ID No. 105, and physiologically acceptable carriers.

Various tests are to be used to assess the *in vitro* and *in vivo* efficacy of anitmicrobial and pharmaceutical compounds that inhibit the activities of *Alloiococcus otitidis* polypeptides, and these are set forth in detail in Examples 5 through 38. For example, an *in vitro* activity of the compounds may be assayed by incubating together a mixture of *Alloiococcus otitidis* or other heterologous bacterial cells such as *E. coli* cells expressing *Alloiococcus otitidis* polypeptides set forth in one of the even numbered sequences from Seq. ID No. 2 to Seq. ID No. 106, and then measuring the activity of the polypeptide using one or more of the assay systems detailed in Example 5 through 38.

The Alloiococcus otitidis polynucleotides, polypeptides, compounds that modulate the activity of an Alloiococcus otitidis polypeptides, and anti-Alloiococcus

otitidis antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration to a host or subject, e.g., a human. Such compositions typically comprise the nucleic acid molecule, protein, antimicrobial compound, or antibody and a pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, such media can be used in the compositions of the invention. Supplementary active compounds can also be incorporated into the compositions.

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A pharmaceutical of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, (e.g., intravenous, intradermal, subcutaneous, intraperitoneal), transmucosal (e.g., oral, rectal, intranasal, vaginal, respiratory), and transdermal (topical). Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water-soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor EL<sup>TM</sup>(BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of

can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser that contains a suitable propellant, *e.g.*, a gas such as carbon dioxide, or a nebulizer. Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (*e.g.*, with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems.

Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods

#### H. DIAGNOSTIC ASSAYS

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The invention also provides methods for detecting the presence of an Alloiococcus otitidis polypeptide or Alloiococcus otitidis polynucleotide, or fragment thereof, in a biological sample. The method involves contacting the biological sample

with a compound or an agent capable of detecting an *Alloiococcus otitidis* polypeptide or mRNA such that the presence of the *Alloiococcus otitidis* polypeptide/encoding nucleic acid molecule is detected in the biological sample. A preferred agent for detecting *Alloiococcus otitidis* mRNA or DNA is a labeled or labelable oligonucleotide probe capable of hybridizing to *Alloiococcus otitidis* mRNA or DNA. The nucleic acid probe can be, for example, a full-length *Alloiococcus otitidis* polynucleotide of one of the odd numbered sequences set forth in SEQ ID NO: 1 through SEQ ID NO: 105, a complement thereof, or a fragment thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to *Alloiococcus otitidis* mRNA or DNA. Alternatively, the sample can be contacted with an oligonucleotide primer of an *Alloiococcus otitidis* polynucleotide of SEQ ID NO: 1 through SEQ ID :105, a complement thereof, or a fragment thereof, in the presence of nucleotides and a polymerase, under conditions permitting primer extension.

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A preferred agent for detecting Alloiococcus otitidis polypeptide is a labeled or labelable antibody capable of binding to an Alloiococcus otitidis polypeptide. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab')2) can be used. The term "labeled or labelable," with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect Alloiococcus otitidis mRNA, DNA or protein in a biological sample in vitro as well as in vivo. For example, in vitro techniques for detection of Alloiococcus otitidis mRNA include Northern hybridizations and in situ hybridizations. In vitro techniques for detection of Alloiococcus otitidis polypeptide include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. Alternatively, Alloiococcus otitidis

polypeptides can be detected *in vivo* in a subject by introducing into the subject a labeled anti-*Alloiococcus otitidis* antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

The polynucleotides according to the invention may also be used in analytical DNA chips, which allow sequencing, the study of mutations and of the expression of genes, and which are currently of interest given their very small size and their high capacity in terms of number of analyses.

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The principle of the operation of these chips is based on molecular probes, most often oligonucleotides, which are attached onto a miniaturized surface, generally of the order of a few square centimeters. During an analysis, a sample containing fragments of a target nucleic acid to be analyzed, for example DNA or RNA labeled, for example, after amplification, is deposited onto the DNA chip in which the support has been coated beforehand with probes. Bringing the labeled target sequences into contact with the probes leads to the formation, through hybridization, of a duplex according to the rule of pairing defined by J.D. Watson and F. Crick. After a washing step, analysis of the surface of the chip allows the effective hybridizations to be located by means of the signals emitted by the labels tagging the target. A hybridization fingerprint results from this analysis which, by appropriate computer processing, will make it possible to determine information such as the presence of specific fragments in the sample, the determination of sequences and the presence of mutations.

The chip consists of a multitude of molecular probes, precisely organized or arrayed on a solid support whose surface is miniaturized. It is at the center of a system where other elements (imaging system, microcomputer) allow the acquisition and interpretation of a hybridization fingerprint.

The hybridization supports are provided in the form of flat or porous surfaces (pierced with wells) composed of various materials. The choice of a support is determined by its physicochemical properties, or more precisely, by the relationship between the latter and the conditions under which the support will be placed during the synthesis or the attachment of the probes or during the use of the chip. It is therefore necessary, before considering the use of a particular support, to consider characteristics such as its stability to pH, its physical strength, its reactivity and its

chemical stability as well as its capacity to nonspecifically bind nucleic acids. Materials such as glass, silicon and polymers are commonly used. Their surface is, in a first step, called "functionalization", made reactive towards the groups which it is desired to attach thereon. After the functionalization, so-called spacer molecules are grafted onto the activated surface. Used as intermediates between the surface and the probe, these molecules of variable size render unimportant the surface properties of the supports, which often prove to be problematic for the synthesis or the attachment of the probes and for the hybridization.

Among the hybridization supports, there may be mentioned glass which is used, for example, in the method of *in situ* synthesis of oligonucleotides by photochemical addressing developed by the company Affymetrix (E.L. Sheldon, 1993), the glass surface being activated by silane. Genosensor Consortium (P. Mérel, 1994) also uses glass slides carrying wells 3 mm apart, this support being activated with epoxysilane.

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The probes according to the invention may be synthesized directly *in situ* on the supports of the DNA chips. This *in situ* synthesis may be carried out by photochemical addressing (developed by the company Affymax (Amsterdam, Holland) and exploited industrially by its subsidiary Affymetrix (United States)) or based on the VLSIPS (very large scale immobilized polymer synthesis) technology (S.P.A. Fodor *et al.*, 1991) which is based on a method of photochemically directed combinatory synthesis and the principle of which combines solid-phase chemistry, the use of photolabile protecting groups and photolithography.

The probes according to the invention may be attached to the DNA chips in various ways such as electrochemical addressing, automated addressing or the use of probe printers (T. Livache *et al.*, 1994; G. Yershov *et al.*, 1996; J. Derisi *et al.*, 1996, and S. Borman, 1996).

The revealing of the hybridization between the probes of the invention, deposited or synthesized *in situ* on the supports of the DNA chips, and the sample to be analyzed, may be determined, for example, by measurement of fluorescent signals, by radioactive counting or by electronic detection.

The use of fluorescent molecules such as fluorescein constitutes the most common method of labeling the samples. It allows direct or indirect revealing of the hybridization and allows the use of various fluorochromes.

Affymetrix currently provides an apparatus or a scanner designed to read its Gene Chip™ chips. It makes it possible to detect the hybridizations by scanning the surface of the chip in confocal microscopy (R.J. Lipshutz *et al.*, 1995).

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The nucleotide sequences according to the invention are also used in DNA chips to carry out the analysis of the expression of the Alloiococcus otitidis genes. This analysis of the expression of Alloiococcus otitidis genes is based on the use of chips where probes of the invention, chosen for their specificity to characterize a given gene, are present (D.J. Lockhart et al., 1996; D.D. Shoemaker et al., 1996). For the methods of analysis of gene expression using the DNA chips, reference may, for example, be made to the methods described by D.J. Lockhart et al. (1996) and Sosnowsky et al. (1997) for the synthesis of probes in situ or for the addressing and the attachment of previously synthesized probes. The target sequences to be analyzed are labeled and in general fragmented into sequences of about 50 to 100 nucleotides before being hybridized onto the chip. After washing as described, for example, by D.J. Lockhart et al. (1996) and application of different electric fields (Sosnowsky et al., 1997), the labeled compounds are detected and quantified, the hybridizations being carried out at least in duplicate. Comparative analyses of the signal intensities obtained with respect to the same probe for different samples and/or for different probes with the same sample, determine the differential expression of RNA or of DNA derived from the sample.

The nucleotide sequences according to the invention are, in addition, used in DNA chips where other nucleotide probes specific for other microorganisms are also present, and allow the carrying out of a serial test allowing rapid identification of the presence of a microorganism in a sample.

Accordingly, the subject of the invention is also the nucleotide sequences according to the invention, characterized in that they are immobilized on a support of a DNA chip.

The DNA chips, characterized in that they contain at least one nucleotide sequence according to the invention, immobilized on the support of the said chip, also form part of the invention.

The chips preferably contain several probes or nucleotide sequences of the invention of different length and/or corresponding to different genes so as to identify, with greater certainty, the specificity of the target sequences or the desired mutation

in the sample to be analyzed.

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Accordingly, the analyses carried out by means of primers and/or probes according to the invention, immobilized on supports such as DNA chips, make it possible, for example, to identify, in samples, mutations linked to variations such as intraspecies variations. These variations may be correlated or associated with pathologies specific to the variant identified and make it possible to select the appropriate treatment.

The invention thus comprises a DNA chip according to the invention, characterized in that it contains, in addition, at least one nucleotide sequence of a microorganism different from *Alloiococcus otitidis*, immobilized on the support of the said chip; preferably, the different microorganism is chosen from an associated microorganism, a bacterium of the *Streptococcus* family, and a variant of the species *Alloiococcus otitidis*.

The principle of the DNA chip as explained above, is also used to produce protein "chips" on which the support has been coated with a polypeptide or an antibody according to the invention, or arrays thereof, in place of the DNA. These protein "chips" make it possible, for example, to analyze the biomolecular interactions (BIA) induced by the affinity capture of target analytes onto a support coated, for example, with proteins, by surface plasma resonance (SPR). Reference may be made, for example, to the techniques for coupling proteins onto a solid support which are described in EP 524 800 or to the methods describing the use of biosensor-type protein chips such as the BIAcore-type technique (Pharmacia) (Arlinghaus et al., 1997, Krone et al., 1997, Chatelier et al., 1995). These polypeptides or antibodies according to the invention, capable of specifically binding antibodies or polypeptides derived from the sample to be analyzed, are thus used in protein chips for the detection and/or the identification of proteins in samples. The said protein chips may in particular be used for infectious diagnosis and preferably contain, per chip, several polypeptides and/or antibodies of the invention of different specificity, and/or polypeptides and/or antibodies capable of recognizing microorganisms different from Alloiococcus otitidis.

Accordingly, the subject of the present invention is also the polypeptides and the antibodies according to the invention, characterized in that they are immobilized on a support, in particular, on a protein chip.

The protein chips, characterized in that they contain at least one polypeptide or one antibody according to the invention immobilized on the support of the said chip, also form part of the invention.

The invention comprises, in addition, a protein chip according to the invention, characterized in that it contains, in addition, at least one polypeptide of a microorganism different from *Alloiococcus otitidis* or at least one antibody directed against a compound of a microorganism different from *Alloiococcus otitidis*, immobilized on the support of the chip.

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The invention also relates to a kit or set for the detection and/or the identification of bacteria belonging to the species *Alloiococcus otitidis* or to an associated microorganism, or for the detection and/or the identification of a microorganism characterized in that it comprises a protein chip according to the invention.

The present invention also provides a method for the detection and/or the identification of bacteria belonging to the species *Alloiococcus otitidis* or to an associated microorganism in a biological sample, characterized in that it uses a nucleotide sequence according to the invention.

The invention also encompasses kits for detecting the presence of an *Alloiococcus otitidis* polypeptide in a biological sample. For example, the kit comprises reagents such as a labeled or labelable compound or agent capable of detecting *Alloiococcus otitidis* polypeptide or mRNA in a biological sample; means for determining the amount of *Alloiococcus otitidis* polypeptide in the sample; and means for comparing the amount of *Alloiococcus otitidis* polypeptide in the sample with a standard. The compound or agent are packaged in a suitable container. The kit further comprises instructions for using the kit to detect *Alloiococcus otitidis* mRNA or protein.

In certain embodiments, detection involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g. U.S. 4,683,195 and U.S. 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR). This method includes the steps of collecting a sample of cells from a patient, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to an *Alloiococcus otitidis* polynucleotide under conditions such that

hybridization and amplification of the *Alloiococcus otitidis*-polynucleotide (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample.

### I. TRANSGENIC ANIMALS

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It is contemplated that in some instances the genome of a transgenic animal of the present invention will have been altered through the stable introduction of one or more of the *Alloiococcus otitidis* polynucleotide compositions described herein, either native, synthetically modified or mutated. As described herein, a "transgenic animal" refers to any animal, preferably a non-human mammal (*e.g.* mouse, rat, rabbit, squirrel, hamster, rabbits, guinea pigs, pigs, micro-pigs, baboons, squirrel monkeys and chimpanzees, *etc*), bird or an amphibian, in which one or more cells contain a heterologous nucleic acid sequence introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly, by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical crossbreeding, or *in vitro* fertilization, but rather is directed to the introduction of a recombinant DNA molecule. This molecule may be integrated within a chromosome, or it may be extrachromosomally replicating DNA.

The host cells of the invention are also used to produce non-human transgenic animals. The non-human transgenic animals are used in screening assays designed to identify infections or compounds, *e.g.*, drugs, pharmaceuticals, *etc.*, which are capable of ameliorating *Alloiococcus otitidis* symptoms or infections. For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which an *Alloiococcus otitidis* polypeptide-coding sequence has been introduced. Such host cells are then used to create non-human transgenic animals in which exogenous *Alloiococcus otitidis* gene sequences have been introduced into their genome or homologous recombinant animals in which endogenous *Alloiococcus otitidis* gene sequences have been altered. Such animals are useful for studying the effects of an *Alloiococcus otitidis* polypeptide and for

identifying and/or evaluating modulators of *Alloiococcus otitidis* polypeptide infectivity.

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A transgenic animal of the invention is created by introducing an *Alloiococcus* otitidis polypeptide-encoding nucleic acid sequence into the male pronuclei of a fertilized oocyte, *e.g.*, by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The human *Alloiococcus* otitidis cDNA sequence of one or more of SEQ ID NO:1 through SEQ ID NO: 4035 can be introduced as a transgene into the genome of a non-human animal.

Moreover, a non-Alloiococcus otitidis homologue of the Alloiococcus otitidis gene can be isolated based on hybridization to the Alloiococcus otitidis cDNA (described above) and used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to the Alloiococcus otitidis transgene to direct expression of an Alloiococcus otitidis polypeptide to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are described, for example, in U.S. 4,736,866 and 4,870, 009, U.S. 4,873,191 and in Hogan, 1986. Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of the Alloiococcus otitidis transgene in its genome and/or expression of Alloiococcus otitidis mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding an Alloiococcus otitidis polypeptide can further be bred to other transgenic animals carrying other transgenes.

In another embodiment, transgenic non-human animals can be produced which contain selected systems that allow for regulated expression of the transgene. One example of such a system is the cre/loxP recombinase system of bacteriophage Pλ. For a description of the cre/loxP recombinase system, see, e.g., Lakso et al., 1992. Another example of a recombinase system is the FLP recombinase system of Saccharomyces cerevisiae (O'Gon-nan et al., 1991). If a cre/loxP recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the Cre recombinase and a selected protein are required.

Such animals can be provided through the construction of "double" transgenic animals, *e.g.*, by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut *et al.*, 1997, and PCT International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, *e.g.*, a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter  $G_o$  phase. The quiescent cell can then be fused, *e.g.*, through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The reconstructed oocyte is then cultured such that it develops to morula or blastocyst and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, *e.g.*, the somatic cell, is isolated.

All patents and publications cited herein are hereby incorporated by reference.

The following examples are carried out using standard techniques, which are well known and routine to those of skill in the art, except where otherwise described in detail. The following examples are presented for illustrative purposes, and should not be construed in any way limiting the scope of this invention.

### EXAMPLE 1

## CONFIRMATION OF THE IDENTITY OF THE ALLOIOCOCCUS OTITIDIS 1104-92 ISOLATE

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The Alloiococcus otitidis isolate 1104-92 was obtained from Dr. Richard Facklam of the Centers for Disease Control in Atlanta. It was isolated from the middle ear fluid of a child in the Atlanta, Georgia area. It was confirmed to be A. otitidis by comparing it to the type strain, ATCC51267, obtained from the American Type Culture Collection [Aguirre, 1992 #1]. Both the 1104-92 and type strain are characterized as Gram positive cocci. Both grow on Columbia agar supplemented with 5% yeast extract, 0.5% polysorbate 80 (Tween 80), and 0.7% phospatidyl choline when incubated at 37°C. On this medium, both strains form slow growing

small white colonies that require nearly two days to be easily observed with the naked eye. Both are sensitive to lysis by hen egg white lysozyme and *Streptococcus globisporus* mutanolysin. Both grow in the presence of 2% sodium azide. Both are killed by incubation at 55°C for 30 minutes. Finally, to further confirm that the 1104-92 was a strain of *A. otitidis*, it was subject to polymerase chain reaction (PCR) identification based on its 16s rRNA gene. This was done using two of the primers specified by Aguirre and Collins [Aguirre, 1992 #2]. The antisense primer used was

5'-ATCTTCCTGCTTGCAGGAAGAGG-3' and the sense primer was 3'-CGCTTCATCTCTGAAGCTAGC-5'. Thus by multiple criteria, the 1104-92 strain was confirmed to be an isolate of *A. otitidis*.

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#### **EXAMPLE 2**

## STORAGE, GROWTH, AND HARVEST OF ALLOIOCOCCUS OTITIDIS 1104-92 FOR ISOLATION OF DNA

The A. otitidis isolate 1104-92 was stored at -70°C in Todd-Hewlett broth containing 40% glycerol. A small portion of the frozen stock was streaked onto the agar medium described in Example 1 and incubated at 37°C for two days. The growth from the plate was swabbed into a  $17 \times 100$  cm tube containing 6 ml of a serum-free broth medium. This broth medium was prepared with 30 g Todd-Hewlett medium, 5 g yeast extract, 10 ml polysorbate 80 (Tween 80), and 1 liter distilled water. This medium was sterilized by autoclaving for 35 minutes. The bacteria were incubated aerobically without shaking in an aerobic incubator at 37°C for two days. The tube containing the growing bacteria was then shaken to resuspend the bacteria and added to a liter of the same medium in a Fernbach flask. This flask, in turn, was incubated aerobically for three days without shaking. The bacteria were harvested by first swirling the flask to suspend the bacteria and then low speed centrifugation at about  $5,000 \times g$  for 30 minutes. The pellet of bacteria was washed by resuspending it in 10 to 15 mL of phosphate buffered saline (PBS), and centrifuging the suspension at about  $8,000 \times g$  for 20 minutes. The pellet of bacteria was retained and stored frozen at -20°C. The yield of wet bacterial pellet was typically about 1 g per liter of broth.

# EXAMPLE 3 PREPARATION OF ALLOIOCOCCUS OTITIDIS GENOMIC DNA

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To prepare genomic DNA, 0.95 g frozen pellet of bacteria was defrosted and suspended in 10 mL of PBS containing 1 mM MgCl<sub>2</sub>. The bacteria were killed by incubating the suspension at 55°C for 20 minutes. The suspension was allowed to cool before adding 25 μl of a 10 mg/mL stock of hen egg white lysozyme and 50 μl of a 25,000 unit/mL stock of *Streptococcus globisporus* mutanolysin to the suspension. It was then incubated for one hour at 37°C. Then 50 μl of a 10 mg/mL stock of RNase was added and the suspension incubated an additional hour at 37°C. After these incubations, sodium dodecylsulfate (SDS) was added to a final concentration of 0.3% (0.3 mL of a 10% stock). This was followed by the addition of 0.3 mL of a 1 mg/mL stock of proteinase K. The suspension was then incubated for two hours at 37°C. After this time, an equal volume of water saturated phenol/chloroform/isopropyl (25:24:1) was added to the digested suspension and gently mixed. The upper aqueous layer was retained after a low speed centrifugation and 2.5 volumes of ethanol were added and the tube gently inverted to mix. The DNA was then spooled out on a glass rod and allowed to air dry.

The DNA at this stage still contained obvious impurities and needed further purification. The DNA dried on the glass rod was soaked in 70% ethanol to remove excess phenol and air-dried once again. It was then suspended in 2 ml of Tris-EDTA buffer to which 2 µl of RNase cocktail was added and incubated at room temperature for 75 minutes. Then 100 µl of protease, 100 µl SDS and 40 µl of 100 mM CaCl<sub>2</sub> were added and the suspension incubated for 3.5 hours. An equal volume of chloroform was added, gently mixed, then centrifuged at a low speed. The aqueous layer was collected and re-extracted with the phenol, chloroform, isopropyl alcohol reagent. In turn, the aqueous layer was extracted with chloroform. At this point, 3 M sodium acetate was added to the aqueous phase collected form the last extraction and then 3.75 ml of ethanol was added and gently mixed. The DNA was spooled out, soaked in 70% ethanol and allowed to air-dry. The DNA was finally suspended in 2 ml of Tris-EDTA buffer. Based on absorption at 260 nm, the final yield of DNA was 482 µg of DNA. The DNA was confirmed to be that of *A. otitidis* by the PCR method described in example 1. This DNA was submitted for sequencing.

#### EXAMPLE 4 ·

## CLONING AND SEQUENCING ALLOIOCOCCUS OTITIDIS GENOME

This invention provides nucleotide sequences of the genome of *Alloiococcus otitidis* which thus comprises a DNA sequence library of *Alloiococcus otitidis* genomic DNA. The detailed description that follows provides nucleotide sequences of Alloiococcus otitidis, and also describes how the sequences were obtained and how ORFs (Open Reading Frames) and protein-coding sequences can be identified.

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To construct a library, genomic DNA was hydrodynamically sheared in an HPLC and then separated on a standard 1% agarose gel. A fraction corresponding to 3000-3500 bp in length was excised from the gel and purified by the GeneClean procedure (BIO101, Inc.).

The purified DNA fragments were then blunt-ended using T4 DNA polymerase. The blunt-ended DNA was then ligated to unique BstX1-linker adapters. These linkers are complimentary to the pGTC vector, while the overhang is not self-complimentary. Therefore, the linkers will not concatermerize nor will the cut-vector religate itself easily. The liner-adapted inserts were separated from the unincorporated linkers on a 1% agarose gel and again purified using GeneClean. The linker-adapted inserts were then ligated to BstX1-cut vector to construct "shotgun" subclone libraries.

Only major modifications to the protocols are highlighted. Briefly, the library was transformed into DH10B competent cells (Gibco/BRL, DH5a transformation protocol). Transformed cells were detected by plating onto antibiotic plates containing ampicillin. The plates were incubated overnight at 37° C. Transformant clones were then selected for sequencing. The cultures were grown overnight at 37°C. DNA was purified using a silica bead DNA preparation (Egelstein, 1996) method. In this manner, 25 mg of DNA was obtained per clone.

These purified DNA samples were then sequenced using ABI dye-terminator chemistry. All subsequent steps were based on sequencing by automated DNA sequencing methods. The ABI dye terminator sequence reads were run on MegaBace™ 10000 (Amersham) machines and the data transferred to UNIX based computers. Base calls and quality scores were determined using the PHRED

software program (Ewing et al., 1998, Genome Res. 8: 175-185; Ewing and Green, 1998, Genome Res. 8:685-734). Reads were assembled using PHRAP (P. Green, Abstracts of DOE Human Genome Program Contractor-Grantee Workshop V, Jan. 1996, p 157) with default program parameters and quality scores.

To identify *Alloiococcus otitidis* genome encoded polypeptides, the complete genomic sequence of *Alloiococcus otitidis* was analyzed essentially as follows: First, all possible stop-to-stop open reading frames (ORFs)  $\geq$  222 nucleotides in all three reading frames were translated into amino acid sequences.

Second, the identified ORFs were analyzed for homology to known protein sequences. Third, the coding potential of non-homologous sequences were evaluated with the GENEMARKTM software program (Borodovsky and McIninch, 1993, Comp. Chem. 17:123). The results of these analysis are set forth in tables 2-16.

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#### EXAMPLE 5

#### IDENTIFICATION OF SPECIFIC GENES IN ALLOIOCOCCUS OTITIDIS

Alloiococcus otitidis homologs of the genes listed in Table 4 were identified as follows:

Protein sequences of interest ("query sequences", Table 4) were extracted from Genbank from one or more species; query species included but were not limited to Staphylococcus aureus, Streptococcus pnuemoniae, Streptococcus pyogenes, Lactococcus lactis, Escherichia coli, and Bacillus subtilis. These queries were compared to the Alloiococcus otitidis sequence by several methods in order to determine which Alloiococcus sequence was the ortholog for the query gene.

First, the query sequences were compared to the translated *Alloiococcus* otitidis ORF set using BLASTP. The ORF set was generated as described in Vaccines patent, except that for each ORF that had multiple potential start codons, only the longest ORF was used. The top 10 *Alloiococcus otitidis* hits for each query were saved, without regard to score.

These Alloiococcus otitidis hits were then compared to NR, the nonredundant Genpept database, using BLASTP. An Alloiococcus otitidis ORF was considered the ortholog of a query sequence if the genes were reciprocal best hits in Alloiococcus

otitidis and the query genome. This analysis is also sumarized in Table 4 (excel file AOT\_PATENT\_FILE.xls, Sheet TopHitsAndClustalKey). Specific numerical cutoffs were not used; however all top hits had Expect values of less than 3 x 10<sup>-28</sup>.

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Several query sequences had more than one high-scoring hit in *Alloiococcus* otitidis. In most cases, however, only the first, best hit to the original query sequence had that query sequence as its reciprocal best hit. For example, the *Streptococcus* pyogenes query sequence GyrA (alpha subunit of DNA gyrase) has two high-scoring hits in *Alloiococcus otitidis*. These were distinguished by the reciprocal blast analysis; the first, ORF\_505 (60% identity, Expect = 0) is the GyrA homolog and the second, ORF\_1907 (38% identity, Expect = 1 x 10 <sup>-154</sup>) is the homolog of the query sequence GrlA or ParC (topoisomerase IV, A subunit). Other examples of closely related proteins include the B subunits of DNA gyrase (GyrB) and Topoisomerase IV (GrlB or ParE); and YphC and Era, both of which are putative GTP binding proteins of unknown function. These *Alloiococcus otitidis* ORFS were assigned based on their top hit in Genpept.

In two cases the multiple high-scoring hits in *Alloiococcus otitidis* were the result of gene duplication. In the case of MurA (UDP-N-acetylglucosamine enolpyruvyl transferase) two separate *Alloiococcus otitidis* ORFS were determined to be the desired orthologs, because both had MurA (or MurZ, alternate notation) as their best hit in Genpept. Likewise, there are two FolC (folylpolyglutamate synthase) homologs in *Alloiococcus otitidis*. It is known that other bacteria, particularly Grampositive bacteria, may carry two homologs of each of these genes.

As a further step in verification of gene assignments, the *Alloiococcus otitidis* ORFS identified as orthologs of the query genes by the analysis above were then compared to an internal copy of the COGS database (Tatusov RL, Natale DA, Garkavtsev IV, Tatusova TA, Shankavaram UT, Rao BS, Kiryutin B, Galperin MY, Fedorova ND, Koonin EV, 2001, Nucleic Acids Res 2001 Jan 1;29(1):22-8. The COG database: new developments in phylogenetic classification of proteins from complete genomes) using BLASTP. The COGS database is a curated set of proteins from a set of finished bacterial genomes, which have been grouped into specific protein families on the basis of protein similarity. In all cases, the *Alloiococcus otitidis* ORF was most closely related to the COGS family of the initial query protein, if that protein had been assigned to a COGS family. Examples of proteins for which

there is no COGS family defined (in our local version of the database) include SrtA (sortase) and MvaK1 (phosphomevalonate kinase).

As a final confirmation, all query proteins were compared to the complete Alloiococcus otitidis nucleotide sequence using TBLASTN, in order to determine if there were additional and/or better hits that had not been predicted as ORFS. In all cases, the same sequence was identified as the best hit by TBLASTN and by BLASTP.

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For one query sequence, sortase, the Alloiococcus otitidis ORF that was the top hit (Expect = 0.42) by the initial BLASTP or TBLASTN using the Staphylococcus aureus sortase sequence as query was found by additional analysis (reciprocal blast) to be a putative ABC-transport protein. The true sortase homolog in Alloiococcus otitidis was identified by construction of a Hidden Markov Model based on a multiple alignment of 72 known and putative sortase proteins that had been identified previously using similar computational methods. The model was constructed using "hmmbuild" and the Alloiococcus otitidis ORF set was searched using "hmmsearch", both of the hmmer package (S.R. Eddy. Profile hidden Markov models. Bioinformatics 14:755-763, 1998). The assignment of ORF\_876 as sortase was then confirmed by reciprocal blast as described above and in Table 2. ORF\_876 was also found to be the top hit in Alloiococcus otitidis when the Bacillus subtilis putative sortase (YhcS) was used as the query sequence in a BLASTP search. The Bacillus halodurans BH3596 Bacillus subtilis YhcS and proteins that are the top hits for RF\_876 have recently been placed into a COGS group of sortases, further confirming the identity of ORF\_876 as the Alloiococcus otitidis sortase.

TABLE 4

TABLE 7										
ORF	DNA	ORF	Protein	ORF	Protein	·				
NO.	SEQ ID NO.r	Start	Start	Stop	SEQ ID No.	Gene				
46b	Seq. ID No. 1	26225	26153	25800	Seq. ID No. 2	ACPS				
48c	Sea. ID No. 3	29105	29090	27696	Seq. ID No. 4	murF				
57b	Seq. ID No. 5	33738	33732	32455	Seq. ID No. 6	murA-2				
65c	Seq. ID No. 7	37245	37242	36634	Seq. ID No. 8	rpoE				
172	Seq. ID No. 9	88726	88726	87785	Seq. ID No. 10	гроА				
228d	Seq. ID No. 11	111563	111542	107883	Seq. ID No. 12	rpoC				
236c	Seq. ID No. 13	115224	115221	111643	Seq. ID No. 14	гроВ				
495c	Seq. ID No. 15	247355	247331	245949	Seq. ID No. 16	dnaB/C				
505b	Seq. ID No. 17	254277	254268	251794	Seq. ID No. 18	gyrA				
509	Seq. ID No. 19	256252	256246	254297	Seq. ID No. 20	gyrB				
515c	Seq. ID No. 21	259131	259116	257914	Seq. ID No. 22	dnaN				
528b	Seq. ID No. 23	263837	263861	265153	Seq. ID No. 24	folC-2				
851b	Seq. ID No. 25	440982	441072	442634	Seq. ID No. 26	murE				
876	Seq. ID No. 27	453874	453898	454509	Seq. ID No. 28	srtA				
956	Seq. ID No. 29	500019	500019	501308	Seq. ID No. 30	folC-1				
959	Seq. ID No. 31	501978	501993	502364	Seq. ID No. 32	folB				
961c	Seq. ID No. 33	502392	502413	502943	Seq. ID No. 34	folK				
1183b	Seq. ID No. 35	626391	626430	627632	Seq. ID No. 36	mvaS				
1184	Seq. ID No. 37	. 629315	629285	627993	Seq. ID No. 38	mvaA				
1263	Seq. ID No. 39	675596	675608	676525	Seq. ID No. 40	murB				
1273	Seq. ID No. 41	685392	685377	684289	Seq. ID No. 42	mvaK2				
1275b	Seq. ID No. 43	686415	686403	685393	Seq. ID No. 44	mvaD				
1277	Seq. ID No. 45	687376	687349	686396	Seq. ID No. 46	mvaK1				
1279	Seq. ID No. 47	687461	687506	688435	Seq. ID No. 48	coaA				
1284	Seq. ID No. 49	691675	691681	692520	Seq. ID No. 50	nadE				
1511	Seq. ID No. 51	815078	815084	815920	Seq. ID No. 52	murl				
1811b	Seq. ID No. 53	985498	985504	986454	Seq. ID No. 54	folP				
1863b	Seq. ID No. 55	1019023	1019050	1019583	Seq. ID No. 56	folA				
1902	Seq. ID No. 57	1040639	1040645	1042606	Seq. ID No. 58	GrlB .				
1907	Seq. ID No. 59	1042729	1042732	1045191	Seq. ID No. 60	grlA				

PCT/US02/36122

Table 4 (Cont'd.)

ORF	DNA	ORF	Protein	ORF	Protein	_
NO.	SEQ ID NO.r	Start	Start	Stop	SEQ ID No.	Gene
1990c	Seq. ID No. 61	1098801	1098798	1097689	Seq. ID No. 62	rpoD
1992b	Seq. ID No. 63	1100670	1100670	1098817	Seq. ID No. 64	dnaG
2003	Seq. ID No. 65	1109198	1109144	1108212	Seq. ID No. 66	era
2016h	Seq. ID No. 67	1115435	1115390	1113879	Seq. ID No. 68	norA
2133	Seq. ID No. 69	1179995	1179938	1175604	Seq. ID No. 70	polC
2181b	Seq. ID No. 71	1203606	1203588	1202281	Seq. ID No. 72	obg
2204	Seq. ID No. 73	1216828	1216804	1215491	Seq. ID No. 74	yphC
2240c	Seq. ID No. 75	1236616	1236607	1233293	Seq. ID No. 76	dnaE
2284	Seq. ID No. 77	1261069	1261063	1259858	Seq. ID No. 78	coaBC
2328	Seg. ID No. 79	1286689	1286668	1285637	Seq. ID No. 80	holA
2333	Seq. ID No. 81	1290847	1290847	1290371	Seq. ID No. 82	coaD ·
2485	Seq. ID No. 83	1374427	1374400	1373168	Seq. ID No. 84	ftsZ
2489	Seq. ID No. 85	1375804	1375792	1374428	Seq. ID No. 86	ftsA
2492b	Seq. ID No. 87	1378075	1378060	1376897	Seq. ID No. 88	murG
2494	Seq. ID No. 89	1379477	1379453	1378050	Seq. ID No. 90	murD
2514	Seq. ID No. 91	1390141	1390135	1389491	Seq. ID No. 92	nadD
2596	Seq. ID No. 93	1437374	1437374	1436709	Seq. ID No. 94	coaE
2602	Seq. ID No. 95	1442399	1442396	1441065	Seq. ID No. 96	murC
2645	Seq. ID No. 97	1467800	1467782	1466751	Seq. ID No. 9	fmhB
2875	Seq. ID No. 99	1605944	1605923	1603701	Seq. ID No. 100	pcrA
2918	Seq. ID No. 101	1631092	1631089	1629779	Seq. ID No. 102	murA-1
3001	Seq. ID No. 103	1680254	1680221	1679229	Seq. ID No. 104	holB
3012	Seq. ID No. 105	1684114	1684102	1682330	Seq. ID No. 106	dnaX

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# EXAMPLE 6 IDENTIFICATION OF THE GENE ENCODING COENZYME A (COA) IN ALLOIOCOCCUS OTITIDIS

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Pantothenate kinase (PanK, CoaA) encoded by the *coaA* gene catalyzes the initial step in Coenzyme A (CoA) biosynthesis. CoA is an essential co-factor in a number of metabolic pathways in bacteria and mammals. Short-chain thioesters such as acetyl-CoA and succinyl-CoA are essential intermediates in carbon metabolism. CoA-thioesters of long chain fatty acids feed into β-oxidation and are also the source of fatty acids for phospholipids. In addition, CoA and its thioesters play important roles in the regulation of several enzymes in intermediary metabolism, including pyruvate dehydrogenase and phosphoenolpyruvate carboxylase. Finally,

synthesis of holo acyl carrier protein (ACP) is dependent on CoA for the 4'-phosphopantetheine moiety linked to ACP. ACP is essential for fatty acid biosynthesis. The two major acyl-carrier groups in cells: CoA and ACP, are derived from pantothenate. Pantothenate can be obtained exogenously through uptake via a permease, the product of the panF gene. Alternately, pantothenate is the product of condensation of pantoate and  $\beta$ -alanine via pantothenate synthetase, the product of the panC gene. The initial step in CoA biosynthesis is the phosphorylation of pantothenate by pantothenate kinase (PanK, CoaA).

The coaA gene was originally identified by Dunn and Snell in S. typhimurium as a temperature sensitive allele. Similarly, a temperature sensitive allele of coaA was reported for E. coli in 1987. CoaA was found to be essential in E. coli in a recent genetic footprinting analysis. In the temperature sensitive strains, accumulation of phosphorylated CoA intermediates rapidly ceased following shift to the non-permissive temperature. CoaA was shown to be a homo-dimer of 35 kDa subunits that bound ATP cooperatively. ATP is bound first in a sequential mechanism of action; CoA has been shown to be a potent inhibitor of the reaction and competitively competes for binding with ATP. Therefore CoaA is under feedback regulation and is the major regulatory step in CoA biosynthesis.

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Lysine 101 in bacterial pantothenate kinase (CoaA) was found to be essential for both ATP and CoA binding. This supports kinetic data that CoA is a competitive inhibitor of ATP binding to CoaA and that both substrates bind to the same site.

Homologues of *E. coli* CoaA have been identified in *B. subtilis, S. pyogenes, M. tuberculosis, H. influenzae* and *V. cholerae*. Homologues have not been identified in either the *S. cerevisiae* genome or in a mammalian expressed sequence tag database. Calder *et al.* identified a homologue, through functional complementation of an *E. coli coaA* ts mutant, in *A. nidulans*. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 47. The protein encoded by the gene is set forth in Seq. ID No. 48.

The A. nidulans gene was then used to identify a yeast homologue. The bacterial and Aspergillus enzymes were found to be 16% identical and 32% similar. Although this level of similarity is quite weak the essential lysine residue involved in nucleotide binding appears to be conserved; however, the sequence surrounding the lysine residue were not conserved and further study will be required to validate this

finding. The most striking difference between the eukaryotic and prokaryotic enzymes is found in the sensitivity of each to competitive inhibition by CoA and acetyl-CoA. The yeast enzyme was most sensitive to acetyl-CoA and less sensitive to CoA, whereas the converse was true for the bacterial enzyme. Later studies demonstrated that mammalian pantothenate kinase is activated by CoA and inhibited by acetyl-CoA.

#### **Nucleotide binding**

Binding of ATP to CoaA is directly demonstrated by equilibrium dialysis employing the non-hydrolyzable ATP analogue ATP $\gamma$ S. The  $K_d$  measured for ATP binding is reported to be 2.1  $\mu$ M.

#### **CoA** binding

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Binding of CoA to CoaA is directyl demonstrated by equilibrium dialysis and the  $K_d$  is reported to be 6.7  $\mu M$ .

#### Pantothenate kinase activity

Specific kinase activity of CoaA is demonstrated using D-[1- $^{14}$ C]pantothenate and capturing 4'-phospho[1- $^{14}$ C]pantothenate on DE81 filters. Using this assay the following kinetic values were derived: specific activity – 470+/- 200 nmol/min/mg; pantothenate  $K_m$  – 36  $\mu$ M;  $K_m$  ATP – 136  $\mu$ M.

#### Suitability of target for anti-infective development

Coenzyme A biosynthesis is essential for bacterial viability. CoaA catalyzes the first step of biosynthesis of CoA and appears to be the point of regulation for the pathway. The essentiality of CoaA is demonstrated through the construction of temperature sensitive alleles in *coaA*. Although the yeast enzyme is found to functionally complement the bacterial temperature sensitive allele, sequence and kinetic differences suggest the possibility of identifying inhibitors of the bacterial enzyme with high selectivity. As CoaA is essential and conserved in gram-negative and gram-positive pathogens, such inhibitors will have broad-spectrum utility.

#### Suitable assays for measuring CoaA function

CoaA is purified by standard methods using widely available molecular tags following expression at high level from *E. coli*. Pantothenate kinase activity is measured as follows: CoaA and D-[1-<sup>14</sup>C]pantothenate is incubated in a buffer consisting of 100 mM Tris (pH 7.4), 2.5 mM MgCl<sub>2</sub>, 2.5 mM ATP for 5-60 minutes at 37°C. Product, 4'-phospho[1-<sup>14</sup>C] pantothenate, is monitored through retention of labeled material on DE81 filters. This assay is amenable to high-throughput screening using high-density well-filter plates.

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#### EXAMPLE 7

#### IDENTIFICATION OF THE GENE ENCODING COABC (DFP) IN ALLOIOCOCCUS OTITIDIS

The *E. coli dfp* gene, which encodes the previously designated Dfp protein, was originally identified as encoding an enzyme required for CoA biosynthesis. The gene, coding for the protein of interest, was renamed *coaBC* to reflect the enzyme function in CoA biosynthesis. CoA is an essential co-factor in a number of metabolic pathways in bacteria and mammals. Short-chain thioesters such as acetyl-CoA and succinyl-CoA are essential intermediates in carbon metabolism.

CoaBC carries out the second and third steps of coenzyme A biosynthesis: the conjugation of 4'-phosphopantetheate with cysteine by the CoaB (PPCS: 4'phosphopantethenoyl cysteine synthase) activity followed by the conversion to 4'-phosphopantetheine by the CoaC (PPCDC: 4'phosphopantenoylcysteine decarboxylase) activity. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 77). The protein encoded by the gene is set forth in Seq. ID No. 78.

#### Enzyme activity of CoaBC (Dfp):

Initially it was demonstrated that Dfp enzyme catalyzing oxidative decarboxylation of (R)-4'-phospho-N-pantothenoylcysteine (*P*PC) to form 4'-phosphopantetheine (*P*P) — the third step in CoA biosynthesis from pantothenate The  $K_M$  for this reaction is 800  $\mu$ M for *P*PC.

Subsequently, it was established that Dfp is a bifunctional enzyme, catalyzing the second step of CoA biosynthesis, coupling of 4'-phosphopantothenate with

cysteine to form *PPC*, as well. This reaction is a two-step process and requires CTP for initial 4'-phosphopantothenate activation. Second step couples cysteine to the phosphopantothenate moiety with a release of CMP. Estimated  $K_{\rm M}$ 's are 300  $\mu{\rm M}$  for 4'-phosphopantothenate and CTP, and 250  $\mu{\rm M}$  for cysteine.

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#### CoaBC as target for antibacterial development.

Coenzyme A (CoA) plays a vital role in the metabolism of living cells. According to a recent report, 4% of all enzymes in the cell require CoA, its thioesters or 4'-phosphopantetheine. Recent genetic footprinting experiments on *E. coli* and direct gene knockout have established that this *coaBC* is essential for bacterial growth. Homologs of *coaBC* have been identified in a number of gram-positive and gram-negative organisms, which suggested the possibility of developing a broad-spectrum antibacterials from *coaBC* inhibitors. Considering the bifunctional nature of CoaBC, it is feasible to identify inhibitors that will inhibit both enzymic functions, thus arresting two steps in the CoA pathway. Another important factor in favor of selecting CoaBC as a target for antibacterials is low homology of the bacterial enzyme to eukaryotic counterparts. In most of the higher organisms including humans, two separate enzymes carry out these functions. Moreover, mammalian (R)-4'-phospho-N-pantothenoylcysteine decarboxylase is a pyruvate-dependent enzyme, while CoaBC requires flavine mononucleotide for its function.

#### Assays for measuring CoaBC activity.

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*P*PC synthetase activity is be monitored by detecting the released pyrophosphate. This is achieved by converting pyrophosphate to inorganic phospate with pyrophosphatase and detection by the Malachite Green assay, or by the MESG assay spectrophotometrically. CoaBC (2 μg) is incubated in the reaction buffer containing 10 mM DTT, 2 mM MgCl<sub>2</sub>, 50 mM Tris-HCl, pH 8, 300 μM 4'-phosphopantothenate, 3.5 mM CTP, 5 μg pyrophosphatase. The reaction is started by addition of appropriate amount (10-500 μM final) of cysteine. The reaction is stopped at different time points by addition of equal volume of 5M  $\rm H_2SO_4$ . The

amount of inorganic phosphate released will be determined according to the one of described techniques.

PPC synthetase activity is also monitored by detecting the release of carbon dioxide from <sup>14</sup>C-labeled cysteine. CoaBC (2 μg) is incubated in the reaction buffer containing 10 mM DTT, 2 mM MgCl<sub>2</sub>, 50 mM Tris-HCl, pH 8, 2.5 μM 4'-phosphopantothenate, 3.5 mM CTP. The reaction is started by addition of appropriate amount (30 mM, final concentration) of <sup>14</sup>C-labeled cysteine. The reaction is stopped at different time points by addition of equal volume of 5M H<sub>2</sub>SO<sub>4</sub>. Amount of released <sup>14</sup>C-labeled CO<sub>2</sub> is determined according to published technique.

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#### Example 8

### Identification of the gene encoding phosphopantetheine adenylyltransferase (CoaD) in Alloiococcus otitidis

Phosphopantetheine adenylyltransferase, (PPAT, CoaD, KdtB) catalyzes the penultimate step in Coenzyme A (CoA) biosynthesis. The fourth step in CoA biosynthesis is the addition of AMP to 4'-phosphopantetheine by phosphopantetheine adenylyltransferase (CoaD) to form 3' dephospho-CoA (dPCoA).

The coaD gene was first identified in E. coli by Geerlof et al. CoaD is essential for viability in E. coli and S. aureus. The enzyme has a mass of 18 kDa and was determined to be a hexamer through cross-linking studies. Crystallography confirmed the oligomeric state of the enzyme. Moreover, co-crystallography of CoaD with dPCoA has also been carried out mapping the binding pocket for the major product of the reaction. Interestingly, in mammals PPAT has been shown to be in a complex with dephospho Coenzyme A kinase (dPCoA kinase, DPCK). This enzyme, purified from pig liver, is referred to as CoA Synthase. The yeast PPAT is associated with a protein complex that is in excess of 375 kDa and composed of six proteins. There is no detectable homology between the bacterial PPAT (CoaD) and the recently identified human PPAT, the activity of which is contained in a bifunctional PPAT/DPCK enzyme. Homologues of E. coli CoaD have been identified in P. aeruginosa, S. pneumoniae, S. aureus, H. influenzae, H. pylori, B. anthracis and M. tuberculosis. Homologue of this gene identified in Alloiococcus otitidis as described in

Example 5 (Seq. ID No 81). The protein encoded by the gene is set forth in Seq. ID No. 82.

#### Enzyme activity

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CoaD (PPAT) carries out the reversible transfer of AMP to 4'-phosphopantetheine, forming dephosphocoenzyme A and releasing PPi. The reverse reaction was demonstrated by Geerlof *et al.* using a coupled assay to tie ATP production to NADP reduction, which is monitored at 340 nm. The following kinetic constants were calculated:  $k_{cat} = 3.3 + -0.1 / sec$ ;  $K_{m(dPCoA)} = 7.0 + 1.4 uM$ ;  $K_{m(PPi)} = 0.22 + 0.04 mM$ .

#### CoaD as target for anti-infective development.

Coenzyme A biosynthesis is essential for bacterial viability. CoaD, phosphopantetheine adenylyltransferase, catalyzes the fourth step in the pathway and was shown to be essential in both *E. coli* and *S. aureus*. There is no measurable homology between CoaD and the human PPAT enzyme, so the liability of poorly selective compounds is quite low. As CoaD is essential and conserved in gramnegative and gram-positive pathogens, inhibitors developed against this target will have broad-spectrum utility.

#### Assays for measuring CoaD function

CoaD will be expressed and purified using standard methodologies for bacterial expression and affinity tag-based purification. Two assay formats can be used to monitor enzymatic activity: the forward reaction and the reverse reaction.

The forward reaction assay was initially described for measuring the activity of the human PPAT activity in the PPAT/DPCK enzyme. The enzyme assay is carried out in 50 mM Tris (pH 8.0), 2 mM MgCl<sub>2</sub>, 5 mM ATP, 5-500 uM 4'-phosphopantotheine, 7.5 mM NADH and enzyme (initially 0.1 – 1.0 µg/ml). The production of PP<sub>i</sub> is detected using the protocol of O'Brien in which PP<sub>i</sub> production is coupled to the oxidation of NADH to NAD. This system requires the addition of 4 enzymes (PP<sub>i</sub>-dependent phosphofructokinase, aldolase, triosephophate isomerase and glycerol-3-P dehydrogenase) to the basic reaction mix and presents the added issue of deconvolution, which limits the use of the assay as a primary screen.

The reverse direction assay is carried out also as a coupled assay to tie ATP production to NADP reduction following the method described by Lamprecht & Trautschold. The assay is set up in reaction buffer containing the following: 50 mM Tris (pH 8.0), 1 mM DTT, 2 mM MgCl<sub>2</sub>, 1 mM NADP, 5 mM glucose, 2 mM PP<sub>i</sub>, 0.1 mM dPCoA. Hexokinase (4 units) and glucose-6-phosphate dehydrogenase (1 unit) will be added to the assay as the coupling enzymes in addition to CoaD (initially 0.1 – 1  $\mu$ g/ml). The assay is monitored at 340 nm. Deconvolution of hits is required with this assay, however with only 2 additional enzymes the task will be less cumbersome when compared to the forward assay described above.

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#### Example 9

### IDENTIFICATION OF THE GENE ENCODING DEPHOSPHOCOA KINASE (DPCK, YACE, COAE) IN ALLOIOCOCCUS OTITIDIS

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DephosphoCoA kinase (DPCK, YacE, CoaE) encoded by the coaE gene catalyzes the final step in Coenzyme A (CoA) biosynthesis. The final step in CoA biosynthesis is the phosphorylation of the 3'-hydroxyl group of dephospho-CoA to form CoA by dephosphocoenzyme A kinase (DPCK, YacE, CoaE).

The determination that the previously identified *yacE* gene encoded the dephosphocoenzyme A kinase activity was reported by Mishra *et al.* These authors previously determined that separate enzymes encode the phosphopantetheine adenyltransferase (PPAT) and dephosphocoenzyme A kinase (DPCK) activity in *Corynebacterium ammoniagenes* in contrast to the eukaryotic enzymes in which the PPAT and DPCK activities are coupled. The *E. coli* gene, encoding a 25 kDa protein, was cloned based on the sequence of the *C. ammoniagenes* gene and found to be identical to the previously described *yacE* gene. The gene was designated *coaE* to follow existing nomenclature in *E. coli. CoaE* (*YacE*) was shown to be essential in *E. coli* through genetic footprinting. CoaE is widely distributed in bacteria. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 93). The protein encoded by the gene is set forth in Seq. ID No. 94.

#### Assays for measuring CoaE function

CoaE carries out the phosphorylation of dephosphocoenzyme A at the 3' hydroxyl group, consuming ATP, to form CoA. Dephosphocoenzyme A kinase activity is measured in a coupled reaction in which NADH oxidation to NAD is tied to ADP production. In this assay, the standard pyruvate kinase/lactose dehydrogenase coupling system is used to generate NAD in a 1:1 molar equivalent to the ADP produced by the test enzyme. NADH oxidation to NAD is monitored at 340 nm in a standard spectrophotometer. The following kinetic values were determined for CoaE:  $K_{m(ATP)} = 0.74 \text{ mM}$ ;  $K_{m (dephospho-CoA)} = 0.14 \text{ mM}$  (7).

The formation of CoA is monitored using a coupled enzyme system in which acetyl-CoA is formed in proportion to the amount of CoA in the assay. Three enzymes (phosphate acetyl transferase, citrate synthase and malate dehydrogenase) are added to the reaction that results in the formation of NADH from NAD, which is monitored at 340 nm.

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#### CoaE as a target for anti-infective development

Coenzyme A biosynthesis is essential for bacterial viability. CoaE, dephosphocoenzyme A kinase, catalyzes the final step in CoA synthesis and is shown to be essential by genetic footprinting in *E. coli*. A degree of homology between CoaE and the human DPCK enzyme has been noted, such that selectivity assays is necessary to determine a high therapeutic index for CoaE inhibitory compounds. CoaE is conserved in gram-negative and gram-positive pathogens and should have broad-spectrum utility in the clinic.

CoaE is expressed and purified using standard methodologies for bacterial expression and affinity tag-based purification. DephosphocoA kinase activity is monitored using a coupled enzyme system to tie ADP production to oxidation of NADH to NAD. The decay of absorbance at 340 nm will be the assay readout. The assay will be setup in the following buffer: 50 mM Tris (pH 8.5), 20 mM KCl, 10 mM MgCl<sub>2</sub>, 10 mM ATP, 0.3 mM NADH and 0.4 mM phosphoenolpyruvate. The coupling enzymes: pyruvate kinase (10 U) and lactate dehydrogenase (4 U) will be added along with dephosphocoenzyme A kinase (initially 0.1- 1.0 ug/ml). The assay will be started by the addition of 0.4 mM dephosphocoenzyme A. In this assay system, the release of ADP is tied to the oxidation of NADH to NAD, and is monitored at 340 nm.

This assay is transferable to a high-density microtiter plate format and suitable for HTS.

#### **EXAMPLE 10**

### 5 IDENTIFICATION OF DNAB AND PCRA, GENES ENCODING HELICASES IN ALLOIOCOCCUS OTITIDIS

Helicases unwind double-stranded DNA in a reaction that couple nucleotide binding and hydrolysis to strand unwinding. Their activity is required for a number of biological processes such as separation of the chromosome during replication, recombination and repair. Homologue of these genes were identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 15 and 99). The protein encoded by the gene is set forth in Seq. ID No. 16 and 100.

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Due to the essential roles modulated by these molecules they represent an important target for antibacterial therapy. Homologs of *dnaB* and *pcrA* genes encoding helicases were identified as described in Example 5. A primary assay, which detects helicase function *in vitro*, is used to identify inhibitors of each enzyme and is described below.

Genes encoding DnaB and PcrA is obtained using polymerase chain reaction amplification of the genomic region encoding them. The genes is subcloned into a standard expression vector either containing an amino acid tag for ease of purification or not. The enzyme is then over-expressed in *Escherichia coli* and purified using a standard tag system.

Most helicases require a region of single-stranded DNA flanking the duplex region that it unwinds. As a result, providing a single stranded region to either the 3' or 5' end of a duplex allows for determination of the polarity of helicase unwinding. These types of experiments have demonstrated that PcrA and DnaB are 3'-5' and 5'-3' helicases, respectively. None the less, a convenient filtration assay has previously been described that is formatted for high-through-put screening of inhibitors of either enzyme, regardless of polarity. Assays (90 ul) contained 15 pM single-stranded M13 DNA to which a radiolabeled oligonucleotide had been annealed as a substrate for unwinding. Reactions are carried out in 96-well GF/C unifilter hydrophobic plates (Polyfiltronics Inc.) in 70 ul helicase buffer [20 mM Hepes (pH 7.6), 4 mM MgCl<sub>2</sub> 4

mM ATP, 100 ug/ml BSA, 5% glycerol and 2 mM DTT] and 10 ul of DMSO or compound. Reactions are initiated by adding 10 ul of purified helicase protein and are incubated for 1 hr at room temperature. 100 ul of 2X capture buffer containing silica beads [25% methanol, 3 M NaI, 0.03% NP-40, and 10% GlassFog beads (BIO101)] were added. The mixture was incubated for 30 min at room temperature. Plates are then washed 5X on a Bio-Teck instruments, Auto Washer EL403) with wash buffer (50% ethanol, 0.2% NP-40 and 50 mM NaCl). Scintillation fluid was added and plates are counted (Packard Topcount).

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#### **EXAMPLE 11**

### IDENTIFICATION OF DNAE, THE GENE ENCODING DNAE-POLYMERASE IN ALLOIOCOCCUS OTITIDIS

DnaE is an enzyme that catalyzes the DNA template directed polymerization of deoxyribonucleotides into deoxyribonucleic acid. The enzyme has been reported to modulate lagging strand synthesis at gram-positive replication forks. Functions for DnaE have been defined biochemically, in *Bacillus subtilis* and *Streptococcus pyogenes*. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 75). The protein encoded by the gene is set forth in Seq. ID No. 76.

Because DnaE is an essential protein in gram-positive bacteria and has high homology to the gram-negative *dnaE*, which is an essential polymerase subunit of the DNA polymerase III holoenzyme, it serves as a good target for antibacterial drug discovery. A primary assay, which detects processive DnaE mediated DNA synthesis *in vitro*, is useful identify inhibitors of the enzyme and is described below.

The gene encoding DnaE I in *Alloiococcus otitidis* was identified as described in Example 5. Purification of DnaE DNA polymerase from *Alloiococcus*. The gene encoding DnaE is obtained using polymerase chain reaction amplification of the *dnaE* gene. The gene is subcloned into a standard expression vector either containing an amino acid tag for ease of purification or not. The enzyme is then over-expressed in *Escherichia coli* and purified using a standard tag system.

Because DnaE catalyzes the incorporation of single deoxyribonucleotides into DNA, the incorporation of radiolabelled deoxyribonucleotides into larger deoxyribonucleic acid molecules is monitored to measure activity of the enzyme. A

filtration assay has been previously described for *Streptococcus pyogenes* DnaE that uses filterplates containing DE81 filters to capture polymerized DNA. This assay is amenable to high-through-put screening format for DnaE. Assays contained 70 ng of 30-mer primed M13mp18 single stranded DNA as a template for replication. The reaction contained 3.3-300 ng of DnaE in 23.5 µl of replication buffer [20 mM Tris-HCL (pH 7.5), 4% glycerol, 0.1 mM EDTA, 5 mM DTT, 2 mM ATP, 8 mM MgCl<sub>2</sub>, 40 µg/ml BSA] and 60 µM of both dGTP and dCTP. NaCl was added to the reaction mixture to a final concentration of 40 mM. DNA synthesis was initiated by the addition of 1.5 µl of 1.5 mM dATP and 0.5 mM [µ-<sup>32</sup>P]dTTP. Reactions were incubated at 37°C for various lengths of time and were quenched by adding an equal volume of 1% SDS and 40 mM EDTA. One-half of the terminated reaction was applied to DE81 filter paper and washed 3X with wash solution (0.3 M Ammonium formate and 0.01 M Sodium pyrophosphate). Filters were then placed in scintillation vials and 1 ml scintillation counter.

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#### EXAMPLE 12

### IDENTIFICATION OF DNAG, THE GENE ENCODING PRIMASE IN ALLOIOCOCCUS OTITIDIS

DnaG is an enzyme that catalyzes the DNA template directed polymerization of ribonucleotides into ribonucleic acid *de novo*. Ribonucleic acid molecules that are synthesized by DnaG primase subsequently serve as primers for synthesis of the leading- and lagging-strands during chromosomal replication. Functions for DnaG have been defined biochemically, and the crystal structure of the RNA polymerase domain has been determined in *Escherichia coli*. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 63). The protein encoded by the gene is set forth in Seq. ID No. 64.

Because DnaG primase plays an essential role in both leading- and laggingstrand synthesis during chromosomal replication, and DnaG has homologs in all prokaryotes but not eukaryotes, it serves as a good target for antibacterial drug discovery. A primary assay, which detects DnaG mediated RNA synthesis *in vitro*, can be used to identify inhibitors of the enzyme and is described below.

## Assay for the activity of DNA polymerase and identification of compounds that inhibit DnaG

The gene encoding DnaG is obtained using polymerase chain reaction amplification of the *dnaG* gene. The gene is subcloned into a standard expression vector either containing an amino acid tag for ease of purification or not. The enzyme is then over-expressed in *Escherichia coli* and purified using a standard tag system.

Because DnaG catalyzes the incorporation of single ribonucleotides into RNA, the incorporation of radiolabelled ribonucleotides into larger ribonucleic acid molecules is monitored to measure activity of the enzyme. A high-throughput scintillation proximity assay (SPA) assay, previously described for *E. coli* DnaG, is used to meadure activity of DnaG activity in a coupled reaction with DnaB helicase. The assay, which was shown to work with DnaG alone, is used to screen for compounds that inhibit DnaG function. Assays are run in 96-well Packard Optiplate plates. First, 1 μl DMSO or test compound was added, followed by 20 μl of DnaG (208 nM) and 3.3 nM M13mp18 single-stranded DNA. Reactions are initiated by adding 10 ul of primase assay buffer [50 mM Tris-HCI (pH 7.5), 4% sucrose, 8 mM DTT, 5 mM MgCl<sub>2</sub>, 40 ug/ml BSA, 0.1 μg/ul Rifampicin, 25 U/ml RNA guard, 100 μM GTP, 100 μM UTP, 3 μM CTP, 1 mM ATP] and 0.4 μCi [³H]CTP. Reactions are incubated at 30°C for 30 min. Next, a suspension of 50 μl of 2.5 mg/ml PVT-PEI SPA beads (Amersham; prepared in 0.3 M NaCitrate, pH 3.0) were added. Plates were read after 1 hr on a Topcount instrument (Packard).

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#### EXAMPLE 13

### DNAN, DNAX, HOLA, HOLB, AND POLC, THE GENES ENCODING THE SUBUNITS OF <u>ALLOIOCOCCUS OTITIDIS DNA POLYMERASE III HOLOENZYME: BETA (β), TAU (T), DELTA</u> (Δ), DELTA' (Δ') AND POLC.

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DNA polymerase III holoenzyme is an enzyme complex comprised of multiple highly conserved subunits that catalyzes the DNA template directed polymerization of deoxyribonucleotides into deoxyribonucleic acid. In gram positive organisms the holoenzyme is composed of a polymerase subunit, PoIC, and accessory proteins. The accessory proteins act in a coordinated manner to clamp the polymerase tightly to the DNA template allowing the polymerase to synthesize DNA with high speed and

processivity. Homologue of these genes identified in *Alloiococcus otitidis* are described in Example 5 (Seq. ID Nos. 21, 105, 79, 103, and 105 respectively). The protein encoded by the gene is set forth in Seq. ID No. 22, 106, 80, 104 and 106 respectively).

Functions for the individual subunits have been defined biochemically and interactions between them have now been deduced structurally by crystallographic analysis of the enzyme from *Escherichia coli*. Tau interacts directly with both delta and delta' to form a clamp loader complex. Upon binding ATP the complex undergoes a conformational change altering an interaction between delta and delta', which allows delta to subsequently interact with the beta-clamp. The beta-clamp is a ring-shaped homomultimer assembly that can be opened by delta and placed onto a primed DNA template. ATP hydrolysis results in closing the clamp around DNA and dissociation of the clamp-loading complex. PoIC then couples with the beta clamp to form a highly processive polymerase.

Because DNA polymerase III holoenzyme is comprised of multiple subunits, the opportunity exists to inhibit its activity at a number of different sites. A primary assay, which detects processive DNA synthesis *in vitro*, can be used to identify inhibitors of the enzyme and is described below. Deconvolution of inhibitors, based on either activity of physical interaction, follow the primary assay.

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#### Assay for the activity of DNA polymerase

Purification of DNA polymerase III holoenzyme subunits from *Alloiococcus*. Genes encoding the subunits of DNA polymerase is obtained using polymerase chain reaction (PCR) amplification of the genomic region encoding them. The genes are subcloned into a standard expression vector either containing an amino acid tag for ease of purification or not. The enzyme is then over-expressed in *Escherichia* coli and purified using a standard tag system.

Because DNA polymerase III catalyzes the incorporation of single deoxyribonucleotides into DNA, the incorporation of radiolabeled deoxynucleotides into larger deoxyribonucleic acid molecules is monitored to measure activity of the enzyme. A filtration assay is previously described for *Streptococcus pyogenes* DNA polymerase III that uses filterplates containing DE81 filters to capture polymerized DNA (2). This assay is amenable to high-through-put screening format. Assays

contained 70 ng of 30-mer primed M13mp18 single stranded DNA as a template for replication. The reaction contained 43 ng of ß and 140 ng of PolC-тдд' complex in 23.5 цl of replication buffer (20 mM Tris-HCL (pH 7.5), 4% glycerol, 0.1 mM EDTA, 5 mM DTT, 2 mM ATP, 8 mM MgCl<sub>2</sub>, 40 цg/ml BSA, and 60 цМ of both dGTP and dCTP. DNA synthesis was initiated by the addition of 1.5 цl of dATP and [цl-<sup>32</sup>P]dTTP. Reactions were incubated at 37°C for various lengths of time and were quenched by adding an equal volume of 1% SDS and 40 mM EDTA. One-half of the terminated reaction was applied to DE81 filter paper and washed 3X with wash solution (0.3 M Ammonium formate and 0.01 M Sodium pyrophosphate). Filters were then placed in scintillation vials and 1 ml scintillation counting liquid was added. Radioactivity was counted using a scintillation counter.

Compounds inhibiting PolC subunit is identified by modifying the above reaction to include only the PolC subunit and using 2.5 µg activated calf thymus DNA as a substrate, instead of singly-primed M13mp18 DNA, as previously described. Several techniques are utilized to determine the interaction of inhibitors with individual subunits. These have been described in the literature and include the following: (1) Nuclear magnetic resonance and capillary electrophoresis.

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### EXAMPLE 14 ERA GTPASE IN ALLOIOCOCCUS OTITIDIS

The era (E. coli Ras) gene was initially identified while sequencing around the mc gene; era lies downstream of mc. While a function for era has yet to be determined, conditional (temperature sensitive) mutants revealed that the product of the era gene, Era, is essential for E. coli viability. A hint as to an in vivo function for Era was uncovered when a suppressor of a dnaG (primase) allele was found to map in the era coding sequence and a second suppressor, which mapped upstream of the era open reading frame, affected expression of era. These data suggest that Era could play one or more roles in DNA replication, regulation of primase activity or otherwise effect cell cycle progression. More recent data has confirmed that the era1 mutant causes a defect in cell growth at the two-cell stage and delays cell division Moreover, Britton et al demonstrated that cell division was coupled with the level of

Era in the cell: division arrest, through reduction in Era levels, is reversed when Era levels return to threshold amount. A current model suggests that Era acts as a checkpoint regulator in the bacterial cell cycle. Era is a GTP-binding protein with GTPase activity, a threshold level of functional/activated Era may be required to initiate septation.

Era is associated with additional cellular functions, specifically translation, as Era specifically interacts with the translation machinery. *E. coli* Era binds both 16S rRNA and the 30S ribosomal subunit; whereas, the *S. pneumoniae* 16S rRNA copurifies with Era. A putative RNA binding "KH motif" has been identified in the carboxyl-terminal domain. The RNA binding activity is critical to Era cellular function as mutation of the putative RNA binding region of the *S. pneumoniae* Era prevents complementation of an *E. coli era* mutant strain. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID No 65). The protein encoded by the gene is set forth in Seq. ID No. 66.

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#### **Nucleotide binding**

Filter-binding assays are utilized to demonstrate nucleotide-binding specific to GTP and not UTP, CTP or ATP. Both GTP and GDP (unlabeled) were capable of inhibiting  $\alpha^{32}$ P-GTP binding. The Kd for GTP and GDP binding were reported to be 5.5 and 1.0  $\mu$ M, respectively.

A large number of GTP-binding proteins have been studied and all members of the family contain three regions of highly homologous amino acid residues that define a GTP-binding pocket. Era contains well-conserved regions defining the so-called G1 (G/AXXXXGKT/S: residues 15-22), G3 (DXXG: residues 62-65) and G4 (NKXD: residues 124-128) consensus sequences. The G2 domain (residues 33-38, see below), located between G1 and G3, is generally more variable.

#### **GTPase activity**

Purified Era showed a significant GTPase activity, which is inhibitable by GTP or GDP but not by UTP, CTP, ATP or ADP. The maximum hydrolysis rate is measured at 9.8 mmol GTP hydrolyzed/min/mol Era. The Km was found to be 9 μM.

It should be noted that Sullivan et al demonstrated, using mant (N-methyl-3'-O-anthraniloyl) labeled GTP and GDP, very rapid exchange kinetics for guanine

nucleotide binding. Era exchanges guanine nucleotides 10-fold more rapidly than the GTP hydrolysis rate suggesting that guanine nucleotide binding and release should be considered as a regulatory point in addition to the more well-studied hydrolysis step.

Autophosphorylation

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When  $\gamma^{32}$ P-GTP is used as a substrate for the GTPase activity , Era is phosphorylated. The autophosphorylation reaction is specific for GTP, as incubation with  $\gamma^{32}$ P-ATP did not result in phosphorylation of Era. Moreover,  $\alpha^{32}$ P-GTP is not a suitable substrate for detection of Era autophosphorylation. Tryptic digestion and HPLC were utilized to resolve the sites(s) of phosphorylation. Using  $\gamma^{32}$ P-GTP as a substrate the major radioactive peak contained the tryptic peptide, ISITSR, corresponding to Era residues 33-38 and containing 3 potential phosphorylation sites. Mutagenesis of both Thr-36 and Ser-37 to alanine abolished enzymatic activity. However, individual alanine substitutions at either site had no effect on Era function. The autophosphorylation site is located in the so-called G2 domain of Era.

#### Suitability of target for anti-infective development

Era is an essential protein for bacterial viability. Knock-down mutations as well as conditional-lethal alleles revealed that Era function is required for cytokinesis. An additional phenotype of the Era-depleted strains is an aberrant response to temperature induced stress. This target is novel and may well lead to the identification of new classes of anti-infectives. The widespread distribution of Era homologues in both gram-negative and gram-positive pathogens suggests that broad-spectrum agents could result from an effort to define Era inhibitory compounds.

#### Assays for measuring Era function

#### 30 NUCLEOTIDE BINDING ASSAYS

Era binding to nucleotide is monitored by a simple filter-binding assay. Era (1-5  $\mu$ g) is incubated with  $\alpha^{32}$ P-GTP (0.2  $\mu$ Ci) in a buffer consisting of 100 mM Tris (pH 7.5), 10 mM MgCl<sub>2</sub>, 0.2% NP-40, 0.2 mg/ml BSA for 30 minutes at 32°C. A

portion of the reaction mix is spotted on nitrocellulose membrane, washed (50 mM Tris (pH 7.5), 5 mM MgCl<sub>2</sub>, 1 mM DTT) and dried. The membrane is then exposed to X-ray film. Alternatively, the spots are excised and counted. This assay is directly amenable to HTS using filter plates.

GTPASE ACTIVITY ASSAY

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The GTP hydrolytic activity of Era is monitored using thin-layer chromatography. Era and α<sup>32</sup>P-GTP is incubated in 50 mM Tris (pH 7.5), 5 mM MgCl2, 0.1% NP-40, 0.2 mg/ml BSA for 30 minutes at 37°C. An aliquot of the reaction is placed on PEI cellulose and the strip developed with 0.5 M KH<sub>2</sub>PO<sub>4</sub>, 1.0 M NaCl (pH 3.7). The spots conforming to GDP and GTP are identified by UV shadowing, excised and counted. This assay represents an acceptable secondary/confirmatory assay.

Alternatively, the hydrolysis of  $\gamma^{32}P$ -GTP is monitored by assaying for liberated P<sub>i</sub>. Obg and  $\alpha^{32}P$ -GTP is incubated in 50 mM Tris (pH 8.5), 1.5 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 100 mM KCl, 10% glycerol for 30 minutes to 3 hours at 37°C. The reaction will be stopped by the addition of a slurry of charcoal in 1 mM Kpi (pH 7.5), which selectively binds the GTP and GDP. The liberated P<sub>i</sub> in the supernatant is monitored by Cerenkov counting. Free P<sub>i</sub> can also be monitored with the Malachite Green reagent.

#### **AUTOPHOSPHORYLATION ASSAY**

Era autophosphorylation is monitored by incubating Era with  $\gamma^{32}$ P-GTP in 50 mM morpholinopropane sulphate (pH 6.8), 5 mM MgCl2, 1 mM DTT at 37°C (14). Samples are analyzed following separation on SDS polyacrylamide gels, drying the gel and exposure to film. This assay represents an acceptable secondary/confirmatory assay for Era activity.

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## EXAMPLE 15 FMHB(FEMX) GENES IN ALLOIOCOCCUS OTITIDIS

The femA, femB, and fmhB(femX) genes have been shown to be essential for incorporation of glycine into the side chain of peptidoglycan precursors in Staphylococcus aureus,. The femABlocus was initially identified as a factor essential for methicillin resistance (fem) based on random insertional inactivation of chromosomal genes and a screen for reduced expression of resistance mediated by the penicillin binding protein 2A (PBP2A). Inactivation of femA or femB was subsequently reported to prevent incorporation of glycine residues at positions 2 to 5 or positions 4 to 5 of the penta-glycine cross bridge since muropeptides cross-linked by one or three glycine residues were detected in the corresponding mutants. Inactivation of fmhB, formerly femX, is lethal, but the construction of a mutant conditionally expressing fmhB under the control of a xylose-inducible promoter showed that the gene was essential for synthesis of branched peptidoglycan precursors. These studies show that the fem gene products were required for incorporation of glycine at positions 1 (FmhB), 2 and 3 (FemA), and 4 and 5 (FemB) of the cross bridge, although the catalytic activity of the proteins has not been directly assessed. Similarly, inactivation of two fmhB homologues in Streptococcus pneumoniae, designated murM (fibA) and murN (fibB), reduced addition of L-Ala or L-Ser to the -amino group of L-Lys and subsequent addition of a second L-Ala residue, respectively. Overall, disruption of the murMN operon reduced the proportion of branched peptide stems in the peptidoglycan from 89 to 33%. In contrast to what occurs in S. aureus, direct cross-linking of L-Lys to D-Ala occurs in S. pneumoniae, and the murMN operon was accordingly reported to be unessential.

BLAST analysis of *Alloiococcus otitis* genome revealed an ORF similar to femX of Weissella viridescent, and fmhB of S. aureus. It suggests that in *Alloiococcus otitis* there is an enzyme with similar to FhmB function. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5 /Table 4 (Seq. ID No 97). The protein encoded by the gene is set forth in Seq. ID No. 98.

#### Assays for measuring FmhB function

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There are no *in vitro* biochemical assays to test enzymatic activity of *S. aureus* FmhB because the reaction occurs at the membrane-bound lipid II precursor GlcNAc-(β-1,4)-*N*- acetylmuramic acid(-L-Ala-D-iGln-L-Lys-D-Ala-D-Ala)-pyrophosphoryl-undecaprenol.

Lipid II is a minor component of bacterial cell membrane which is detected by thin-layer chromatography separation of presolubilized membranes supplied with the cytoplasmic precursors, UDP-N-acetylmuramyl-pentapeptide (UDP-MurNAcpentapeptide) and [14C]UDP-N-acetylglucosamine ([14C]UDP-GlcNAc).

The *in vitro* biosynthesis of branched lipid II of *S. aureus* requires whole-cell membranes, cytoplasmic PG precursors, glycine (<sup>14</sup>C labeled for detection of reaction products), purified tRNA, and an intracellular fraction that contains tRNA-activating enzymes. Therefore, the *in vitro* assay of *S. aureus* FmhB is a tedious procedure.

One way to facilitate this procedure is to use Weissella viridescens FemX or E. faecalis UDP-MurNac-pentapetide:L-alanine ligase. Recombinant Weissella viridescens FemX and E. faecalis UDP-MurNac-pentapetide:L-alanine ligase were purified, and their in vitro activity was demonstrated. The distinctive feature of these enzymes is that they catalyze the addition of a branching amino acid (Ala) to the cytoplasmic cell wall precursor UDP-MurNac-pentapetide.

Other bacteria for which the biosynthesis of Gly-containing branched UDP-MurNac-hexapeptide in cytoplasm was shown are *Streptomyces lividans* and *Streptomyces hydroscopicus*, although the enzymes were not isolated and their ligase activity remain to be demonstrated.

These new data open an opportunity to develop an assay to detect the activity of FmhB(FemX) by using cytoplasmic UDP-MurNac-pentapetide. Products of the reaction are detected by HPLC. HPLC separation of precursors are performed by the method of Flouret et al. The precursors are separated by reverse-phase HPLC on a µBondapak C<sub>18</sub> column (3.9 by 300 mm; Waters) in 50 mM ammonium formate (pH 3.9) at a flow rate of 0.5 ml/min. The elution of precursors is monitored at a wavelength of 254 nm.

## EXAMPLE 16 FOLA- DIHYDROFOLATE REDUCTASE (DHFR)

The *Alloiococcus* ORF-1863 encodes a homolog of *S. aureus* dihydrofolate reductase that catalyzes the NADPH-dependent conversion of dihydrofolate to tetrahydrofolate, one of the steps in bacterial folate biosynthesis. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 55). The protein encoded by the gene is set forth in Seq. ID No. 56.

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#### FOLA as a target for anti-infective development

Folate is an essential cofactor in many important metabolic processes in bacteria, such as purine, pyrimidine, amino acid and pantothenate biosynthesis. Unlike mammalian cells, bacteria are unable to utilize exogenous folate derivatives, and therefore must synthesize folate *de novo*. Bacterial folate biosynthesis occurs via two converging pathways, the non-essential para-amino-benzoate (PABA) synthesis pathway, and synthesis of the pterin precursor, to which pABA is subsequently attached to form the folate precursor. Bacterial DHFRs are essential for viability and well conserved across all bacterial species. Although bacterial DHFR shares similarity with human DHFR, selective inhibitors against bacterial DHFR have been identified in the past such as trimethoprim which specifically blocks the bacterial DHFR step. Thus DHFR still remains an attractive target for development of broadspectrum antibacterial agents.

#### Assays for measuring DHFR activity

DHFR activity is monitored spectrophotometrically, recording the change of absorbance at 340 nm due to the equimolar consumption of NADPH in the course of dihydrofolate substrate reduction. DHFR (10 ng) is preincubated in reaction buffer containing 50 mM 2-(N-morpholino)ethanesulfonic acid, 25 mM Tris-HCl, 25 mM ethanolamine, and 100 mM NaCl at pH 7.5 for 3 minutes. The reaction is started by addition of 0.5-10  $\mu$ M 7,8-dihydrofolate. The amount of processed substrate is calculated from the decrease of absorbance at 340 nm due to oxidation of NADPH ( $\square$ =11800 M<sup>-1</sup>cm<sup>-1</sup>) to NADP<sup>+</sup>.

## EXAMPLE 17 FOLB- DIHYDRONEOPTERIN ALDOLASE (DHNA)

The Alloiococcus otitidis ORF-959 encodes a homolog of *S. aureus* dihydroneopterin aldolase that catalyzes the conversion of 7,8-dihydroneopterin to 6-hydroxymethyl-7,8-dihydropterin, one of the early steps in bacterial folate biosynthesis. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 31). The protein encoded by the gene is set forth in Seq. ID No. 32.

#### FOLB as a target for anti-infective development

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Folate is an essential cofactor in many important metabolic processes in bacteria, such as purine, pyrimidine, amino acid and pantothenate biosynthesis. Unlike mammalian cells, bacteria are unable to utilize exogenous folate derivatives, and therefore must synthesize folate *de novo*. Bacterial folate biosynthesis occurs via two converging pathways, the non-essential para-amino-benzoate (*p*ABA) synthesis pathway, and synthesis of the pterin precursor, to which *p*ABA is subsequently attached to form the folate precursor. Enzymes that catalyze steps in the folate biosynthesis pathway are essential and well conserved across all bacterial species, and those that act in early steps such as FolB have no direct homologs in mammals. Thus FolB becomes an attractive target for development of broad-spectrum antibacterial agents.

#### Assays for measuring FoLB activity

FoIB (DHNA) 7,8-dihydroneopterin aldolase activity is monitored individually or in conjunction with downstream enzymes in folic acid biosynthesis pathway (FoIK and Sul).

FoIB activity is monitored directly by HPLC assay. FoIB substrate (7,8-dihydro-D-neopterin) is commercially available from Schircks Laboratories (Swizerland). FoIB (0.5  $\mu$ g) is preincubated in reaction buffer containing 50 mM Tris-HCI (pH 8.0), 50 mM KCI, 0.1 mg/ml BSA, 2.5 mM dithiothrietol for 5 min. Reaction is started by addition of stock solution of 7,8-dihydro-D-neopterin in DMSO (100  $\mu$ M

final concentration). Reaction is terminated by addition of 1/3 of reaction volume of 1% I<sub>2</sub>, 2% KI in 1M HCI with subsequent incubation at room temperature for 5 minutes. Quenched reaction will be applied directly to HPLC. Oxidized starting material and reaction products are efficiently separated on ODS (C18) column. Reaction components are detected and quantified by analysis of UV absorbance at 254 nm, or fluorescence (excitation at 365 nm; emission at 446 nm).

FolB activity are also monitored in the coupled assay with FolK (HPPK) and Sul (DHPS) enzymes. FolB activity is measured by detection of radioactive dihydropteroate formation as described in FolK and Sul assays, under conditions of excess of the later enzymes. FolB enzyme and substrate 7,8-dihydro-D-neopterin are added to the described assay to replace the 6-hydroxymethyl-7,8-dihydropterin (FolK substrate).

## EXAMPLE 18 FOLC- DIHYDROFOLATE SYNTHASE (DHFS)

The *Alloiococcus otitidis* ORF-956 and ORF-528 both encode a homolog of *B. subtilis* dihydrofolate synthase that catalyzes the conversion of 7,8-dihydropteroate and glutamate to dihydrofolate, one of the steps in bacterial folate biosynthesis [. Homologue of this gene identified in *Alloiococcus otitidis* as described in Example 5 (Seq. ID Nos. 29 and 23). The protein encoded by the gene is set forth in Seq. ID Nos. 30 and 24.

#### Use of FoLC as a target for anti-infective development

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Folate is an essential cofactor in many important metabolic processes in bacteria, such as purine, pyrimidine, amino acid and pantothenate biosynthesis. Unlike mammalian cells, bacteria are unable to utilize exogenous folate derivatives, and therefore must synthesize folate *de novo*. Bacterial folate biosynthesis occurs via two converging pathways, the non-essential para-amino-benzoate (*p*ABA) synthesis pathway, and synthesis of the pterin precursor, to which *p*ABA is subsequently attached to form the folate precursor. Enzymes that catalyze steps in the folate biosynthesis pathway are essential, and are well conserved across all bacterial species. Bacterial FolC appears to be a bifunctional enzyme possessing both

dihydrofolate synthase (DHFS) activity and folyl-polyglutamate synthetase (FPGS) activity, which are probably mediated through different sites of the protein. The bacterial DHFS activity but not the FPGS activity is essential for viability. Although bacterial FolC shares similarity with human FPGS, the human enzymes apparently lack DHFS activity and display a folate substrate specificity quite distinct from that of bacterial enzymes. Thus targeting bacterial FolC/DHFS activity selectively might lead to identification of broad-spectrum antibacterial agents.

#### Assays for measuring FoLC activity

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FoIC (DHFS) 7,8-dihydrofolate synthase activity in the presence or absence of antimicrobial compounds or putative inhibitory compounds are monitored by several methods.

In one method, FolC activity is monitored directly by simple HPLC assay. FolC substrate (7,8-dihydropteroic acid) is commercially available form Schircks Laboratories (Switzerland). FolC (15 ng) is added to reaction mix, containing 10 mM glutamate, 5 mM ATP, 50 mM Tris-HCl (pH 8.0), 20 mM Mg<sub>2</sub>Cl, 50 mM KCl, 0.1 mg/ml BSA, 5 mM dithiothreitol. Reaction is started by addition of stock solution of 7,8-dihydropteroic acid in DMSO (10 µM final concentration). Reaction is terminated by addition of equal volume of 8M Guanidinium hydrochloride. Stopped reaction is applied directly to HPLC. Starting material and reaction products are efficiently separated on ODS (C18) column. Reaction components are detected and quantified by analysis of UV absorbance at 254 nm, or fluorescence (excitation at 280 nm; emission at 420 nm).

In another method, the FolC activity monitoring is by detection of ADP accumulation. ADP is released in the amount equimolar to the amount of the product formed. ADP detection is performed by coupling its conversion to ATP by pyruvate kinase in the presence of phospho(enol)pyruvate producing pyruvate. Lactate dehydrogenase reduces pyruvate to S-lactate in the presence of NADH. Course of reaction is monitored by decrease in absorbance at 340 nm due to oxidation of NADH ( $\epsilon$ =6220 cm<sup>-1</sup>M<sup>-1</sup>) to NAD<sup>+</sup>. Reaction conditions are as following: 5 mM dithiothreitol, 5 mM ATP, 380  $\mu$ M NADH, 10 mM glutamate, 2 mM phospho(enol)pyruvate, 50 mM KCl, 20 mM Mg<sub>2</sub>Cl, 50 mM Tris-HCl, 50  $\mu$ g of

pyruvate kinase, 50  $\mu g$  of S-lactate dehydrogenase. Reaction is started by addition 7,8-dihydropteroic acid in DMSO (10  $\mu M$  final concentration).

In yet another method, FolC activity is monitored through detection of inorganic phospate release. Amount of inorganic phosphate in solution is quantified by:

- (i) its conversion by purinenucleoside phosphorylase leading to phosphorylation of MESG. Later assay kit is available from Molecular Probes as EnzCheck™ Phosphate Assay Kit;
- (ii) its reaction with Malachite Green reagent; and
- (iii) detecting the release of radioactive inorganic phosphate in reaction with  $\gamma$   $^{33}$ P-labeled ATP following the absorption of unprocessed ATP by charcoal.

First method is applied in rate-based assay format; the later two in end-point format. Reaction conditions are similar to the ones described in HPLC-based assay.

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#### **EXAMPLE 19**

#### FOLK- 6-HYDROXYMETHYL-7, 8-DIHYDROPTERIN PYROPHOSPHOKINASE (HPPK)

The *Alloiococcus otitidis* OFR-961 (Seq. ID No. 33) encodes a homolog of *S. aureus* 6-hydroxymethyl-7,8-dihydropterin pyrophosphokinase that catalyzes pyrophosphoryl transfer from ATP to 6-hydroxymethyl-7,8-dihydropterin, one of the early steps in bacterial folate biosynthesis. The protein encoded by this ORF is set forth in Seq. ID No. 34. (see Example 5/Table 4).

#### 25 Use of Folk as a target for anti-infective development

Folate is an essential cofactor in many important metabolic processes in bacteria, such as purine, pyrimidine, amino acid and pantothenate biosynthesis. Unlike mammalian cells, bacteria are unable to utilize exogenous folate derivatives, and therefore must synthesize folate *de novo*. Bacterial folate biosynthesis occurs via two converging pathways, the non-essential para-amino-benzoate (*p*ABA) synthesis pathway, and synthesis of the pterin precursor, to which *p*ABA is subsequently attached to form the folate precursor. Enzymes that catalyze steps in the folate biosynthesis pathway are essential and well conserved across all bacterial species, and those that act in early steps such as FolK have no direct homologs in mammals.

Thus FolK is an attractive target for the development of broad-spectrum antibacterial agents.

#### Assays for measuring Folk activity

Folk (HPPK) 7,8-dihydroxymethylpterin-pyrophosphokinase activity is monitored individually or in conjunction with downstream enzyme in folic acid biosynthesis pathway.

FolK activity is monitored directly by HPLC assay. FolK substrate (7,8-dihydro-6-hydroxymethylpterin) is commercially available from Schircks Laboratories (Swizerland). FolK is preincubated in reaction buffer containing 50 mM Tris-HCl (pH 8.0), 50 mM KCl, 20 mM MgCl<sub>2</sub>, 5 mM ATP, 0.1 mg/ml BSA, 2.5 mM dithiothrietol. Reaction is started by addition of stock solution of 7,8-dihydro-6-hydroxymethylpterin in DMSO (100  $\mu$ M final concentration). Reaction is terminated by addition of equal volume of 8M Guanidinium hydrochloride and applied directly on HPLC. Starting material and reaction products are efficiently separated on ODS (C18) column. Reaction components are detected and quantified by analysis of UV absorbance at 254 nm.

Folk activity is monitored by end-point assay coupled with excess of Sul enzyme. Activity is calculated from quantification of the radioactivity incorporated in final product (7,8-dihydropteroate).

#### **EXAMPLE 20**

#### ALLOIOCOCCUS OTITIDIS ENCODED FOLP (SUL)- DIHYDROPTEROATE SYNTHASE (DHPS)

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The *Alloiococcus otitidis* ORF-1811 (Seq. ID No. 53) encodes a homolog of *B. subtilis* dihydropteroate synthase that catalyzes the condensation of pABA (para-aminobenzoic acid) with 6-hydroxymethyl-7,8-dihydropterin pyrophosphate, one of the early steps in bacterial folate biosynthesis. The polypeptide encoded by this ORF is set forth in Seq. ID No. 54. (see Example 5/Table 4)

#### FOLP AS A TARGET FOR ANTI-INFECTIVE DEVELOPMENT

Folate is an essential cofactor in many important metabolic processes in bacteria, such as purine, pyrimidine, amino acid and pantothenate biosynthesis. Unlike

mammalian cells, bacteria are unable to utilize exogenous folate derivatives, and therefore must synthesize folate *de novo*. Bacterial folate biosynthesis occurs via two converging pathways, the non-essential para-amino-benzoate (*p*ABA) synthesis pathway, and synthesis of the pterin precursor, to which *p*ABA is subsequently attached to form the folate precursor. Enzymes that catalyze steps in the folate biosynthesis pathway are essential and well conserved across all bacterial species, and those that act in early steps such as FoIP (SuI) have no direct homologs in mammals. In fact, dihydropteroate synthase (FoIP or SuI) is the target for known antibiotics sulfonamides which are competitive inhibitors of FoIP/SuI as *p*ABA analogues. Thus FoIP (SuI) still remains an attractive target for development of broad-spectrum antibacterial agents.

#### Suitable assays for measuring FoIP/Sul activity

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Sul (DHPS) 6-hydroxymethy-7,8-dihydroneopteroate synthase activity is monitored individually or in conjunction with upstream enzymes in folic acid biosynthesis pathway (FoIB and/or FoIK).

DHPS activity is monitored directly by counting the amount of radioactivity incorporated in 6-hydroxymethy-7,8-dihydroneopteroate when using radioactively labeled *p*-aminobenzoic acid (*p*ABA). Final product is separated from unreacted *p*ABA by thinlayer chromatography, paper chromatography or on HPLC equipped with radioactivity detector. DHPS substrate (6-hydroxymethyl-7,8-dihydropterin pyrophosphate) is not commercially available, but is quantitatively synthesized in one step from its oxidized precursor available from Schircks Laboratories (Swizerland). DHPS (20 ng) is added in reaction buffer containing 50 mM Tris-HCl, pH 8.0, 20 mM MgCl<sub>2</sub>, 0.1 mg/ml BSA, 5 mM dithiothreitol and 0.5 – 10 μM PABA. Reaction is started by addition of stock solution of substrate (6-hydroxymethyl-7, 8-dihydropterin pyrophosphate, 0.05 - 1 μM final concentration). Reaction is terminated by acidification of reaction volume with addition of equal volume of citrate/phosphate or ammonium acetate/acetate buffer, pH 4 containing excess of unlabelled *p*ABA. Quenched reaction is separated by chromatography and the amount of formed product calculated.

DHPS activity is determined in coupled assay with excess of FolB and FolK enzymes. The advantage of coupled assay is that it makes it possible to use

commercially available FolB (7,8-dihydro-D-neopterin), or FolK (6-hydroxymethyl-7,8-dihydropterin) substrates, thus forming DHPS substrate *in situ*.

#### **EXAMPLE 21**

### ALLOIOCOCCUS OTITIDIS ENCODED FILAMENTATION TEMPERATURE SENSITIVE GENE A (FTSA)

The *Alloiococcus* otitidis ORF-2489 (Seq. ID No. 85) encodes a homolog of *E. faecalis* FtsA, one of the essential components of bacterial cell division. The "fts" stands for <u>filamentation temperature sensitive</u> and has been assigned to most bacterial cell division genes due to the fact that these genes were generally discovered by the isolation of conditional mutants that form filaments at nonpermissive temperature. The *ftsA* allele was first isolated and identified in *E. coli* by Ricard and Hirota in 1973, and mapped along with *ftsZ* in 1980. The protein encoded by this ORF is set forth in Seq. ID No. 86. (see Example 5/Table 4)

Bacterial cell division requires formation of a septum at mid-cell that begins with the polymerization of FtsZ into a ring structure at the nascent division site. FtsZ, another key component of bacterial septation is the first known protein to localize to the division site. In *E. coli*, shortly after the formation of the FtsZ ring, FtsA and ZipA (another key division component present only in gram-negative bacteria) [7] are independently recruited to the septal ring, most likely through their direct interaction with FtsZ. Subsequent assembly of other division components at the septum requires FtsA as well as FtsZ.

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#### FtsA as a target for anti-infective development

Like FtsZ, FtsA homologs are present and highly conserved in almost all eubacteria. FtsA is essential for cell division and its deletion leads to impaired cell division and sporulation defect. In addition, *E. coli* cells have to maintain critical ratio of FtsA to FtsZ in order for proper cell division to occur. FtsA belongs to the actin/DnaK/sugar kinase family of proteins. In *B. subtilis*, FtsA acting as a dimer not only binds ATP but also hydrolyzes ATP. As briefly stated above, *in vivo* and *in vitro* evidence have demonstrated that FtsA and FtsZ from various bacterial species

directly interact. Taken all together, targeting at FtsA especially at its interaction with FtsZ might lead to identification of broad-spectrum antibacterial agents.

#### Assays for measuring FtsA activity

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ATPase activity of FtsA is assayed by following the formation of  $^{32}$ Pi from [ $\gamma$ - $^{32}$ P]-ATP. The reaction mixture containing 50 mM Tris-HCl (pH7.2), 50 mM potassium acetate, 1 mM DTT, 10 mM MgCl<sub>2</sub> and different concentrations of [ $\gamma$ - $^{32}$ P]-ATP is incubated for 5 minutes at 37°C. The reaction is started by addition of 50 nM purified FtsA of *Alloiococcus*. The reaction is stopped with 1.5% ammonium molybdate in 0.5N sulfuric acid, and the radioactive Pi extracted into isoamyl alcohol and counted.

Interaction between FtsA and FtsZ is detected quantitatively using yeast two-hybrid system as described. Briefly, *Alloiococcus ftsZ* is cloned into yeast two-hybrid bait vector pLexA (Clontech) to generate a LexA-FtsZ fusion with DNA-binding property. *Alloiococcus ftsA* is cloned into the target vector pB42AD (Clontech) to fuse FtsA to the activating domain. Both plasmids are then transformed into a *Saccharomycyces cerevisiae* strain containing a *lacZ* reporter under the control of multiple LexA operators. β-Galactosidase activity is determined to quantify relative strength of FtsA-FtsZ interaction.

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#### **EXAMPLE 21**

### ALLOIOCOCCUS OTITIDIS ENCODED FILAMENTATION TEMPERATURE SENSITIVE GENE Z (FTSZ)

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FtsZ is an essential protein that forms a cytokinetic ring (Z-ring) that drives cell division in bacteria. FtsZ has been identified in most prokaryotic species with the exception of *Chlamidia*, a *Ureaplasma* species and *Crenarchaea*. FtsZ and Z-ring formation are most extensively studied in *E. coli*. FtsZ is an abundant cytoplasmic protein which is present at ~ 10<sup>4</sup> copies per cell, and is the first protein to be localized to the division site. Z-ring is required throughout septation and directs the ingrowth of septum in part by recruiting other cell division protein to the division site. Another function is suggested by FtsZ homology to eukaryotic tubulins. Like tubulin, FtsZ is a GTPase and undergoes GTP/GDP-dependent polymerization. Recent studies showed that Z-ring is a very dynamic structure suggesting that GTP-dependent

assembly/disassembly of Z-ring might provide constriction force to power cell division. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 83). The protein encoded by the gene is set forth in Seq. ID No. 84.

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#### **GTPase activity**

FtsZ is a GTPase that contains the tubulin-signature nucleotide-binding motif GGGTGS/TG. Like in DD-tubulin dimer, the active site for GTP-hydrolysis appears to be shared between two subunits where the GTP-binding pocket is provided by one subunit while the GTPase-activating T7 loop comes from the other subunit. This view is supported by genetic analysis as various mutations that inhibit FtsZ GTPase activity map in the T7-loop region and a conserved Asp-residue in T7-loop is found to be involved in the coordination of the cation involved in GTP hydrolysis. FtsZ GTPase activity is Mg²+-dependent and is stimulated by KCI.

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#### **Polymerization**

In vivo, about 75% of FtsZ is present as multimers. In vitro, FtsZ forms a variety of structures at various conditions. FtsZ assembles into thin protofilaments with GTP and formation of FtsZ polymers is coupled to GTP hydrolysis: when GTP runs out, polymers disassemble. Protofilaments assemble into sheets and bundles in the presence of multimolar amounts of either Mg²+ or Ca²+ or by addition of DEAE-dextran. In addition, ZipA protein induces bundling of FtsZ polymers. With GDP, FtsZ assembles into curved filaments and minirings.

#### Interactions with other proteins

In *E. coli*, at least nine different proteins are localized to the division septum and are required for cell division to proceed. Among them two proteins, ZipA and FtsA, are shown to interact directly with FtsZ. Both of these proteins localize to the division site independently from each other, but require FtsZ for localization. ZipA is an integral membrane protein which is thought to mediate invagination of cell membrane by linking the membrane to constricting Z-ring. Interaction between ZipA and FtsZ is confined to C-terminal portion of ZipA (residues 185-328) and conserved 17-amino acid region on C-terminus of FtsZ. FtsA is an actin-like membrane-associated protein

which possesses ATPase activity and might provide energy required for Z-ring dynamics. Interaction between FtsZ and FtsA is not studied in great detail, it is shown that C-terminus of FtsZ is required. The remaining division proteins require both ZipA and FtsA for their localization to Z-ring.

FtsZ as a target for anti-infective development

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FtsZ is an essential protein for cell division/bacterial viability. Knock-out *ftsZ* mutants fail to divide and, as a result, filament and die. The target is widely conserved throughout bacterial kingdom implying that FtsZ-specific inhibitor would have a broad-spectrum antibacterial activity. The potential drawbacks of the target might include the presence and the essential role of a homolog (tubulin) in eukaryotes and an intrinsic difficulty in inhibiting protein-protein interactions by small molecules. Although this target is being studied extensively, no FtsZ-specific compounds are reported up to date.

#### Assays for measuring FtsZ function

Polymerization of FtsZ is measured by light scattering assay as described previously. FtsZ (12.5  $\mu$ M) is incubated in 200  $\mu$ I of polymerization buffer (50 mM MES/NaOH, pH 6.5, 50 mM KCl, 5 mM MgCl<sub>2</sub>, 10 mM CaCl<sub>2</sub>) in a fluorescence cuvette with a 1 cm path length. The sample is maintained at 30°C, polymerization is induced by addition of 20-500  $\mu$ M GTP. Light scattering is measured at 90°, both excitation and emission wavelengths are set to 350 nm, slit width is 2 nm. Alternatively, the amount of polymerized FtsZ is analyzed by sedimentation and subsequent quantification of precipitated FtsZ by SDS-PAGE, Coomassie staining and densitometric scanning. In addition, polymers are observed by electron microscopy. This assay represents either primary or secondary/confirmatory assay.

GTP binding of FtsZ is monitored by the covalent cross-linking of [γ-<sup>32</sup>P]GTP (3000 Ci/mmol) to FtsZ in a previously described competition assay. FtsZ (3 μg) is incubated in 20 μl of 50 mM MES/NaOH, pH 6.5, 100 mM KCl, 4 mM MgCl<sub>2</sub>, 1 mM EDTA, 0.1 mM EGTA and 0.5 mM DTT. Various amounts of non-labeled competing nucleotide (GTP or GTP analogs) and 0.1 mM [γ-<sup>32</sup>P]GTP are added, samples are incubated at 0°C for 15 min, then UV cross-linked for 5 min and analyzed by SDS-

PAGE on 12% gel, autoradiography and densitometric scanning. This assay represents a secondary/confirmatory assay.

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The GTP hydrolytic activity of FtsZ is monitored by thin-layer chromatography (TLC) as described previously. Briefly, the reaction mixture consists of 5 mM of [Y-3²P]GTP (40 mCi/mmol), 15 mM magnesium acetate and 0.25-2 mg/ml of FtsZ in reaction buffer (40 mM Tris-acetate, pH7, 200 mM potassium acetate, 2 mM EDTA, 1 mM DTT and 0.5% Triton X-100), aliquots are separated by TLC and amount of GTP converted to GDP is determined by spot-densitometry. Alternatively, GTPase activity is measured either by quantitation of the non-radioactive inorganic phosphate with the malachite green-molybdate reagent as described previously or by quantitation by scintillation counting of radioactive inorganic phosphate released after hydrolysis of [Y-3²P]GTP (26). This assay represents either primary or secondary/confirmatory assay.

Among interactions of FtsZ with various cell division proteins, interaction between FtsZ and ZipA is characterized the best. ZipA –induced bundling of FtsZ is measured by the light scattering assay that is described above, both proteins are used at  $\geq 5~\mu\text{M}$ .

#### EXAMPLE 22

## ALLOIOCOCCUS OTITIDIS ENCODED GYRA/GYRB (DNA GYRASE, TOPOISOMERASE II) AND GRLA/GRLB (TOPOISOMERASE IV)

DNA topoisomerases: topoisomerases modulate the topological state of DNA in cells. This involves binding to DNA, introducing single or double stranded breaks in the DNA, passing DNA molecules through the break and rejoining the break. This controls the levels of positive and negative supercoiling of DNA and functions in catenation/decatenation. Controlling the topological state of DNA is critical to the fundamental processes of transcription, recombination, replication and partitioning of the chromosome. There are two main categories of topoisomerases, type I and type II. Type I topoisomerases introduce single stranded breaks in DNA whereas type II enzymes introduce double stranded breaks. GyrA/GyrB (gyrase) and GrIA/GrIB (topoisomerase IV) are both type II enzymes that are essential for cell viability.

DNA gyrase (GyrA/GyrB) is a type II topoisomerase that functions to control the degree of supercoiling in double stranded DNA. It is essential for viability and

plays central roles in replication, repair, recombination and transcription of DNA. Gyrases have the ability to introduce double stranded breaks in DNA molecules while remaining bound to the DNA through phosphotyrosine bonds, pass uncut DNA through the break and then rejoin the breaks, with repeated cycles being driven by the hydrolysis of ATP. Gyrase has the unique ability to introduce negative supercoils in closed circular DNA and also functions to catenate/decatenate DNA duplexes. The generation of negative supercoiling is important for initial stages in replication. DNA gyrase from Escherichia coli has been studied in detail. It is a complex of two subunits of GyrA (encoded by gyrA) and two subunits of GyrB (encoded by gyrB) (ie.  $A_2B_2$  complex). The subunits are organized in discreet domains. An N-terminal domain of GyrB harbors ATPase activity while the C-terminal domain is thought to interact with the GyrA subunit, and is involved in DNA binding. The N-terminal domain of GyrA is apparently involved in DNA strand breakage-ligation reactions while the C-terminal segment is involved in DNA binding. Crystal structures of the DNA strand breakage/reunion domain of E. coli GyrA, and the N-terminal ATPase domain of E. coli GyrB have been determined. DNA gyrase has also been purified and characterized from gram positive organisms such as S. aureus. Comparison of DNA gyrases from several bacteria reveal a high degree of conservation of important domains.

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Topoisomerase IV (GrIA/GrIB) is a type II topoisomerase but unlike gyrase it does not possess negative supercoiling activity. Its primary role in replication appears to be in the decatenation of multiply linked daughter chromosomes, important for terminal stages of the replication process. Topoisomerase IV has been purified and characterized from gram negatives eg. *E. coli*, (where the GrIA/GrIB subunit homologs are designated ParC and ParE), and gram positives eg *S. aureus*. Homologs of the gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID Nos 17 and 19). The proteins encoded by the genes are set forth in Seq. ID Nos. 18 and 20.

GyrA/GyrB (Gyrase) and GrlA/GrlB (topoisomerase IV) as targets for antiinfective development:

Alloiococcus otitidis is an infectious organism associated with disease, and consequently, novel antimicrobials to combat these infections are desirable. DNA gyrase and Topoisomerase IV is essential for bacterial viability and is a well-established and validated antibacterial target.

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Purification of DNA gyrase and topoisomerase IV from Alloiococcus otitidis

Genes encoding the GyrA/GyrB and GrlA/GrlB subunits or their functional domains are obtained using polymerase chain reaction amplification of the genomic region encoding them. The genes are then subcloned into standard expression vectors, with or without affinity tags. The enzyme is then overexpressed in *Escherichia coli* and purified using a standard tag system or conventional chromatography.

Measurement of gyrase and topoisomerase IV by kinetoplast DNA decatenation assay:

Type II topoisomerases introduce double stranded breaks in DNA and mediate catenation/decatenation of DNA. Topoisomerase IV activity is readily determined with decatenation assays using as substrate kinetoplast DNA (KDNA) from *Crithidia fasciculata*. The DNA isolated in this procedure is a highly networked series of catenated double stranded minicircles and is easily be pelleted by centrifugation. The activity of topoisomerase II enzymes results in the release of decatenated DNA minicircles from the networked KDNA. These have a high mobility in agarose gels and migrate into the gel ahead of the networked material, which has very low mobility, allowing for determination of decatenation activity using ethidium bromide stained agarose gel electrophoresis.

Alternatively, using radiolabeled KDNA, the level of decatenation activity is measured by counting radioactivity remaining in reaction supernatants following centrifugation to pellet the networked material. Typical conditions used for assaying decatenation activity of *S. aureus* and *E. coli* topoisomerase IV activity are as follows: *C. fasciculata* KDNA (0.9 mg/ml) is incubated in 40 µl of reaction buffer (50 mM Tris-HCl, pH 7.7, 5 mM MgCl<sub>2</sub>, 5 mM DTT, 50 µg/ml bovine serum albumin, 1.5 mM ATP

and 350 mM potassium glutamate) with appropriate amounts of the Grl subunits, for 1 hour at 37° C. If non radiolabeled KDNA is used, these reactions can be stopped and analyzed by agarose gel electrophoresis, or for radioassays, the reaction is stopped by gentle mixing with 10 µl of stop solution (50 % glycerol, 50 mM EDTA (pH 8.0), 2.5 % SDS and 0.1 % bromphenyl blue) and centrifuged at 15 000 x g for 5 min at 20° C. Decatenation activity is determined by counting radioactivity in 25 µl of the supernatant in a scintillation counter. Alternatively, a modified assay employing flow injection fluorometry of 4', 6-diaminidino-2-phenylindole (DAPI) treated supernatants has been described that could be suitable for moderate throughput non radioactive assays, or filtration of the reactions through appropriate filters may efficiently separate the decatenated species from KDNA. Although the above described assays were used for topoisomerase IV, modified decatenation reactions using KDNA isolated from *Leishmania donovani* reveal significant decatenation activity by gyrase from *E. coli* and *Mycobacterium smegmatis*, indicating the applicability of the assay to prokaryotic gyrases.

#### DNA Supercoiling/relaxation assays.

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DNA gyrase function is directly assayed using a simple supercoiling assay typified by that described for the measurement of *Escherichia coli* DNA gyrase activity. Briefly, incubation of relaxed closed circular plasmid DNA (pUC18, 7.5 nM) in the presence of DNA gyrase (approximately 10 nM) in 40 mM Tris-HCl (pH 8.0) buffer containing 25 mM KCl, 4 mM MgCl2, 2.5 mM spermidine and 1.4 mM ATP buffer results in the introduction of supercoils in the plasmid DNA. Changes in DNA supercoiling status are readily observed by the alteration of mobility of the DNA in agarose gels stained with ethidium bromide and comparison to the mobility of relaxed and supercoiled plasmid template. This strategy is employed for screening for DNA gyrase inhibitors.

Topoisomerase IV activity is assayed by measuring relaxation of supercoiled plasmid DNA. A typical relaxation assay used for *S. aureus* topoisomerase IV activity is as follows: topoisomerase IV enzyme and supercoiled plasmid DNA (pBR322, 0.6 µg) is incubated in 40 µl 50 mM Tris-HCl, pH 7.7, containing 5 mM MgCl<sub>2</sub>, 5 mM DTT, 50 µg/ml bovine serum albumin, 1.5 mM ATP, 5 mM spermidine and 20 mM KCl, for 30 min at 37°C. Changes in DNA supercoiling status can be

readily observed by the alteration of mobility of the DNA in agarose gels stained with ethidium bromide and comparison to the mobility of relaxed and supercoiled plasmid template

The ATPase activity of topoisomerases is measured using a coupled spectrophotometric ATPase assay described for the GyrB subunit of E. coli. ATPase activity is assayed in 300  $\mu$ l of 40 mM Tris-HCl (pH 8.0), containing 25 mM KCl, 2.5 mM spermidine, 4 mM MgCl2, 400  $\mu$ M phosphoenolpyruvate, 250  $\mu$ M NADH, 3  $\mu$ l of pyruvate kinase /lactate dehydrogenase mix and ATP (0.5 – 3.5 mM). The reaction is started by the addition of truncated N-terminal derivatives of the GyrB protein (5  $\mu$ M) containing the ATPase domain. ATPase activity is reflected as a decrease in absorbance of light at 340 nanometer wavelength.

### DNA cleavage assay.

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Quinolone drugs interfere with the DNA strand breakage-ligation cycle activity of many topoisomerases. Incubation of topoisomerase and linear or supercoiled pBR322 plasmid DNA, or small linear DNA fragments, in the presence of quinolones and magnesium results in the trapping of a complex of topoisomerase, DNA with a double stranded break and the drug. The topoisomerase remains bound to the cleaved DNA, however treatment with a denaturant such as SDS or proteinases remove/degrade the gyrase, releasing the cut DNA. Certain consensus sequences representing preferred cut sites of E. coli gyrase in plasmid pBR322 have been identified in template DNA molecules used in these assays. This assay is useful for mode of action studies of inhibitors of gyrase/topoisomerase IV activity and in particular of the strand breakage-ligation function. Cleavage reactions are performed with linear or supercoiled DNA. A typical cleavage reaction using linear DNA to measure cleavage by E. coli and S. aureus gyrase and topoisomerase IV in the presence of drugs is as follows: gyrase/ topoisomerase IV is incubated in 20 µl 25 mM Tris-HCI (pH 7.5) containing 0.5 mM EDTA, 0.5 mM DTT, 3 µg bovine serum albumin per ml, 10 mM MgCl<sub>2</sub>, 120 mM KCL<sub>1</sub> 10 mM ATP, 10 000 dpm of 3' end labeled linear pBR322 plasmid DNA and drug for 1 hour at 37°C. (Note: for S. aureus, KCI is replaced with 0.7 M potassium glutamate). Reactions are terminated by adding 5 µl 2.5% SDS-2.5 mg proteinase K per ml and incubating at 37°C for 30 minute, then adding 5 µl 30% glycerol-1% SDS-50 mM EDTA-0.05 % bromophenol

blue. Cleavage products are resolved on 1% agarose gels and visualized by autoradiography.

Additional cleavage assays are also used that measure 1) the linearization of supercoiled plasmid DNA (pBR322), with linearization measured using scanning densitometry of DNA species separated on 1 % agarose gels, or 2) the cleavage of small linear DNA molecules of approximately 100 bp encompassing the preferred cleavage sequence 5'- GGCTGGATGGCCTTCCCCAT - 3' from position 990 in plasmid pBR322. In the latter case, the fragment is produced by PCR and radiolabeled with y-32P ATP at the 5' end of the top strand. This DNA is incubated with 1.3 pmol DNA gyrase in a total volume of 10 µl 35 mM Tris-HCl (pH 8.0), 24 mM KCI, 2 mM spermidine, 4 mM MgCl2 and inhibitor compound at 37°C for 10 min. Reactions are stopped by addition of 8 mM EDTA and 1% SDS, then treated with 500 µg/ml proteinase K for 2 hours at 37°C. The DNA is then cleaned by phenolchloroform extraction and ethanol precipitation, resuspended in TE buffer (pH 8.0), and loaded and resolved on 12 % sequencing gels containing 7M urea. In the presence of inhibitors of the strand breakage-ligation function, radioactive cleavage products are detectable by autoradiography. Modifications of this assay whereby one strand of the DNA substrate is labeled with an affinity tag such as biotin and the other is radiolabeled or fluorescently labeled should facilitate rapid separation and detection of cleavage products using streptavidin coated columns or plates, resulting in higher assay throughput.

#### GYRASE ACTIVITY ASSAYS: DNA REPLICATION:

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Early work by Fuller and Kornberg revealed that a partially purified crude soluble fraction derived from *Escherichia coli* cells (designated fraction II) contained the components necessary for replication of plasmids containing oriC (*E. coli* chromosomal origin of replication). Replication mediated by this fraction specifically required supercoiled plasmids. Although the exact makeup of the protein complex mediating the replication was not known, the replication reaction was inhibited by 1) rifampicin, and 2) nalidixic acid and novobiocin, indicating essential roles for both RNA polymerase and DNA gyrase, respectively. Subsequently the reaction was reproduced using replication machinery reconstituted from purified protein HU, DnaA,

DnaC, DnaB, single stranded binding protein (SSB), primase, DNA polymerase holoenzyme, RNA polymerase holoenzyme and GyrA/GyrB.

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The requirement for gyrase activity for replication is exploited for the identification of gyrase inhibitors using a replication-based high throughput screen. Gyrase specific inhibitors are identified from the overall pool of replication inhibitors using the secondary assays detailed below. Screening for inhibitors of gyrase in a setting where gyrase is participating in an overall reaction that is essential in bacteria might better select physiologically relevant inhibitors

An assay suitable for high throughput screening of inhibitors of replication (including gyrase and DnaA inhibitors) is based on the replication reaction of Kaguna and Kornberg. This reaction was set up as follows; standard reaction in 25 μl: 40 mM Hepes (pH 7.6), 2 mM ATP, 0.5 mM GTP, CTP and UTP, 50 μg/ml bovine serum albumin, 6 mM phospho creatine, 100μM dATP, dGTP, dCTP and dTTP, γ-<sup>33</sup>P dTTP (50-150 cpm/pmol total nucleotides) 11mM magnesium acetate,100 μg/mL creatine kinase,85 ng SSB, 48 ng DnaB, 40 ng DnaC, 20 ng primase, 160 ng DNA polymerase III holoenzyme, 800 ng RNA polymerase, 150 ng GyrA, 350 ng GyrB, 120 ng DnaA, 2.5 units topoisomerase I, 190 ng HU, 0.15 ng Rnase H 200 ng supercoiled plasmid template. The reaction is assembled at 0 °C and initiated by incubation at 30°C. Replication reactions are terminated by the addition of EDTA to 20 mM. Incorporation of nucleotides into DNA is measured by filtration through 96 well DEAE filter plates and counting retained radioactivity.

Compounds inhibiting gyrase activity in Alloiococcus otitidis are found as part of a larger program directed at replication. This reaction described above uses the replication machinery of a gram-negative organism, which differs somewhat from the replication machinery of gram positives such as *Staphylococcus aureus* with respect to the specific protein subunits involved. Therefore a similar system specific to *Alloiococcus otitidis* is assembled from the relevant proteins purified from *Alloiococcus otitidis*. Several techniques are then utilized to determine the interaction of inhibitors with Gyr A and GyrB. These are described in the literature and include a) Nuclear magnetic resonance; and b) Capillary electrophoresis.

#### Example 23

### ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYMES MURA

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Bacterial cell wall peptidoglycan (murein) is a large macromolecule of periodic structure whose basic unit, a disaccharide-peptapeptide, is polymerized linearly via the disaccharide motif and cross-linked laterally via the peptide motif. The process of bacteria cell wall biosynthesis starts from the transferase MurA, which transfers the addition of an enolpyruvyl moiety to the 3'-hydroxyl-UDP-N-acetyl glycosamine (UDP-GluNAc). Subsequently, the reductase MurB reduces the enol ether to the lactyl ether, utilize one equiv. of NADPH and a solvent proton to form UDP-*N*-acetyl muramic acid (UDP-MurNAc). Next a series of ATP dependent amino acid ligases (MurC, MurD, MurE and MurF) catalyze the stepwise synthesis of the pentapeptide side chain using the newly synthesized carboxylate as the first acceptor site. Each enzyme is responsible for the addition of one more residue except MurF, catalyzes D-ala-D-ala. MurE in gram negative bacteria catalyzes the meso-2, 6-diaminopimelate (DAP), while in gram positive bacteria MurE catalyzes L-lysine.

The product of MurF, UDP-NAM pendapeptide is the final product of the cytoplasm enzymes and is the most important precusor for further peptidoglycan biosynthesis. UDP-MurNAc pendapeptide is then and catalyzed at the plasma membrane by the membrane bound enzymes such as the translocase MraY and transferase MurG.

UDP-*N*-acetylglucosamine enolpyruvyl transferase (MurA) catalyzes the first committed step in bacterial cell wall biosynthesis. The enzyme transfers an enolpyruvyl group from phosphoenolpyruvate (PEP) to UDP-*N*-acetylglucosamine (UDP-GluNAc) to the 3'-OH of UDP-GlcNAc by an addition-elimination mechanism that proceeds through a tetrahedral ketal intermediate. MurA product enolpyruvate UDP-*N*-acetylglucosamine (EP-UNAG) is a precursor to UDP- N-acetylmuramate (UDP-MurNAc), an essential building block for the bacterial cell wall. MurA is conserved across both gram-positive and gram-negative bacterial species: gramnegative bacteria have one copy of the *murA* and gram-positive bacteria have two copies. *Alloiococcus otitidis mur*A was identified as described in Example 5/Table 4 and its genomic structure set forth in Seq. ID No. 101. The amino acid sequence of the protein encoded by this gene is set out in Seq. Id No. 102.

## Alloiococcus otitidis murA as a target for anti-infective development

MurA in E. coli and Streptococcus pneumoniae has been shown to be essential by gene deletion technique. The essentiality of MurA in gram-positive bacteria such as Streptococcus pneumoniae was demonstrated in that its deletion is fetal. No mammalian homolog to MurA has been reported. MurA is specifically inhibited by the natural product antibiotic fosfomycin. Thus the importance of MurA in peptidoglycan biosynthesis makes it an attractive target for the design of novel antibacterial agent.

#### 10 Assays for measuring MurA function

#### Phosphate detection:

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*Mur*A activity is detected by quantitating the UDP-GluNAc-dependent Pi from PEP and assayed by Lanzetta's malachite Green-ammonium molybdate assay. Pi is quantitated by measuring the optical density at A660 nm.

#### Coupled assay with MurB:

A coupled assay in access of MurB, which reduces the MurA product EP-UNAG G to UDP-MurNAc, couples the MurA transferase activity with NADPH oxidation. The oxidation of NADPH is monitored at 340 nm and is stoichometric with the production of EP-UNAG.

#### Fluorescence experiments

Fluorescence experiments to detect *murA* are performed using the hydrophobic fluorescence probe 8-anilino-1-naphthalene sulfonate (ANS). The fluorescence quenching of MurA/ANS solutions upon addition of UDP-GlcNAc or pyruvate-P is concentration dependent and in a saturating manner.

### Isothermal titration calorimetry

The binding of UDP-GluNAc to MurA is studied in the absence and presence of the antibiotic fosfomycin by isothermal titration calorimetry. Fosfomycin binds covalently to MurA in the presence of UDP-GluNAc and also in its absence as

demonstrated by MALDI mass spectrometry. Novel Fosfomycin analogs and other antibiotics that bind to *mur*A are also identifiable using isothermal titration chemistry.

## Capillary electrophoresis-based enzyme assay

A capillary electrophoresis-based enzyme assay for MurA is described by Dai and colleagues. This method, based on UV detection, provides baseline separation of one of the reaction products, EP-UNAG, from substrates PEP and UDP-GlcNAc within 4 min. The other product, phosphate, is not detectable by UV at 200 nm. Quantitation of individual components, substrates or product, is be accomplished based on the separated peaks. This assay is also used to detect novel antibiotics, which inhibit murA activity.

#### EXAMPLE 23

## ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYMES MURB

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MurB, the UDP-*N*-acetyl enolpyruvyl glucosamine reductase, commits the second step of bacterial cell wall biosynthesis in cytoplasm and is responsible for the reduction of the enol ether to the lactyl ether, utilizes one equiv. of NADPH and a solvent proton. The product of MurB is UDP-N-acetylmuramic acid (UDP-MurNAc), the linker of the peptide and glycan portions of cell wall precursor UDP muramyl-pentapeptide. MurB from *E. coli* is a 342 amino acid protein, which has a distinctive yellow color characteristic of bound flavin as its co-factor. The biochemistry characterization and X-ray crystal structure of MurB in *E. coli*, in *Staphylococcus aureus* and *Streptococcus pneumoniae* have been studied extensively. The gene Alloiococcus oitidis murB was identified as disclosed as described in Example 5, and is set out in Seq. ID No. 39. The amino acid sequence of the protein encoded by this gene is set out in Seq. ID No. 40.

## Alloiococcus oitidis murB as a target for anti-infective development

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The essentiality and unique function of MurB in prokaryotic cells and the absence of homologue in eukaryotic cells make it an attractive novel antibacterial target. To date, no small molecule inhibitors of MurB have been reported.

Alloiococcis oititidis ORF-1263 (*murB* ) (Seq. ID No. 39) encodes enzyme UDP-*N*-acetylenolpyruvylglucosamine Reductase (MurB) as shown by sequence homology.

# 5 Assays for measuring *MurB* activity Spectrophotometric assay monitoring NADPH consumption:

MurB activity is typically monitored by its biochemical reaction in which NADPH reduces the bound FAD and resulting decrease in absorbance at 340 nm. Enzyme is maximally activated in the presence of K+, NH<sup>4</sup> at cation concentrations between 10-50 mM.

#### Coupled assay with MurC:

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In designing an end point assay for high through put screen (HTS), a novel coupled assay in access of UDP-MurNAc L-alanine synthase (*Mur*C) was developed at Wyeth. This assay utilizes the biochemically synthesized *Mur*A product EP-UNAG as substrate, coupled with limited *Mur*B and excess *Mur*C in the reaction with all other substrates/components involved. In this assay, *Mur*B is responsible for the reduction of the enol ether to the lactyl ether, and the follow up enzyme *Mur*C catalyzes the ATP dependent ligation of the first of the five amino acids of UDP-peptapeptide with a release of one molecule of phosphate. After 60 minutes of incubation, color reagent malachite green was added and phosphate was detected spectrophotometrically.

### Fluorescence binding assay

A fluorescence method developed at Wyeth is used to determine the binding potency (Kd value), stoichiometry and nature of binding site of substrates and inhibitors interactions with *Mur*B enzymes. This assay is based on changes in intrinsic fluorescence of inhibitor and/or enzyme, upon formation of enzyme-inhibitor complex. Oxidized form of *Mur*B consists of two fluorescent groups, namely tryptophan residues and the cofactor FAD. Upon binding inhibitor or substrate, local changes in the solvent environment of these groups or overall conformational and electronic changes occur in the enzyme due to which the fluorescence emission is altered. For instance, inhibitor binding significantly quenched the fluorescence and

altered the solvent environment of FAD to a less polar environment. The changes in the fluorescence of the FAD moiety are used to estimate binding constants for *MurB* inhibitors. Binding experiments are set up in which a fixed concentration of enzyme is titrated with increasing concentrations of the inhibitor. In typical inhibitor binding experiments, the fluorescence emission of the FAD moiety is quenched due to specific interactions of the inhibitor with *MurB* enzymes and the binding site was saturated at micromolar concentrations of inhibitor. The changes in the fluorescence are fitted to mathematical binding models to determine binding affinity.

### 10 Temperature-jump isothermal denaturation procedure

Temperature-jump isothermal denaturation procedure with various methods of detection is used to evaluate the quality of putative inhibitors of *Mur*B discovered by high-throughput screening. Three optical methods of detection-ultraviolet hyperchromicity of absorbance, fluorescence of bound dyes, and circular dichroism-as well as differential scanning calorimetry are used to dissect the effects of two chemical compounds and a natural substrate on the enzyme. The kinetics of the denaturation process and binding of the compounds detected by quenching of flavin fluorescence are used to quantitate the dose dependencies of the ligand effects.

#### 20 NMR studies

NMR studies are performed using perdeuterated, uniformly 13C/15N-labeled samples of *Mu*rB. In the case of substrate-free *Mu*rB, one or more backbone atoms are assigned for 334 residues (96%). For NADP+-complexed *Mu*rB, one or more backbone atoms are assigned for 313 residues. The strategies used for obtaining resonance assignments are known. Localizing the NADP+ binding site on the MurB enzyme is also studied by NMR methodology.

#### EXAMPLE 25

## ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYME, MURC

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Uridine diphosphate-N-acetylmuramate:L-alanine ligase (MurC) catalyzes the third chemical step of bacterial cell wall biosynthesis. This enzyme is a nonribosomal peptide ligase which utilize ATP to form an amide bond between L-alanine and UDP-

N-acetylmuramic acid (UDP-MurNAc). This ATP-dependent ligation adds the first of five amino acids to the sugar moiety of the peptidoglycan precursor. Also, in this reaction, ATP is converted to ADP with release of one molecule of inorganic phosphate. Thus MurC reaction is an essential step in cell wall biosynthesis for both gram-positive and gram-negative bacteria. The genetic, biochemistry analysis and crystal graphic studies of MurC in gram-negative bacteria *E. coli* have been extensively studied. Characterizations of MurC in other pathogens such as *Staphylococcus aureus* and *Pseudomonas aeruginosa* have also been documented.

## 10 Alloiococcis otitidis encoded MurC as a target for anti-infective development

The Alloiococcis otitidis ORF-2602 (murC, Seq. ID No. 95) encodes enzyme UDP-MurNAc:L-alanine ligase (MurC) as determined by sequence homology. This enzyme presents a target for the development of novel anti-infectives to treat the disease(s) caused by this pathogen. Novel compounds identified using combinatorial chemistries are assayed for their inhibitory effect on MurC activity using one of the asssays set out below.

### Assays for measuring MurC activity

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## Spectrophotometric assay detecting phosphate release:

MurC activity is detected by the inorganic phosphate production. Typically the reaction mixture contains substrates ATP, L-alanine, UDP-MurNAc, DTT, MgCl<sub>2</sub> and MurC enzyme. After 20 minutes incubation, the reaction is quenched with the addition of malachite Green-ammonium molybdate for a colored reaction. Absorbance at 660 nm is read 5 minutes after the quench. Absorbance values are converted to concentration of Pi with standard curves using KH<sub>2</sub>PO<sub>4</sub>, which is prepared under identical conditions without the enzyme MurC.

## Spectrophotometric assay detecting formation of ADP

Due to the conversion of ATP to ADP in MurC reaction, the production of ADP is monitored in coupled enzymes spectrophotometrically. In this reaction, in addition to MurC substrate UDP-MurNAc, L-alanine and ATP, NADH, phosphoenolpyruvate, MgCl<sub>2</sub> and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, two other coupled enzymes pyruvate

kinase and lactase dehydrogenase are also presented. Reaction mixtures without ATP and MurC are incubated at 37°C for 10 min before ATP is added for another minute. Reaction is then started by the addition of MurC. The decrease of NADH absorbance at 340 nm is monitored spectrophotometrically. One unit of activity corresponds to 1 umol of ADP formed per hour.

#### L-Alanine radio-labeled assay:

The MurC enzyme activity in this assay is measured as endpoint using <sup>14</sup>C-L-alanine and ATP incubated with MgCl<sub>2</sub>, and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> in 100 mM Tris/HCl, pH 8.0. Reaction is initiated by the addition of the catalytic amounts of MurC. Samples of the reaction mixture are then mixed with glacial acetic acid and then stored at 4°C. Remaining <sup>14</sup>C -L-alanine is separated from <sup>14</sup>C -UDPMurNAc on SCX columns run under vacuum. Quenched reaction samples are supplemented with equilibration buffer and counted using a liquid scintillation counter.

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#### EXAMPLE 26

## ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYMES MURD

Bacterial UDP-N-acetylmuramyl-L-alanine:D-glutamate ligase (MurD), a cytoplasmic peptidoglycan biosynthetic enzyme, catalyzes the fourth step of bacterial cell wall biosynthesis. In this reaction, MurD catalyzes ATP-dependent addition of D-glutamate to an alanyl residue of the UDP-N-acetylmuramyl-L-alanine (UDP-MurNAc-L-Ala) precursor, generating the UDP-MurNAc-dipeptide. The formation of a peptide linkage between the amino function of D-glutamate and the carboxy terminius of UDP-N-acetylmuramuamyl-L-alanine is generated through this reaction. The stoichiometric consumption of ATP supplies the energy needed for this peptide bond formation with concomitant generation of ADP and orthophosphate. The murD genes were cloned and characterized from gram-positive bacteria of Staphylococcus aureus and Streptococcus pyogenes, and gram-negative bacteria from Escherichia coli, Haemophilus influenzae, Bacillus subtilis. Structures of MurD from E. coli and MurD complexed with its substrate UDP-MurNAc-L-Ala have been solved to 2.0 A resolution. The role of specific amino acids at the active site of MurD have been extensively studied using the ortholog and paralog amino acid invariants. Homologue

of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 89). The protein encoded by the gene is set forth in Seq. ID No. 90.

## Alloiococcus otitidis encoded MurD as a target for anti-infective development

Due to its high specificity and essentiality, MurD is an attractive target for the development of novel antimicrobial agents. *Alloiococcis otitidis* ORF-2494, by sequence homology, has been shown to encode enzyme UDP-N-acetylmuramyl-L-alanine:D-glutamate ligase (MurD) (Seq. ID. No. 89). Inhibition of MurD activity is used to identify novel antimicrobial agents.

## Assays for measuring MurD activity

## Spectrophotometric assay detecting phosphate release:

MurD activity in the presence or absence of a putative inhibitory molecule of MurD is detected by the orthophosphate production in test tube or in 96-well format. Typically the reaction mixture contains substrates ATP, D-glutamine, UDP-MurNAc-L-Ala, DTT, MgCl2 and MurD enzyme. After 20 minutes incubation, the reaction is quenched with the addition of malachite Green-ammonium molybdate for a colored reaction. Absorbance at 660 nm is read 5 minutes after the quench using Molecular Devices SpectraMax 250 plate reader. Absorbance values are converted to concentration of Pi using orthophosphate standards, which are prepared under identical conditions without the enzyme MurD.

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# Spectrophotometric assay for detecting formation of ADP in the presence or absence of a putative inhibitory mollecule of MurD:

Due to the conversion of ATP to ADP in MurD reaction, the production of ADP is monitored with coupled enzymes of pyruvate kinase and lactase dehydrogenase spectrophotometrically. In this reaction, in addition to MurD substrate UDP-MurNAc-L-ala and ATP, MgCl<sub>2</sub> and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, there is also in significant access of NADH, phosphoenolpyruvate, and two coupled enzymes pyruvate kinase and lactase dehydrogenase. This protocol monitors ADP formation

in the MurD catalyzed reaction, in the presence or absence of a putative inhibitory mollecule of MurD, by the decrease of NADH absorbance at 340 nm.

### L-Glutamate radio-labeled assay:

The MurD enzyme activity in the presence or absence of putative inhibitors of MurD is also measurable using D-14C- glutamate as an endpoint assay. The reaction mixture contains D-14C- glutamate UDP-MurNAc-L-Ala, ATP, MgCl<sub>2</sub>, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> in 100 mM Tris/HCl, pH 8.0. An HPLC assay with online UV and flow scintillation detects the formation of UDP-MurNAc-L-Ala-D-14C Glu and ADP in each reaction.

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#### **EXAMPLE 27**

## ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYME, MURE

The fifth step in the cytoplasmic peptidoglycan biosynthetic is catalyzed by MurE. In this step, the monomer units in the *Escherichia coli* and *Staphylococcus aureus* cell wall peptidoglycans differ in the nature of the third amino acid in the L-alanyl-gamma-D-glutamyl-X-D-alanyl-D-alanine side chain, where X is meso-diaminopimelic acid or L-lysine, respectively. Therefore, MurE from *E. coli* is the UDP-N-acetylmuramoyl-L-alanyl-D-glutamate: meso-diaminopimelic acid ligase, and MurE from *S. aureus* is the UDP-N-acetylmuramoyl-L-alanyl-D-glutamate: L-lysine ligase. Thus represents the major difference of MurE from other murein enzymes in cytoplasm. The amino acid residues catalyzed by MurE plays a key role in the integrity of sacculus since it is directly involved in the peptide cross-linkage. MurE reaction is also ATP-dependent, which supplies the energy needed for the peptide bond formation with concomitant generation of ADP and orthophosphate.

The essentiality of *MurE* has been well documented in *E. coli*, in *S. aureus*, as well as other pathogens such as *Haemophilis influenzae*, *Vibrio cholerae* and *Corynebacterium glutamicum*. Gene *murE* has been shown to be essential in bacteria. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 25). The protein encoded by the gene is set forth in Seq. ID No. 26.

Alloiococcus otitidis MurE as a target for anti-infective development

Alloiococcis otitidis ORF-851, by sequence homology encodes enzyme UDP-N-acetylmuramyl-L-alanine-D-glutamate ligase: meso-diaminopimelic acid/or L-Lysine (MurE) (Seq. ID No 25). MurE activity in the presence or absence of a putative inhibitory molecule of MurE activity is used to identify novel antimicrobial I agents, which may be used ti treat disease caused by Alloiococcis otitidis.

## Assays for measuring MurE activity

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## Radio labeled substrate assay: meso-A2pm-adding activity

Activity of MurE from *Alloiococcis otitidis* in the presence or absence of a putative inhibitory molecule of MurE activity is measured by using radio-labeled meso-<sup>14</sup>C A2pm mixing with ATP, MgCl<sub>2</sub> UDP-MurNAc-L-Ala-D-Glu, DTT in 100 mM Tris/HCl and MurE from *Alloiococcis otitidis*.

## 15 Radio labeled substrate assay: L-lysine adding activity

Activity of MurE from *Alloiococcis otitidis* in the presence or absence of a putative inhibitory molecule of MurE activity is measured by using radio-labeled UDP-MurNAc-L-Ala-D-14C-Glu mixing with ATP, MgCl<sub>2</sub>, DTT, L-lysine in 100 mM Tris/HCl and MurE from *Alloiococcis otitidis*.

In both cases, mixtures are incubated at 37°C for 30 min, and reactions stopped by the addition of acetic acid. Reaction product is separated by high votage electrophoresis in 2% formic acid for 45 min. The radio active spots corresponding to substrate and reaction product are detected by overnight autoradiography, or with radio scanner. The spots are also cut out and counted using liquid scintillation counter.

#### Example 28

## ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYME, MURF

The D-alanyl-D-alanine-adding enzyme MurF encoded by the *murF* gene catalyzes is the last step of the cytoplasmic peptidoglycan biosynthesis. MurF performs the ATP-dependent formation of UDP-N-acetylmuramyl-L-gamma-D-Glumeso-diaminopimelyl-D-Ala-D-Ala (UDP-MurNAc-pentapeptide). The product of MurF, UDP-MurNAc pendapeptide, is the final product of the cytoplasm enzymes and

is the most important precusor for further peptidoglycan biosynthesis. UDP-MurNAc pendapeptide is then catalyzed by the plasma membrane bound enzymes such as the translocase MraY and transferase MurG. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 3). The protein encoded by the gene is set forth in Seq. ID No. 4.

## Alloiococcus otitidis MurF as a target for anti-infective development

Due to its high specificity, essentiality, and importance of its product UDP-MurNAc pentapeptide, MurF is attractive as an antibacterial target. The *Alloiococcis otitidis* ORF-48, by sequence homology,encodes enzyme UDP-N-acetylmuramyl-L-alanine-D-glutamate ligase: meso-diaminopimelic acid/or L-Lysine -alanyl-D-alanine-adding enzyme (MurF) (Seq. ID No. 3). MurF activity in the presence or absence of a putative inhibitory molecule of MurF activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by *Alloiococcis otitidis*.

Assays for measuring MurF activity

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## Spectrophotometric assay detecting phosphate release:

Activity of MurF from *Alloiococcis otitidis* in the presence or absence of a putative inhibitory molecule of MurF activity is detected by the inorganic phosphate release in the ATP dependent MurF reaction. This assay detects nonomole amount of Pi in the reaction mixture contains substrates ATP, D-ala-D-ala, UDP-MurNActripeptide, DTT, MgCl<sub>2</sub> and MurF enzyme. After 5 minutes incubation, the reaction is quenched with the addition of malachite Green-ammonium molybdate for a colored reaction.

## Coupled spectrophotometric assay detecting formation of ADP

Due to the conversion of ATP to ADP in MurF reaction, the production of ADP in the presence or absence of a putative inhibitory molecule of MurF activity, is monitored with coupled enzymes of pyruvate kinase and lactase dehydrogenase spectrophotometrically. In this reaction, the decrease at 340 nm is observed as NADP is consumed in MurF reaction process. The reaction typically contains tris

buffer, substrates ATP, D-ala-D-ala, UDP-MurNAc-tripeptide, DTT, MgCl<sub>2</sub>, phosphoenopyruvate, NADPH and MurF enzyme.

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#### **EXAMPLE 29**

### ALLOIOCOCCUS OTITIDIS ENCODED CELL WALL BIOSYNTHETIC ENZYME, MURG

MurG, the last enzyme involved in the intracellular phase of peptidoglycan synthesis, is a membrane-associated glycosyltransferase. MurG catalyzes the transfer of *N*-acetyl glucosamine from UDP to the C4 hydroxyl of a lipid-linked N-acetyl muramic acid derivative (lipid I) to form lipid II. Lipid II is a linked disaccharide that is the minimal subunit of peptidoglycan. Once lipid II is formed, this disaccharide is translocated across the bacterial membrane where it is polymerized and cross-linked to form the peptidoglycan layers. MurG has been shown to be essential for bacterial survival. The inactivation of MurG gene rapidly inhibits peptidoglycan synthesis in exponential growing cells. As a result, various alterations of cell shape are observed, and cell lysis finally occurs. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 87). The protein encoded by the gene is set forth in Seq. ID No. 88.

#### Allojococcus otitidis MurG as a target for anti-infective development

MurG is shown to be associated with the inner face of cytoplasmic membrane, and establishing that the entire peptidoglycan monomer unit assembled before being transferred across the membrane. MurG is a key enzyme at the border line between cytoplasmic and membrane of pepdidoglycan synthesis, thus makes it an attractive target for novel antibacterial agent. Further, no mammalian analogues of MurG have been identified. Due to its high specificity, essentiality, and importance, MurG is attractive as an antibacterial target.

The Alloiococcis otitidis ORF-2492 has been shown to encode, by sequence homology, glycosyltransferase (MurG) (Seq. ID No.....). MurG activity in the presence or absence of a putative inhibitory molecule of MurG activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by Alloiococcis otitidis.

## Assays for measuring MurG function

### Radiolabeled reaction

Activity of MurG from *Alloiococcis otitidis* in the presence or absence of a putative inhibitory molecule of MurG activity is measured by using <sup>14</sup>C labeled *N*-UDP-GluNAc in the reaction containing UDP-MurNAc-pentapeptide, MgCl<sub>2</sub>, ATP and MurG protein. The reaction is stopped after 30 min incubation and by boiling for 3 min. The reaction mixtures are applied to a Whatman I filter paper and subject to descending chromatography overnight. Radioactivity is located and countered with a scanner. This assay is also used to identify the specificity of inhibitor of MraY or MurG, based on the detection of radiolabeled <sup>14</sup>C GluNAc incorporated into membrane precursors.

## Fluorometric assay

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Based on the decrease in NADPH fluorescence at 465 nm, MurG reaction is also monitored in a reaction mixture of HEPES buffer, MgCl<sub>2</sub>, Triton, phosphoenolpyruvate, and coupled enzymes of lactic dehydrogenase and pyruvate kinase, UDP-GluNAc and synthesized lipid I analogue in the presence or absence of putative inhibitors of MurG activity. One micromolar UDP corresponds to 500-fluorescence unit under the instrument setting.

# EXAMPLE 30 ALLOIOCOCCUS OTITIDIS ENCODED BY HMG COA REDUCTASE (MVAA)

Two pathways for isopentenyl diphosphate (IPP) synthesis have been described in bacteria: the mevalonate pathway and the non-mevalonate (MEP or GAP-pyruvate) pathway. The mevalonate pathway predominates in the archaebacteria, gram-positive organisms, yeast and mammals; whereas the MEP pathway is found in gram-negative organisms, *B. subtilis*, chlamydia, and mycobacterium. The first HMG CoA reductase gene to be sequenced was cloned from *P. mevalonii*, in which HMG CoA reductase permits growth on mevalonate as a sole carbon source. A number of genes of the mevalonate pathway were identified in *S. aureus*, *S. epidermidis*, *S. pyogenes*, *S. pneumoniae*, *E. faecalis* and *E. faecium*. One of the genes, which encodes for HMG-CoA reductase (*mvaA*), when deleted

severely attenuated for virulence in a mouse model indicating that *mvaA* is essential. Due to its high specificity, essentiality, and importance, *mvaA* is attractive as an antibacterial target. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 37). The protein encoded by the gene is set forth in Seq. ID No. 38.

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## HMG-CoA reductase (MvaA) as a target for anti-infective development

The Alloiococcis otitidis ORF- has been shown to encode, by sequence homology, HMG-CoA reductase (mvaA) (Seq. ID No 37). MvaA activity in the presence or absence of a putative inhibitory molecule of HMG-CoA reductase (mvaA) activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by Alloiococcus otitidis.

## Assays for measuring HMG-CoA reductase (mvaA) activity

MvaA is purified by standard methods using widely available molecular tags following expression at high level from *E. coli*. Enzymatic activity is monitored in the presence or absence of a putative inhibitory molecule of HMG-CoA reductase activity by following oxidation of NADPH to NADP spectrophotometrically at 340 nm. The assay is carried out in the following buffer: 0.25 mM NADPH, 0.25 mM HMG-CoA, 50 mM NaCl, 1 mM EDTA, 5 mM DTT, 25 mM KH<sub>2</sub>PO<sub>4</sub> (pH 7.5). The assay is amenable to HTS in high density screening microtiter plates.

Forward reaction: Activity of HMG-CoA reductase (mvaA) from Alloiococcus otitidis in the presence or absence of a putative inhibitory molecule of HMG-CoA reductase activity is measured by reductive deacylation of HMG-CoA to mevalonate as measured the consumption of NADPH to NADP. Unlike other class II HMG Coa reductases, MvaA from Alloiococcus otitidis, like S. aureus, can use either NADPH or NADH cofactor in the reaction. The following kinetic data describe the reaction:  $K_{m(HMG CoA)} = 40 \ \mu M$ ,  $K_{m(NADPH)} = 70 \ \mu M$ ,  $K_{m(NADP)} = 100 \ \mu M$  (12). This assay is inhibitable by the statin drug fluvastatin; the  $K_i$  was measured at 320  $\mu M$ , which is four orders of magnitude higher than the  $K_i$  for class I HMG-Coa reductases.

Reverse reaction: The oxidative acylation of mevalonate to HMG-CoA in the presence or absence of a putative inhibitory molecule of HMG-CoA reductase activity is also monitored. The following kinetic data describes the reaction:  $K_{m(mevalonate)} = 670 \, \mu M$ ,  $K_{m(CoASH)} = 390 \, \mu M$ ,  $K_{m(NADP)} = 580 \, \mu M$  (12).

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#### **EXAMPLE 31**

### ALLOIOCOCCUS OTITIDIS ENCODED DIPHOSPHOMEVALONATE DECARBOXYLASE (MVAD)

Diphosphomevalonate decarboxylase, encoded by *mvaD*, the final enzyme acting in the mevalonate pathway of IPP synthesis was cloned from *S. aureus* by Wilding *et al* in 2000. Insertional inactivation of *mvaD* could only be accomplished when the strains were supplemented with mevalonate, indicating that *mvaD* is essential. The final step of the mevalonate pathway leading to IPP is the decarboxylation and dehydration of mevalonate-5-pyrophosphate to form isopentenyl diphosphate by MvaD (diphosphomevalonate decarboxylase).

MvaD homologues are well represented in gram-positive organisms (10). Phylogenetic analysis revealed that the cluster of gram-positive enzymes (39-80% identity) were well separated from the eukaryotic homologues, suggesting utility as an antibacterial target. The *Alloiococcis otitidis* ORF- 1275b has been shown to encode, by sequence homology, diphosphomevalonate decarboxylase (MvaD) (Seq. ID No. 43). MvaD activity in the presence or absence of a putative inhibitory molecule of diphosphomevalonate decarboxylase (MvaD) activity is used to identify novel antimicrobial agents, which may be used to treat the disease(s) caused by *Alloiococcus otitidis*. The protein encoded by the gene is set forth in Seq. ID No. 44.

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# Example 32 ALLOIOCOCCUS OTITIDIS ENCODED HMG COA SYNTHASE (MVAS)

The second step of the mevalonate pathway leading to IPP is the irreversible condensation of acetoacetyl-CoA and acetyl-CoA to form HMG-CoA by MvaS (HMG CoA synthase). It has been shown that *mvaS* knockout mutant of *S. pneumoniae* was attenuated for virulence. Due to its high specificity, essentiality, and importance, *mvaS* is attractive as an antibacterial target. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 35). The protein encoded by the gene is set forth in Seq. ID No. 36.

## HMG COA SYNTHASE (MVAS) AS A TARGET FOR ANTI-INFECTIVE DEVELOPMENT

The Alloiococcis otitidis ORF- has been shown to encode, by sequence
homology, MvaS (HMG CoA synthase) (Seq. ID No. 35). MvaS activity in the
presence or absence of a putative inhibitory molecule of HMG-CoA synthase (mvaS)
activity is used to identify novel antimicrobial agents, which may be used to treat
disease caused by Alloiococcus otitidis.

## Assays for measuring MvaS function

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MvaS is purified by standard methods using widely available molecular tags following expression at high level from *E. coli*. HMG-CoA synthase activity in the presence or absence of a putative inhibitory molecule of HMG-CoA synthase (mvaS) is assayed by measuring the loss of the enolate form of acetoacetyl-CoA spectrophotometrically. The reaction is carried out in a buffer containing 50 mM Tris (pH 9.75), 5.0 mM MgCl<sub>2</sub>, 500  $\mu$ M acetyl-CoA, 20  $\mu$ M acetoacetyl-CoA and enzyme. The enolate formed is monitored at 302 nm; therefore, as the acetoacetyl-CoA is consumed the signal is depleted. Using this assay the following kinetic data is measured:  $K_{m(acetyl-CoA)} = 350 \ \mu$ M;  $K_{m}^{app}_{(acetoacetyl-CoA)} = 10 \ \mu$ M. This assay is amenable to HTS in high- high density screening microtiter plates.

#### Example 33

# ALLOIOCOCCUS OTITIDIS ENCODED NICOTINAMIDE ADENINE DINUCLEOTIDE ADENYLYL TRANSFERASE (NADD)

Nicotinamide adenine dinucleotide (NAD) is an essential molecule in all living cells. NAD is synthesized via a multi-step *de novo* pathway or via a pyridine salvage pathway. The enzyme nicotinic acid mononucleotide adenylyl transferase (NaMN AT, EC2.7.7.18) catalyzes the conversion of ATP and nicotinic acid mononucleotide (NaMN) to nicotinic acid adenine dinucleotide (NaAD). The *nadD* gene, encoding bacterial NaMN AT, is essential for NAD biosynthesis and bacterial cell survival. NadD contains well-conserved the nucleotidyl transferase consensus sequence (GXFXXXHXGH). The adenylyl transferase encoded by the *nadD* gene prefers NaMN over nicotinomide mononucleotide (NMN) as substrate. Due to its high specificity, essentiality, and importance, *nadD* is attractive as an antibacterial target. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 91). The protein encoded by the gene is set forth in Seq. ID No. 92.

## NICOTINAMIDE ADENINE DINUCLEOTIDE ADENYLYL TRANSFERASE (NADD) AS A TARGET FOR ANTI-INFECTIVE DEVELOPMENT

The Alloiococcis otitidis ORF- has been shown to encode, by sequence homology, niotinomide adenine dinucleotide adenyl transferase (NadD) (Seq. ID No. 91). NadD activity in the presence or absence of a putative inhibitory molecule of NadD activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by Alloiococcus otitidis.

## Assays for measuring NadD function Discontinuous assay

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NadD activity in *Alloiococcus otitidis* is measured in the presence or absence of a putative inhibitory molecule of NadD activity. NadD converts nicotinic acid mononucleotide (NaMN) and adenosine triphosphate (ATP) to nicotinic acid dinucleotide (NaAD) and pyrophosphate (PP<sub>i</sub>). Each PP<sub>i</sub> molecule produced by the NadD reaction is then converted to two phosphate

 $(P_i)$  molecules in the presence of inorganic pyrophosphatase (PPase). The  $P_i$  molecules present are quantitated with a malachite green reagent at 660 nm.

HPLC-based assay: Enzyme activity is measured by HPLC quantitation of the reaction products. A neutralized aliquots from the reaction described above was injected into an HPLC system utilizing a 250 x4.6 mm Supelcosil LC-18 5μm reversed-phase column. The elution conditions: 9 min at 100% buffer A (0.1 M potassium phosphate buffer, pH6.0,6 min at up to 12% buffer B (buffer a, containing 20% methanol, 2.5 min at up to 45% buffer B, 2.5 min at up to 100% buffer B, and hold at 100% buffer B for 5.5 min. The eluate absorbance was monitored at 254 nm.

#### Continuous assay

In bacteria, NadD combines nicotinic acid mononucleotide (NaMN) and adenosine triphosphate (ATP) to form nicotinic acid adenine dinucleotide (NaAD). NadE then converts NaAD into nicotinamide adenine dinucleotide (NAD) in the presence of ammonia and ATP. In the assay, the NAD product is reduced to NADH with alcohol dehydrogenase (ADH) and ethanol, thus permitting direct spectrometric detection of NADH at 340 nm wavelength. The coupled reaction above also includes inorganic pyrophosphatase (PPase) to prevent accumulation of the pyrophosphate byproduct from the consumption of ATP.

#### **EXAMPLE 34**

## ALLOIOCOCCUS OTITIDIS ENCODED NICOTINAMIDE ADENINE DINUCLEOTIDE SYNTHASE (NADE)

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NAD is a central compound in cellular metabolism. The final metabolic step in the pathway is conversion of nicotinamide adenine dinucleotide – product of NadD reaction – to NAD, a step catalyzed by the enzyme NAD synthetase (NadE). NaMN – substrate for NadD – can be formed by three different enzymatic reactions: in the *de novo* pathway from quinolinate, in Preiss-Handler salvage pathway from nicotinic acid, and in the nucleoside salvage pathway by deamindation of nicotinamide mononucleotide. In bacteria, there are no known alternatives for the metabolic steps between NaMN and NAD. Mutants blocked in these steps cannot be recovered as auxotrophs since the required metabolites are not taken up by cells. In

the bacterial cells, the second substrate for NadE is ammonium, as opposed to glutamine for eukaryotes. NadE is an essential and conserved protein in the eubacterial nicotinamide adenine dinucleotide (NAD) biosynthesis pathway. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 49). The protein encoded by the gene is set forth in Seq. ID No. 50.

## Assays for measuring NadE function:

10 The Alloiococcis otitidis ORF- has been shown to encode, by sequence homology, niotinomide adenine dinucleotide adenyl synthase (NadE) (Seq. ID No. 49). NadE activity in the presence or absence of a putative inhibitory molecule of NadE activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by Alloiococcus otitidis.

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### **DISCONTINUOUS ASSAY:**

In assay, NadE converts nicotinic acid adenine dinucleotide (NaAD) into nicotinamide adenine dinucleotide (NAD) in the presence of ammonia and ATP. Each PP<sub>i</sub> molecule produced by the NadE reaction can then be converted to two phosphate (P<sub>i</sub>) molecules in the presence of inorganic pyrophosphatase (PPase). The P<sub>i</sub> molecules present can then be quantitated with a malachite green reagent at 660 nm.

#### HPLC-based assay:

Enzyme activity can be measured by HPLC quantitation of the reaction products. A neutralized aliquots from the reaction described above was injected into an HPLC system utilizing a 250 x4.6 mm Supelcosil LC-18 5µm reversed-phase column. The elution conditions: 9 min at 100% buffer A (0.1 M potassium phosphate buffer, pH6.0,6 min at up to 12% buffer B (buffer a, containing 20% methanol, 2.5 min at up to 45% buffer B, 2.5 min at up to 100% buffer B, and hold at 100% buffer B for 5.5 min. The eluate absorbance was monitored at 254 nm (1).

#### Continuous assay:

Coupled NadD-NadE assay. NadD and NadE can be detected in one continuous coupled assay. In first reaction, NadD combines nicotinic acid mononucleotide (NaMN) and adenosine triphosphate (ATP) to form nicotinic acid adenine dinucleotide (NaAD). NadE then converts NaAD into nicotinamide adenine dinucleotide (NAD) in the presence of ammonia and ATP. In the assay, the NAD product is reduced to NADH with alcohol dehydrogenase (ADH) and ethanol, thus permitting direct spectrometric detection of NADH at 340 nm wavelength. The coupled reaction above also includes inorganic pyrophosphatase (PPase) to prevent accumulation of the pyrophosphate byproduct from the consumption of ATP (this method can be use as HTS format).

NadE assay. In assay, NadE converts NaAD into nicotinamide adenine dinucleotide (NAD) in the presence of ammonia and ATP. The NAD product is reduced to NADH with alcohol dehydrogenase (ADH) and ethanol, thus permitting direct spectrometric detection of NADH at 340 nm wavelength. The reaction above also includes inorganic pyrophosphatase (PPase) to prevent accumulation of the pyrophosphate byproduct from the consumption of ATP (this method can be use as HTS format).

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#### EXAMPLE 35.

## ALLOIOCOCCUS OTITIDIS ENCODED PUTATIVE MEMBRANE PROTEIN NORA

An efflux transporter NorA that was originally identified in *Staphylococcus* aureus belongs to the family of multidrug resistance (MDR) transporters. NorA is encoded by chromosomally-located *norA* gene, it has broad substrate specificity and mediates resistance to various lipophilic and monocationic compounds such as ethidium bromide (EtBr), cetrimide, benzalkonium chloride, rhodamine 6G, tetraphenylphosphonium (TPP), chloramphenicol as well as some hygrophilic quinolones such as norfloxacin, ciprofloxacin and oxafloxacin. Increased levels of *norA* expression are associated with single nucleotide changes upstream of *norA* in a putative promoter/operator region and lead to increased pleiotropic resistance. NorA is a putative membrane protein with 12 predicted membrane-spanning domains and is classified as a member of major facilitator superfamily (MFS), a subgroup of MDR

transporters characterized by the presence of 12-14 transmembrane segments and the use of proton motive force as an energy source for drug efflux. NorA homologs that belong to MFS family include Bmr and Blt of *Bacillus sub*tilis, EmeA of *Enterococcus faecalis* and PmrA of *Streptococcus pneumonia*. The expression of *bmr* gene in *B. subtilis* is upregulated by the product of adjacent *bmR* gene in the presence of inducers (rhodamine 6G and TPP), and there is an evidence that expression of *norA* in *S. aureus* is regulated by AlrS-AlrR two-component regulatory system.

It remains unknown whether the efflux of various toxins is a primary function of NorA. When overexpressed in E. coli, norA produces resistance to a broad range of substrates including fluoroquinolones. Everted membrane vesicles prepared from norA-expressing E. coli exhibit energy-dependent transport of norfloxacin, the transfer is abolished by cyanide m-chlorophenylhydrazone (CCCP) and nigericin but not by valinomycin indicating that NorA-mediated transfer is coupled to the proton gradient of cell membrane. Norfloxacin uptake in everted vesicles as well as NorAassociated resistance phenotype is inhibited by reserpine and verapamil that also inhibit other MDR transporters and are toxic to mammalian cells. Histidine-tagged NorA (NorA-His) was recently overexpressed and purified from E. coli, reconstituted into both everted membrane vesicles and proteoliposomes and was shown to function as a self-sufficient efflux pump using fluorescent dye Hoechst 33342. Due to its high specificity, essentiality, and importance, norA is attractive as an antibacterial target. Homologue of this gene identified in Alloiococcus otitidis is described in Example 5/Table 4 (Seq. ID No 67). The protein encoded by the gene is set forth in Seq. ID No. 68.

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#### NORA AS A TARGET FOR ANTI-INFECTIVE DEVELOPMENT

The *Alloiococcis otitidis ORF*- has been shown to encode, by sequence homology, NorA (Seq. ID No. 67). NorA activity in the presence or absence of a putative inhibitory molecule of NorA activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by *Alloiococcus otitidis*.. Because of broad substrate specificity of NorA, NorA inhibitors should be particularly useful against pathogens that possess multiple drug resistance.

Whole-cell high-throughput screen (HTS) assay that measures NorA activity in the presence or absence of a putative inhibitory molecule of Alloiococcis otitidis NorA activity is used to identify potential inhibitors of NorA activity. The assay utilizes B. subtilis strain (AANA) that has both Bmr and Blt genetically inactivated while Alloiococcis otitidis NorA is supplied on the plasmid expression vector. The screen is based on the reversing of the resistance of AANA to EtBr. The exponentially growing cells are inoculated into the wells of a 96-well plate to OD<sub>600</sub>=0.001, the compounds are added at 20 µg/ml and EtBr is added at 10 µg/ml. Plates are incubated for 18 hrs at 37°C and examined for growth. Compounds that inhibit growth are subsequently tested in the presence/absence of EtBr for toxicity and effectivity. The efflux of EtBr from cells is monitored as described previously. The exponentially growing cells are loaded with EtBr at a concentration of 10 Dg/ml for 20 min at 37°C in the presence of reserpine (20 □g/ml). Cells are centrifuged, resuspended to an OD<sub>600</sub>=0.2 in a minimal medium GM1 alone or in the presence of inhibitor compound. Fluorescence of EtBr is monitored on a fluorimeter at an excitation 

of 530 nm and emission of 600 nm..

#### MONITORING OF HOECHST 33342 EFFLUX

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The efflux of fluorescent dye Hoechst 33342 from either everted membrane vesicles prepared from *Alloiococcus otitidis* His-NorA overexpressing *E. coli* or a proteoliposomes reconstituted with *Alloiococcus otitidis* His-NorA is also used to monitor NorA activity in the presence or absence of putative inhibitors of NorA. Everted membrane vesicles are diluted into 2 ml of 50 mM potassium HEPES (pH 7.2), 8.5 mM NaCl, 2 mM magnesium sulfate at a final protein concentration of 40 µg/ml. NorA is activated by the addition of either 0.5 mM lactate or 0.1 mM Mg<sup>2+</sup>-ATP. Hoechst 33342 is used in a range of 12.5 to 200 nM. Inhibitors are added at various concentrations prior to the addition of Hoechst 33342. Fluorescence change is monitored at excitation and emission wavelenghths of 355 and 457 nm respectively in a FluoroMax spectrofluorimeter. For proteoliposome assay, the His-NorA proteoliposomes are diluted into a cuvette containing 2 ml of 20 mM potassium phosphate, 50 mM potassium sulfate, 2 mM magnesium sulfate (pH 7.0) at a protein concentration of 10 µg/ml. The inhibitor compounds and Hoechst 33342 are added at various concentrations and the fluorescence is measured as described previously.

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# EXAMPLE 36 ALLOIOCOCCUS OTITIDIS ENCODED OBG GTPASE

The *obg* gene is the second gene in a two-gene operon along with the stage-O sporulation gene *spoOB* in *B. subtilis*. SpoOB is central to the phospho-relay signal cascade that initiates sporulation. Obg is a member of the GTPase superfamily by virtue of homology throughout a small portion of the protein that in other members of the family is responsible for nucleotide (GTP/GDP) binding. Obg is essential for growth. Initiation of sporulation is thought to be triggered by changes in the GTP content of the cell; therefore, the presence of a GTP binding protein in an operon with a central player in the process is suggestive of a role for Obg in sensing GTP levels and transmitting a signal to SpoOB.

It has been shown that Obg is involved in activation of the  $\sigma^B$  transcription factor in *B. subtilis* in response to environmental stress. Cells were depleted of Obg utilizing a construct that put *obg* under the control of an inducible ( $P_{lac}$ ) promoter. Depletion of IPTG resulted in bacteria that failed to activate  $\sigma^B$ . These studies further showed by yeast-two-hybrid analysis that Obg interacted with several known  $\sigma^B$  regulators, the so-called Rsb proteins.

The role Obg plays in transmitting signals important for sporulation and activation of the stress sigma factor may be indicative of the activities that small GTP binding proteins carry out in triggering cell division in response to GTP levels. Due to its high specificity, essentiality, and importance, obg is attractive as an antibacterial target. Homologue of this gene identified in *Alloiococcus otitidis* is described in Example 5/Table 4 (Seq. ID No 71). The protein encoded by the gene is set forth in Seq. ID No. 72.

#### **OBG AS A TARGET FOR ANTI-INFECTIVE DEVELOPMENT**

Obg is essential for bacterial viability. Conditional lethal alleles revealed that Obg is required for early events in sporulation and is involved in transmitting signals require for activation of the stress sigma factor. The *Alloiococcis otitidis ORF*- has been shown to encode, by sequence homology, obg (Seq. ID No.71). Obg activity in the presence or absence of a putative inhibitory molecule of Obg activity is used to

identify novel antimicrobial agents, which may be used to treat disease caused by Alloiococcus otitidis..

#### **Nucleotide binding**

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Obg binding to nucleotide in the presence or absence of putative antimicrobials, which inhibit Obg activity, is monitored by a simple filter-binding assay. *Alloiococcus otitidis* Obg (1-5 μg) is incubated with α<sup>32</sup>P-GTP (0.2 μCi) in a buffer consisting of 50 mM Tris (pH 8.5), 1.5 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 200 mM KCl, 10% glycerol for 30 minutes to 3 hours at 37°C. A portion of the reaction mix is spotted on nitrocellulose membrane, washed (50 mM Tris (pH 8.5), 1.5 mM MgCl<sub>2</sub>, 1 mM DTT) and dried. The membrane is then exposed to X-ray film. Alternatively, the spots are excised and counted. This assay is directly amenable to HTS using filter plates.

#### **GTPase activity**

The GTP hydrolytic activity of Obg is monitored using thin-layer chromatography (1, 2, 10). Obg and  $\alpha^{32}$ P-GTP are incubated in 50 mM Tris (pH 8.5), 1.55 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 200 mM KCl, 10% glycerol for 30 minutes at 37°C. An aliquot of the reaction is placed on PEI cellulose and the strip developed with 0.5 M KH<sub>2</sub>PO<sub>4</sub>, 1.0 M NaCl (pH 3.7). The spots conforming to GDP and GTP are identified by UV shadowing, excised and counted.

Alternatively, the hydrolysis of  $\gamma^{32}$ P-GTP is monitored by assaying for liberated P<sub>i</sub> (12). Obg and  $\alpha^{32}$ P-GTP are incubated in 50 mM Tris (pH 8.5), 1.5 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 100 mM KCl, 10% glycerol for 30 minutes to 3 hours at 37°C. The reaction is stopped by the addition of a slurry of charcoal in 1 mM Kpi (pH 7.5), which selectively binds the GTP and GDP. The liberated P<sub>i</sub> in the supernatant is monitored by Cerenkov counting. Free P<sub>i</sub> is also monitored with the Malachite Green reagent.

## Autophosphorylation

Obg autophosphorylation is monitored by incubating Obg with  $\gamma^{32}$ P-GTP in 50 mM Tris (pH 8.5), 1.5 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 100 mM KCl, 10% glycerol for 30

minutes at 37°C. Samples are analyzed following separation on SDS polyacrylamide gels, drying the gel and exposure to film.

#### **EXAMPLE 37**

## RPOA, RPOB, RPOC, AND RPOD, THE GENES ENCODING THE SUBUNITS COMPRISING ALLOICOCCUS OTTTIDIS RNA POLYMERASE: ALPHA, BETA, BETA', AND SIGMA.

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RNA polymerase is an enzyme comprised of multiple highly conserved subunits which catalyzes the DNA template directed polymerization of ribonucleic nucleotides into ribonucleic acid. It is composed of a core enzyme,  $\Box 2,\Box,\Box'$ , along with a fifth subunit present in stoichiometric amounts,  $\Box\Box\Box$ which can catalyze RNA synthesis non-specifically. Holoenzyme is formed by the introduction of the subunit  $\Box\Box\Box$ , which enhances gene promoter recognition and allows specificity. Homologs of the genes identified in *Alloiococcus otitidis* are described in Example 5/Table 4 (Seq. ID Nos 7, 9, 11, and 13). The amino acid sequence of the protein encoded by these genes are set forth in Seq. ID Nos. 8, 10, 12 and 14.

Functions for the individual subunits have been defined biochemically, and interactions between them have now been deduced structurally by crystallographic analysis of the enzyme from *Thermatoga thermophila*, and to a lesser extent, *Escherichia coli*. The alpha subunit, encoded by *rpoA*, is required for enzyme assembly. It also interacts with transcription factors and with DNA elements involved in enhanced promoter strength. Beta, encoded by *rpoB*, is involved in initiation and elongation of the polymerization product. Beta' (encoded by *rpoC*), is responsible for binding of the enzyme to the DNA template. Omega is required to restore denatured RNA polymerase to function *in vitro*. Finally, sigma, encoded by *rpoD*, directs the enzyme to promoters on the template to enhance specificity of transcription (polymerization).

ALLOIOCOCCUS OTITIDIS RNA POLYMERASE: ALPHA, BETA, BETA', AND SIGMA AS A TARGET FOR ANTI-INFECTIVE DEVELOPMENT

Bacterial RNA polymerase is a validated target for antimicrobial chemotherapy in that several inhibitors have been identified and at least one, rifampin, is in use clinically. Alloiococcus otitidis RNA polymerase holoenzyme is essential for bacterial viability. The *Alloiococcis otitidis ORFs*- have been shown to encode, by sequence homology, RNA polymerase holoenzyme (Seq. ID Nos. 7, 9, 11 and 13). Alloiococcus otitidis RNA Polymerase activity in the presence or absence of a putative inhibitory molecule of Alloiococcus otitidis RNA Polymerase activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by *Alloiococcus otitidis*.

#### Assays for the activity of RNA polymerase

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Genes encoding the subunits of *Alloiococcus otitidis* RNA polymerase can be obtained using polymerase chain reaction amplification of the genomic region encoding them. The genes are subcloned into a standard expression vector either containing an amino acid tag for ease of purification or not. The enzyme are overexpressed in *Escherichia* coli and purified using a standard tag system or conventional chromatography.

Because RNA polymerase catalyzes the incorporation of single ribonucleotides into RNA, the incorporation of radiolabelled nucleotides into larger oligonucleotides is monitored to measure activity of the enzyme in the presence or absence of putative inhibitors of RNA polymerase activity. An automated high throughput filtration assay has been previously described for *E. coli* polymerase which uses filterplates containing a hydrophobic membrane and DEAE beads to capture polymerized RNA. G-less supercoiled DNA is used as a template at 6 ug/ml. Reaction contained 0.5 mM ATP, 0.1 mM UTP, 0.3 mM CTP, approximately 100,000 counts per minute (per 100 ul) [γ-<sup>33</sup>P] CTP (2000 Ci/mmol, NEN/DuPont), 4 % polyethylene glycol, 4 mM DTT, 10 mM MgCl<sub>2</sub>, in 50 mM Tris-acetate (pH 7.8), and 100 mM potassium acetate. The reaction is carried out at 34 degrees C for 40 minutes, with 10% DMSO present in all reactions. The reaction was stopped by adding 100 ul 15% DEAE-Sephacel bead slurry in 50% methanol, 20 mM EDTA, and 0.02% NP-40. The reaction was incubated for 40-60 minutes at room temperature without shaking, and then transferred to a unifilter plate on a filtermate cell harvester. The wells were washed

six times with 2X PBS and 0.1% NP-40. After washing the bottom of the plate was sealed, and 50 ul scintillation counting liquid was added. Radioactivity was counted using a microplate scintillation counter.

Deconvolution assays are carried out by measuring the inhibition of sigma activity. Because sigma is required only for promoter specificity, polymerization may occur non-specifically if sigma is inhibited: Consequently a second assay is described above that is used to deconvolute activity against sigma.

The binding of putative inhibitory compounds to core enzyme. Several techniques are utilized to determine the interaction of inhibitors with individual subunits and include nuclear magnetic resonance and capillary electrophoresis.

## EXAMPLE 38

# YPHC, ENCODING A SMALL GTPASE OF UNKNOWN FUNCTION FROM ALLOIOCOCCUS OTITIDIS

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The yphC was initially identified in Bacillus subtilis in a collaboration between Wyeth and Millennium pharmaceuticals as being essential for growth by insertional mutagenesis. Subsequently it was determined that YphC, the encoded protein, contained two GTPase domains and had some homology to era. It was further identified in Thermatoga maritima and Escherichia coli. While no function has yet been determined for yphC, it appears that the carboxy terminal may contain an RNA binding site. In addition, site directed mutagenesis of four amino acids in the carboxy region were found to be lethal (unpublished results, Millennium). Under non-permissive conditions, strains carrying temperature sensitive alleles of the gene in E. coli become elongated, and chromosome segregation becomes abberrant, suggesting a role in cell division. Homologue of this gene identified in Alloiococcus otitidis is described in Example 5/Table 4 (Seq. ID No 73). The protein encoded by the gene is set forth in Seq. ID No. 74.

## YphC from Alloiococcus otitidis as a target for antimicrobial chemotherapy

YphC is an essential protein in *Bacillus subtilis* and *E. coli*, and is conserved among bacteria including *Alloiococcus otitidis*. The *Alloiococcis otitidis ORF*- has been shown to encode, by sequence homology, YphC (Seq. ID No. 73). YphC activity in the presence or absence of a putative inhibitory molecule of YphC activity is used to identify novel antimicrobial agents, which may be used to treat disease caused by *Alloiococcus otitidis*.. Consequently it is proposed here that an assay which identified inhibitors of YphC from *Alloiococcus* would result in small molecules which can be developed into effect antimcrobial agents. Additionally, because of the conservation of the enzyme among bacteria, inhibitors of the protein's function from this organism should have broad spectrum activity.

## Assays for the GTP hydrolysis by YphC

The YphC gene from Alloiococcus otitidis is obtained using polymerase chain reaction amplification of the genomic region encoding it. The gene is subcloned into a standard expression vector either containing an amino acid tag for ease of purification or not. The enzyme is then overexpressed in *Escherichia* coli and purified using a standard tag system or conventional chromatography. Activity of YphC in the presence or absence putative antimicrobial agents is monitored using the assay system described below.

GTP hydrolysis – detection by thin layer chromatography: Reaction is carried out in a 50 ul reaction of 50 mM Tris-Cl (pH 7.5), 400 mM KCl, 5 mM MgCl2, 1 mM DTT, 10 uM [a-32P] GTP, and 10 ug purified YphC, at 37 degrees for 10 minutes. The reaction is terminated by transfer of 5 ul samples to 10 ul of ice-cold 20 mM EDTA. Portions are spotted onto polyethyleneimine-cellulose thin layer chromatography plates, which are developed in 0.75 KH2PO4 (pH 3.65). The plate is autoradiographed to identify hydrolysis products.

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  - Sood, P., C. G. Lerner, T. Shimamoto, Q. Lu, and M. Inouye. 1994. Characterization of the autophosphorylation of Era, an essential Escherichia coli GTPase. Mol Microbiol 12:201-8.

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#### WHAT IS CLAIMED IS:

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5 1. A purified or isolated *Alloiococcus otitidis* nucleic acid sequence comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, wherein expression of said nucleic acid is essential for the proliferation of a cell.

- 10 2. A purified or isolated nucleic acid of Alloiococcus otitidis comprising a fragment of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105 said fragment selected from the group consisting of fragments comprising at least 10, at least 20, at least 25, at least 30, at least 50 and more than 50 consecutive nucleotides of one of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.
  - 3. A purified or isolated antisense nucleic acid comprising a nucleotide sequence complementary to at least a portion of an intragenic sequence, intergenic sequence, sequences spanning at least a portion of two or more genes, 5' noncoding region, or 3' noncoding region within an operon comprising a proliferation-required gene of Alloiococcus otitidis whose activity or expression is inhibited by an antisense nucleic acid and selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.
- 4. A purified or isolated nucleic acid comprising a nucleotide sequence having at least 70% identity to a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, fragments comprising at least 25 consecutive nucleotides selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, the nucleotide sequences complementary to one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, and the sequences complementary to fragments comprising at least 25 consecutive nucleotides

of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

- A vector comprising a promoter operably linked to a nucleic acid encoding a polypeptide whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence of any one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.
- 10 5. A purified or isolated polypeptide of Alloiococcus otitidis comprising a polypeptide whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or a fragment selected from the group consisting of fragments comprising at least 5, at least 10, at least 20, at least 30, at least 40, at least 50, at least 60 or more than 60 consecutive amino acids of one of the said polypeptides.
- 6. A purified or isolated *Alloiococcus otitidis* polypeptide comprising a amino acid sequence having at least 25% amino acid identity to a polypeptide

  20 whose expression is inhibited by a nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or at least 25% amino acid identity to a fragment comprising at least 10, at least 20, at least 30, at least 40, at least 50, at least 60 or more than 60 consecutive amino acids of a polypeptide whose expression is inhibited by a nucleic acid comprising a nucleotide sequence selected from the group consisting of one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.
- 30 7. A purified or isolated Alloiococcus otitidis polypeptide comprising selected from one of the even numbered sequences set forth in Seq. ID Nos: 2 to Seq. ID Nos: 106, wherein the polypeptide is essential for the proliferation of a cell.

8. A method of producing an Alloiococcus otitidis polypeptide comprising introducing into a cell a vector comprising a promoter operably linked to a nucleic acid comprising a nucleotide sequence encoding a polypeptide whose expression is essential for the proliferation and viability of Alloiococcus otitidis, and which is inhibited by an antisense nucleic acid, and which is selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105.

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- 9. A method of inhibiting the proliferation of Alloiococcus otitidis in an individual comprising inhibiting the activity or reducing the amount of a gene product whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105 or inhibiting the activity or reducing the amount of a nucleic acid encoding said gene product.
- 10. A method for identifying a compound which influences the activity of an

  Alloiococcus otitidis gene product, which is required for proliferation, said
  gene product comprising a gene product whose expression is inhibited by an
  antisense nucleic acid comprising a nucleotide sequence selected from one
  of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105,
  said method comprising:

- (a) contacting said gene product with a candidate compound; and
- (b) determining whether said compound influences the activity of said gene product.
- 30 11. A method for identifying a compound or an antisense nucleic acid having the ability to reduce activity or level of a *Alloiococcus otitidis* gene product, which is required for proliferation, said gene product comprising a gene product whose activity or expression is inhibited by an antisense nucleic acid

comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, said method comprising the steps of:

- (a) contacting a target gene or RNA encoding said gene product with a candidate compound or antisense nucleic acid; and
- (b) measuring the activity of said target.

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- 13. A method for inhibiting cellular proliferation of *Alloiococcus otitidis* comprising introducing an effective amount of a compound with activity against a gene whose activity or expression is essential for cellular proliferation, and which is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or a compound with activity against the product of said gene into a population of *Alloiococcus otitidis* cells expressing said gene.
  - 13. A composition comprising an effective concentration of an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, or a proliferation-inhibiting portion thereof in a pharmaceutically acceptable carrier.
  - 14. A method for identifying a compound having the ability to inhibit proliferation of *Alloiococcus otitidis* cell comprising:
    - (a) identifying a homologue of a gene or gene product whose activity or level is inhibited by a nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, in a test cell, wherein said test cell is not Alloiococcus otitidis;
    - (a) identifying an inhibitory nucleic acid sequence which inhibits the activity of said homologue in said test cell;
    - (b) contacting said test cell with a sublethal level of said inhibitory nucleic acid, thus sensitizing said cell;
    - (c) contacting the sensitized cell of step (c) with a compound; and

(d) determining the degree to which said compound inhibits proliferation of said sensitized cell relative to a cell which does not contain said inhibitory nucleic acid.

5 16. A method for identifying a compound having activity against a biological pathway required for proliferation comprising:

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(a) sensitizing a cell by providing a sublethal level of an antisense nucleic acid complementary to a nucleic acid encoding a gene product required for proliferation, wherein the activity or expression of said gene product is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, in said cell to reduce the activity or amount of said gene product;

(a) contacting the sensitized cell with a compound; and

- (b) determining the degree to which said compound inhibits the growth of said sensitized cell relative to a cell which does not contain said antisense nucleic acid.
- 20 17. A method for identifying a compound having the ability to inhibit one of the *Alloiococcus otitidis* polypeptides encoded by a polynucleotide selected from one of odd numbered sequences set forth in Seq. ID Nos: 1 to Seq. ID Nos: 105, and which is essential for cellular proliferation comprising:
  - (a) contacting a cell which expresses the polypeptide with the compound; and
  - (b) determining whether said compound reduces proliferation of said contacted cell by acting on said gene product.
- 18. A method for identifying a compound having the ability to inhibit one of the purified and isolated *Alloiococcus otitidis* polypeptides selected from one of the even numbered sequences set forth in Seq. ID No.: 2 to Seq. ID No.: 106, and which is essential for cellular proliferation comprising:

(c) contacting the purified and isolated polypeptide with the compound in vitro in the presence or absence of a substrate, which is essential for the activity of the polypeptide; and

- (d) determining the effect of the compound on the polypeptide by measuring the effect of the polypeptide on the substrate.
- 19. A compound which interacts with an Alloiococcus otitidis polypeptide selected from one of the even numbered sequences set forth in Seq. ID No.: 2 to Seq. ID No.: 106 and inhibits its activity.

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- 20. A method for manufacturing an antimicrobial compound comprising the steps of screening one or more candidate compounds to identify a compound that reduces the activity or level of an *Alloiococcus otitidis* polypeptide selected from one of the even numbered sequences set forth in Seq. ID No.: 2 to Seq. ID No.: 106, said polypeptide comprising a gene product whose activity or expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105; and manufacturing the compound so identified.
- 21. A compound which inhibits proliferation of *Alloiococcus otitidis* by interacting with a gene encoding a polypeptide that is required for proliferation or with a polypeptide required for proliferation, wherein said polypeptide is selected from the group consisting of a gene product having at least 70% nucleotide sequence identity from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105, polypeptide encoded by a nucleic acid having at least 70% nucleotide sequence identity to a nucleic acid encoding a polypeptide whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105, a polypeptide having at least 25% amino acid identity to a gene product whose expression is inhibited by an antisense nucleic acid comprising a nucleotide sequence selected one of the odd numbered sequences set forth in Seq. ID No.: 1 to

Seq. ID No. 105, a polypeptide encoded by a nucleic acid comprising a nucleotide sequence which hybridizes to a nucleic acid selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105 under stringent conditions, a gene product encoded by a nucleic acid comprising a nucleotide sequence which hybridizes to a nucleic acid selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105 under moderate conditions, and a gene product whose activity may be complemented by the gene product whose activity is inhibited by a nucleic acid selected from one of the odd numbered sequences set forth in Seq. ID No.: 1 to Seq. ID No. 105.

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#### SEQUENCE LISTING

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Asp Ile Gln Asp Val His Ser Leu Leu Glu Ile Leu Asn Glu Met Asn 60 55

Val Lys Thr Asp Phe Asp Gly Asn Thr Leu Thr Ile Asp Pro Arg Glu

Met Val Ser Ile Pro Met Pro Ser Gly Lys Ile Gln Ser Leu Arg Ala 90

Ser Tyr Tyr Phe Met Gly Ala Leu Leu Ala Lys Phe Gly Lys Gly Val 105 100

Val Gly Leu Pro Gly Gly Cys Phe Leu Gly Pro Arg Pro Ile Asp Gln 115

His Leu Lys Gly Phe Arg Leu Leu Gly Ala Asp Val Asp Asn Glu Met 135

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145 150

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Ile	Tyr	Leu	Asp	Val 165	Val	Ser	Ile	Gly	Ala 170	Thr	Ile	Asn	Ile	Met 175	Leu
Ala	Ala	Val	Arg 180	Ala	Gln	Gly	Arg	Thr 185	Val	Ile	Glu	Asn	Ala 190	Ala	Arg
Glu	Pro	Glu 195	Ile	Ile	Asp	Val	Ala 200	Thr	Leu	Leu	Asn	Lys 205	Met	Gly	Ala
Lys	Ile 210	Arg	Gly	Ala	Gly	Thr 215	Asp	Met	Ile	Arg	Ile 220	Glu	Gly	Val	Asp
Gln 225		Thr	Gly	Cys	Gln 230	His	Ser	Ile	Ile	Pro 235	Asp	Arg	Ile	Glu	Ala 240
Gly	Thr	Tyr	Leu	Ala 245	Ile	Ala	Ala	Ala	Ala 250	Gly	Glu	Asp	Val	Leu 255	Val
Asn	Asn	Val	Ile 260	Val	Glu	His	Ile	Asp 265	Ser	Leu	Ile	Ala	Lys 270	Leu	Asp
Glu	lle	Gly 275		Asp	Leu	Asp	Ile 280	Gly	Glu	Asp	Ser	Ile 285	Arg	Val	Lys

Ala Pro Ser Lys Pro Leu Gln Pro Val Thr Ile Lys Thr Leu Pro Tyr

Pro Gly Phe Ala Thr Asp Leu Gln Gln Pro Ile Thr Pro Leu Leu

Leu Ala Lys Gly Glu Ser Val Ile Thr Asp Thr Ile Tyr Pro Lys Arg

Val Lys His Ile Pro Glu Leu Glu Arg Met Gly Ala Asn Ile Arg Val

Glu Ser Asp Ile Ile Leu Ile Glu Gly Gly His Pro Leu Lys Gly Ala

360

330

315

295

325

290

Gly Ala Met Tyr Leu Lys Thr Ser Asp Ser Gly Leu Val Gly Ser Arg

Glu Val Glu Ala Ser Asp Leu Arg Ala Gly Ala Cys Leu Ile Asn Ala

375

Gly Leu Ile Ala Glu Gly Gln Thr Glu Ile Thr Gly Val Asp Lys Ile 385 390 395 400

Leu Arg Gly Tyr Ser His Ile Val Glu Lys Leu Asn Asp Leu Gly Ala 405 410 415

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Val Ala Lys Ala Ile Leu Asp Gln Val His Asp Leu Met His Phe Asn

40

45

gac ctc ttg agt gaa gtg tct gaa tat cta gac ttg tca gat gac gag
Asp Leu Ser Glu Val Ser Glu Tyr Leu Asp Leu Ser Asp Asp Glu
50 55 60

atc gaa agc ggt atg ggc caa ttt tac acc gat tta aat att gac ggt

Ile Glu Ser Gly Met Gly Gln Phe Tyr Thr Asp Leu Asn Ile Asp Gly

65 70 75

cgc ttc atc tct tta ggc gac aac cat tgg ggc tta cgt gaa tgg tat
Arg Phe Ile Ser Leu Gly Asp Asn His Trp Gly Leu Arg Glu Trp Tyr
80 85 90 95

cca gtc gat tct atc gat gaa gag ttg acc cac gac aat gac ctg gag
Pro Val Asp Ser Ile Asp Glu Glu Leu Thr His Asp Asn Asp Leu Glu
100 105 110

aag gtc aca ccc aag cag gcg gaa gac ggc ttt gat gac tta gag cat

Lys Val Thr Pro Lys Gln Ala Glu Asp Gly Phe Asp Asp Leu Glu His gtc gaa aaa gaa gtg atg gat gac gca aaa gaa gaa tta gat gac cag 432 Val Glu Lys Glu Val Met Asp Asp Ala Lys Glu Glu Leu Asp Asp Gln 135 gcc gtc aat gaa gat gaa gaa aat gtt gct cca gat gaa atc acc gac 480 Ala Val Asn Glu Asp Glu Glu Asn Val Ala Pro Asp Glu Ile Thr Asp 150 528 gat gga gat gaa gac aag ctg gat gaa tac tct agc gat atc gaa gac Asp Gly Asp Glu Asp Lys Leu Asp Glu Tyr Ser Ser Asp Ile Glu Asp 165 170 ctc gaa gat gat cgt aag gct agc caa gac aag ctg tcc att gtt gac 576 Leu Glu Asp Asp Arg Lys Ala Ser Gln Asp Lys Leu Ser Ile Val Asp 185 180 612 gac gaa gat gtc tta aca aat gat gac gat gag taa Asp Glu Asp Val Leu Thr Asn Asp Asp Asp Glu

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Lys Glu Phe Asp Gly Lys Lys Lys Glu Glu Leu Ala Met Ile Asp Val 20 25 30

Ala Lys Ala Ile Leu Asp Gln Val His Asp Leu Met His Phe Asn Asp . 35 40 45

Leu Leu Ser Glu Val Ser Glu Tyr Leu Asp Leu Ser Asp Asp Glu Ile 50 55 60

Glu Ser Gly Met Gly Gln Phe Tyr Thr Asp Leu Asn Ile Asp Gly Arg 65 70 75 80

Phe Ile Ser Leu Gly Asp Asn His Trp Gly Leu Arg Glu Trp Tyr Pro 85 90 95

Val Asp Ser Ile Asp Glu Glu Leu Thr His Asp Asn Asp Leu Glu Lys 100 105 110

Val Thr Pro Lys Gln Ala Glu Asp Gly Phe Asp Asp Leu Glu His Val Glu Lys Glu Val Met Asp Asp Ala Lys Glu Glu Leu Asp Asp Gln Ala 135 130 Val Asn Glu Asp Glu Glu Asn Val Ala Pro Asp Glu Ile Thr Asp Asp 145 150 Gly Asp Glu Asp Lys Leu Asp Glu Tyr Ser Ser Asp Ile Glu Asp Leu Glu Asp Asp Arg Lys Ala Ser Gln Asp Lys Leu Ser Ile Val Asp Asp 180 185 Glu Asp Val Leu Thr Asn Asp Asp Asp Glu . 195 <210> 9 <211> 942 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (1) .. (942) <223> <400> 9 48 atg atc gaa att gaa aag cca gta att gaa aca gta gag atc agt gaa Met Ile Glu Ile Glu Lys Pro Val Ile Glu Thr Val Glu Ile Ser Glu 10 96 gat ggc aaa ttc ggt aag ttt gtt gtt gaa cca ttg gaa cgt ggt tat Asp Gly Lys Phe Gly Lys Phe Val Val Glu Pro Leu Glu Arg Gly Tyr 25 ggg act acc tta ggg aat tcc tta cgc cgc atc tta tta tca tca cta 144 Gly Thr Thr Leu Gly Asn Ser Leu Arg Arg Ile Leu Leu Ser Ser Leu 192 ccg ggt gct gcg gtc acc aat att caa att gat ggt gtt ttg cat gag Pro Gly Ala Ala Val Thr Asn Ile Gln Ile Asp Gly Val Leu His Glu 50 240 ttt aca gct att gat ggt gtg gtt gaa gat gtg act tcc atc atc tta Phe Thr Ala Ile Asp Gly Val Val Glu Asp Val Thr Ser Ile Ile Leu 70 aac ctg aaa aaa ctg gct tta aaa ctt cat act gaa gaa aca aaa aca 288

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Asn	Leu	Lys	Lys	Leu 85	Ala	Leu	Lys	Leu	His 90	Thr	Glu	Glu	Thr	Lys 95	Thr	
								gct Ala 105								336
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			_					cac His			_	_	_	_		432
								cac His								480
								att Ile								528
								gtt Val 185								576
								gat Asp							_	624
								atg Met								672
								gaa Glu								720
								ctt Leu								768
			_				Asn	tgt Cys 265	_		_	_				816
act Thr	gtc Val	caa Gln 275	gaa Glu	cta Leu	acg Thr	gac Asp	aaa Lys 280	act Thr	gaa Glu	ccg Pro	gaa Glu	atg Met 285	atg Met	aaa Lys	gtt Val	864
cgc Arg	aat Asn 290	ctc Leu	gga Gly	cgt Arg	aag Lys	tca Ser 295	tta Leu	gaa Glu	gaa Glu	gtt Val	aaa Lys 300	aac Asn	aag Lys	ctt Leu	gat Asp	912
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305 310

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<212> PRT

<213> Alloiococcus otitidis

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Gly Thr Thr Leu Gly Asn Ser Leu Arg Arg Ile Leu Leu Ser Ser Leu

Pro Gly Ala Ala Val Thr Asn Ile Gln Ile Asp Gly Val Leu His Glu

Phe Thr Ala Ile Asp Gly Val Val Glu Asp Val Thr Ser Ile Ile Leu 70

Asn Leu Lys Lys Leu Ala Leu Lys Leu His Thr Glu Glu Thr Lys Thr 85 90

Ile Glu Leu Asp Ile Glu Gly Pro Ala Glu Val Thr Ala Ala Asp Ile

Ile Thr Asp Ser Asp Val Glu Ile Met Asn Pro Asp Leu Tyr Leu Cys

Thr Val Ser Glu Gly Gly His Leu His Ile Arg Met Glu Ala Glu Thr

Gly Arg Gly Tyr Val Asn Ala Glu His Asn Lys His Asp Asp Met Pro 145 150 155

Ile Gly Val Leu Pro Ile Asp Ser Ile Tyr Thr Pro Ile Ser Arg Val 165

Asn Tyr Thr Val Glu Asp Thr Arg Val Gly Glu Arg Glu Gln Tyr Asp 185

Lys Leu Thr Leu Asp Ile Trp Thr Asp Gly Ser Ile Ser Pro Glu Asp 195 200 205

Gly Leu Ser Leu Ala Ala Lys Ile Met Asn Glu His Leu Asn Ile Phe 210 215 220

Ile Asn Leu Thr Glu Gln Ala Arg Glu Ala Asp Ile Met Val Glu Lys 225 230 235

Glu Glu Asp Gln Lys Glu Lys Met Leu Glu Met Thr Ile Glu Glu Leu 245 250 255

Asp Leu Ser Val Arg Ser Tyr Asn Cys Leu Lys Arg Ala Gly Ile Asn 260 265 270

Thr Val Gln Glu Leu Thr Asp Lys Thr Glu Pro Glu Met Met Lys Val 275 280 285

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Gln Ile Gly Leu Ala Ser Pro Glu Lys Ile Arg Ser Trp Ser His Gly
15 20 25

gaa gtg aag aaa cct gaa acc att aac tac cgg aca tta aaa cct gaa 147 Glu Val Lys Lys Pro Glu Thr Ile Asn Tyr Arg Thr Leu Lys Pro Glu 30 35 40

aaa gac ggt ttg ttc tgc gaa cgc att ttt ggc cca acc aag gac tat

195
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tgt Cys 75	gac Asp	cgt Arg	tgc Cys	ggt Gly	gtt Val 80	gaa Glu	gtc Val	acc Thr	aag Lys	tcg Ser 85	agt Ser	gtc Val	aga Arg	cga Arg	gaa Glu 90	291
cgc Arg	atg Met	ggc	cac His	ttg Leu 95	gaa Glu	tta Leu	gca Ala	gct Ala	cct Pro 100	gtc Val	acc Thr	cac His	att Ile	tgg Trp 105	tac Tyr	339
ttc Phe	aag Lys	ggt Gly	att Ile 110	cca Pro	agt Ser	cgg Arg	atg Met	ggc Gly 115	ctt Leu	atc Ile	tta Leu	gat Asp	atg Met 120	agc Ser	cca Pro	387
aga Arg	tcc Ser	ttg Leu 125	gaa Glu	gaa Glu	att Ile	atc Ile	tat Tyr 130	ttt Phe	gcç Ala	tct Ser	tat Tyr	gtt Val 135	gtt Val	att Ile	gac Asp	435
ggt Gly	ggg Gly 140	gat Asp	acc Thr	ccg Pro	ctt Leu	gaa Glu 145	cgc Arg	aaa Lys	cag Gln	ctc Leu	tta Leu 150	act Thr	gaa Glu	cgt Arg	gaa Glu	483
tac Tyr 155	cgg Arg	gaa Glu	aac Asn	aaa Lys	agc Ser 160	aag Lys	tac Tyr	ggc	aat Asn	gaa Glu 165	ttc Phe	caa Gln	gct Ala	gaa Glu	att Ile 170	531
gga Gly	gct Ala	gaa Glu	gct Ala	gtt Val 175	cgg Arg	acc Thr	ttg Leu	cta Leu	aaa Lys 180	aat Asn	gtc Val	gat Asp	ttg Leu	gaa Glu 185	caa Gln	579
gaa Glu	gtt Val	gct Ala	gac Asp 190	ctc Leu	aaa Lys	gaa Glu	atc Ile	tta Leu 195	gaa Glu	act Thr	gca Ala	act Thr	ggc Gly 200	caa Gln	aaa Lys	627
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tcc Ser	aac Asn 220	aac Asn	aaa Lys	ccg Pro	gaa Glu	tgg Trp 225	atg Met	gtc Val	ttg Leu	gat Asp	gct Ala 230	att Ile	cca Pro	att Ile	atc Ile	723
cca Pro 235	Pro	gaa Glu	ctc Leu	cgc Arg	cca Pro 240	atg Met	gta Val	caa Gln	cta Leu	gaa Glu 245	ggt Gly	ggc	cgg Arg	ttt Phe	gca Ala 250	771
					Asp								cgg Arg			819
cgg Arg	ttg Leu	aaa Lys	cgc Arg 270	Leu	ctt Leu	gac Asp	ttg Leu	aat Asn 275	Ala	ccc Pro	cac	att	atc Ile 280	Val	caa Gln	867

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tct Ser 315	ctt Leu	tct Ser	cac His	atg Met	ttg Leu 320	aaa Lys	ggg ggg	aaa Lys	caa Gln	ggg Gly 325	cgc Arg	ttc Phe	cgt Arg	cag Gln	aac Asn 330	1011
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GJA aaa	cca Pro	Thr	ctt Leu 350	aaa Lys	atg Met	tac Tyr	caa Gln	tgt Cys 355	ggt Gly	cta Leu	ccg Pro	aaa Lys ·	gaa Glu 360	atg Met	gcc Ala	1107
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gta Val	Pro 460	Leu	agt Ser	gat Asp	gaa Glu	gcc Ala 465	cag Gln	gca Ala	gaa Glu	gcc Ala	cgc Arg 470	Ile	tta Leu	atg Met	ctg Leu	1443
ggt Gly 475	Ala	caa Gln	aat Asn	atc Ile	tta Leu 480	aac Asn	cct Pro	aaa Lys	gat Asp	ggt Gly 485	Gln	cca Pro	gtc Val	gtt Val	acc Thr 490	1491
cct Pro	tcc Ser	caa Gln	gac Asp	atg Met 495	Val	cta Leu	GJ A aaa	aac Asn	tac Tyr 500	Tyr	cta Leu	acc Thr	atg Met	gaa Glu 505	gaa Glu	1539

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gaa Glu	ggt Gly	aaa Lys	att Ile 510	ggt Gly	gaa Glu	gga Gly	act Thr	gtc Val 515	ttc Phe	tcc Ser	agt Ser	gct Ala	tct Ser 520	gag Glu	gct Ala	1587
atc Ile	caa Gln	gcc Ala 525	tac Tyr	caa Gln	aca Thr	ggc Gly	tat Tyr 530	gtc Val	cac His	ctc Leu	cac His	acc Thr 535	cgg Arg	gtt Val	gcg Ala	1635
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gac Asp 555	aag Lys	tac Tyr	ttg Leu	att Ile	acc Thr 560	aca Thr	gtc Val	ggt Gly	aag Lys	att Ile 565	atc Ile	ttt Phe	aat Asn	gaa Glu	att Ile 570	1731
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								Leu				gaa Glu 695				2115
_		Val		_	_			_	_	_	_	gaa Glu				2163
	Leu										Ile	ttt Phe				2211
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	aaa Lys 875	att Ile	gtc Val	gat Asp	gcc Ala	ggt Gly 880	att Ile	gaa Glu	gaa Glu	gtt Val	act Thr 885	atc Ile	cgg Arg	tct Ser	gcc Ala	ttc Phe 890	269:	L
	tgc Cys	tgc Cys	aac Asn	acc Thr	aac Asn 895	cac His	ggt Gly	gtc Val	tgc Cys	aag Lys 900	cac His	tgc Cys	tat Tyr	ggc Gly	cgt Arg 905	aac Asn	· 2739	)
	ttg Leu	gca Ala	act Thr	ggc Gly 910	cgg Arg	gaa Glu	gtt Val	gaa Glu	gtt Val 915	ggt Gly	gaa Glu	gca Ala	gtt Val	gga Gly 920	act Thr	atc Ile	2787	,
	gct Ala	gcc Ala	caa Gln 925	tcc Ser	att Ile	Gly ggg	gaa Glu	ccc Pro 930	ggt Gly	acc Thr	caa Gln	ttg Leu	acc Thr 935	atg Met	cgg Arg	acc Thr	2835	;
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Glu Arg Thr I	aag acc gtc act Lys Thr Val Thr 990	gtt aag ggg Val Lys Gly 995	aat gtt gac caa Asn Val Asp Gln 1000	Arg Asp
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gaa gtc caa t Glu Val Gln T	ac gct tac cgg Yr Ala Tyr Arg 1055	tct caa ggg Ser Gln Gly 1060	gtc gaa att ggt Val Glu Ile Gly )	gac aag 3219 Asp Lys 1065
His Val Glu V	rtt atg gtg cga Val Met Val Arg .070	caa atg ctc Gln Met Leu 1075	cgt aaa gtc cgt Arg Lys Val Arg 1080	Val Leu
caa cca ggg g Gln Pro Gly A 1085	ac act gat atc sp Thr Asp Ile	ctg cct ggt Leu Pro Gly 1090	acc atg att gac Thr Met Ile Asp 1095	ctc cac 3315 Leu His
gac ttc aag g Asp Phe Lys G 1100	aa cgc aac caa lu Arg Asn Gln 110	Glu Thr Leu	atg tcc ggt ggc Met Ser Gly Gly 1110	caa ccc 3363 Gln Pro
gca act gct a Ala Thr Ala A 1115	ga ctg gtc cta rg Leu Val Leu 1120	ctg ggt att Leu Gly Ile	acc aag gcc tcc Thr Lys Ala Ser 1125	ctt gaa 3411 Leu Glu 1130
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1215

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Glu Arg Ile Phe Gly Pro Thr Lys Asp Tyr Glu Cys Ala Cys Gly Lys 50 55 60

Tyr Lys Arg Val His Tyr Lys Gly Ile Val Cys Asp Arg Cys Gly Val 65 70 75 80

Glu Val Thr Lys Ser Ser Val Arg Arg Glu Arg Met Gly His Leu Glu . 85 90 95

Leu Ala Ala Pro Val Thr His Ile Trp Tyr Phe Lys Gly Ile Pro Ser 100 105 110

Arg Met Gly Leu Ile Leu Asp Met Ser Pro Arg Ser Leu Glu Glu Ile 115 120 125

Ile Tyr Phe Ala Ser Tyr Val Val Ile Asp Gly Gly Asp Thr Pro Leu 130 135 140

Glu Arg Lys Gln Leu Leu Thr Glu Arg Glu Tyr Arg Glu Asn Lys Ser 145 150 155 160

Lys Tyr Gly Asn Glu Phe Gln Ala Glu Ile Gly Ala Glu Ala Val Arg

175 170 165 Thr Leu Leu Lys Asn Val Asp Leu Glu Gln Glu Val Ala Asp Leu Lys 180 190 Glu Ile Leu Glu Thr Ala Thr Gly Gln Lys Arg Thr Arg Ala Ile Arg 200 Arg Leu Asp Ile Ile Asp Ser Phe Lys Ser Ser Asn Asn Lys Pro Glu Trp Met Val Leu Asp Ala Ile Pro Ile Ile Pro Pro Glu Leu Arg Pro Met Val Gln Leu Glu Gly Gly Arg Phe Ala Thr Ser Asp Leu Asn Asp 250 245 Leu Tyr Arg Arg Val Ile Asn Arg Asn Asn Arg Leu Lys Arg Leu Leu . 260 265 270 Asp Leu Asn Ala Pro His Ile Ile Val Gln Asn Glu Lys Arg Met Leu 280 275 Gln Glu Ala Val Asp Ala Leu Ile Asp Asn Gly Arg Arg Gly Arg Ala 290 Val Asn Gly Pro Gly Asn Arg Pro Leu Lys Ser Leu Ser His Met Leu 305 310 315 Lys Gly Lys Gln Gly Arg Phe Arg Gln Asn Leu Leu Gly Lys Arg Val Asp Tyr Ser Gly Arg Ser Val Ile Val Val Gly Pro Thr Leu Lys Met Tyr Gln Cys Gly Leu Pro Lys Glu Met Ala Ile Glu Leu Phe Lys Pro 360 355 Phe Val Met Arg Glu Leu Val Glu Arg Asp Ile Ala Asn Asn Ile Lys 370 Asn Ala Lys Arg Lys Val Glu Arg Met Glu Asp Asp Val Trp Pro Val 390 395

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Leu	Glu	Asp	Val	Ile	Lys	Glu	His	Pro	Val	Leu	Leu	Asn	Arg	Ala	Pro
		_		405					410					415	

- Thr Leu His Arg Leu Gly Ile Gln Ala Phe Glu Pro Val Leu Val Asn 420 425 430
- Gly Lys Ala Ile Arg Leu His Pro Leu Ala Cys Glu Ala Tyr Asn Ala 435 440 445
- Asp Phe Asp Gly Asp Gln Met Ala Val His Val Pro Leu Ser Asp Glu 450 450
- Ala Gln Ala Glu Ala Arg Ile Leu Met Leu Gly Ala Gln Asn Ile Leu 465 470 475 480
- Asn Pro Lys Asp Gly Gln Pro Val Val Thr Pro Ser Gln Asp Met Val 485 490 495
- Leu Gly Asn Tyr Tyr Leu Thr Met Glu Glu Glu Gly Lys Ile Gly Glu 500 505 510

1

- Gly Thr Val Phe Ser Ser Ala Ser Glu Ala Ile Gln Ala Tyr Gln Thr 515 520 525
- Gly Tyr Val His Leu His Thr Arg Val Ala Ile Arg Ala Val Asp Leu 530 540
- Pro Asp Lys Pro Phe Thr Asp Trp Gln Lys Asp Lys Tyr Leu Ile Thr 545 550 555
- Thr Val Gly Lys Ile Ile Phe Asn Glu Ile Met Pro Ala Glu Phe Pro 565 570 575
- Phe Leu Asn Glu Pro Ser Lys Val Asn Leu Glu Gln Gln Thr Pro Asp 580 585 590
- Lys Tyr Phe Val Asp Arg Gly Gln Asn Leu Lys Asp Leu Ile Ala Asp 595 600 605
- Arg Pro Leu Val Gln Pro Phe Lys Lys Gln Asp Leu Ser Asn Ile Ile 610 615 620

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Ala Glu Val Phe Asn Asn Phe Gln Val Thr Glu Thr Ser Lys Met Leu 630

Asp Arg Met Lys Asn Leu Gly Tyr Lys Tyr Ser Thr Arg Ser Gly Ile

Thr Val Gly Ile Ala Asp Val Ser Val Leu Glu Ala Lys Pro Glu Ile 665

Leu Lys Glu Ala His Ala Lys Val Asp Lys Ile Asn Ala Thr His Arg 680 675

Arg Gly Leu Ile Thr Glu Glu Glu Arg Tyr Asp Asn Val Ile Asp Val 695 700

Trp Gln Lys Ala Lys Asp Glu Ile Gln Asp Ala Leu Met Asp Ser Leu · 710 715

Asp Pro Arg Asn Asn Ile Phe Met Met Ser Asp Ser Gly Ala Arg Gly 725

Asn Ile Ser Asn Phe Thr Gln Leu Ala Gly Met Arg Gly Leu Met Ala 740 745

Ala Pro Ser Gly Glu Ile Met Glu Leu Pro Ile Thr Ser Asn Phe Arg

Glu Gly Leu Ser Val Leu Glu Met Phe Ile Ser Thr His Gly Ala Arg

Lys Gly Met Thr Asp Thr Ala Leu Lys Thr Ala Asp Ser Gly Tyr Leu

Thr Arg Arg Leu Val Asp Val Ala Gln Asp Val Ile Ile Arg Glu Glu 815 805 810

Asp Cys Gly Thr Lys Arg Gly Leu Lys Val Ser Ala Ile Gln Val Gly 820

Asn Glu Gln Ile Glu Ser Leu Ser Asp Arg Ile Leu Gly Arg Tyr Ala 840 835

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- Gln Glu Thr Val Thr His Pro Glu Thr Gly Glu Val Ile Val His Lys 855
- Asp Glu Leu Ile Asp Glu Gly Lys Thr Arg Lys Ile Val Asp Ala Gly 875 870
- Ile Glu Glu Val Thr Ile Arg Ser Ala Phe Cys Cys Asn Thr Asn His
- Gly Val Cys Lys His Cys Tyr Gly Arg Asn Leu Ala Thr Gly Arg Glu 900 905 910
- Val Glu Val Gly Glu Ala Val Gly Thr Ile Ala Ala Gln Ser Ile Gly 920 915
- Glu Pro Gly Thr Gln Leu Thr Met Arg Thr Phe His Thr Gly Gly Val
- Ala Gly Asp Asp Ile Thr Gln Gly Leu Pro Arg Val Gln Glu Ile Phe 950 945
- Glu Ala Arg His Pro Lys Gly Gln Ala Thr Ile Thr Glu Val Asn Gly 970
- Gln Ile Gln Glu Ile Val Glu Asp Pro Glu Glu Arg Thr Lys Thr Val 985
- Thr Val Lys Gly Asn Val Asp Gln Arg Asp Tyr Ser Leu Pro Ile Asn 995 1000
- Ala Arg Met Lys Val Glu Val Gly Asp Tyr Val Glu Arg Gly Asp Ala 1020 1010 1015
- Leu Asn Glu Gly Ser Ile Asp Pro Lys Glu Leu Leu Ala Val Ser Asp 1025 1040
- Met Met Lys Leu Gln Lys Tyr Leu Leu Gln Glu Val Gln Tyr Ala Tyr 1050
- Arg Ser Gln Gly Val Glu Ile Gly Asp Lys His Val Glu Val Met Val 1065 1060
- Arg Gln Met Leu Arg Lys Val Arg Val Leu Gln Pro Gly Asp Thr Asp

27/235

1075 1080 1085

Ile Leu Pro Gly Thr Met Ile Asp Leu His Asp Phe Lys Glu Arg Asn 1090 1095 1100

Gln Glu Thr Leu Met Ser Gly Gly Gln Pro Ala Thr Ala Arg Leu Val

Leu Leu Gly Ile Thr Lys Ala Ser Leu Glu Thr Asn Ser Phe Leu Ser 1125 1130 1135

Ala Ala Ser Phe Gln Glu Thr Thr Arg Val Leu Thr Asp Ala Ala Ile 1140 1145 1150

Arg Gly Lys Val Asp Asp Leu Val Gly Leu Lys Glu Asn Val Ile Ile 1155 1160 1165

Gly Lys Ser Ile Pro Ala Gly Thr Gly Met Arg Ala Tyr Ser Asn Ile 1170 1175 1180

Glu Pro Lys Lys Val Gly Val Val Ser Glu Asn Val Tyr Ser Ile Asn 1185 1190 1195 1200

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Ser Glu Lys

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gtt cgt aga agt tac tca cga atc aac gaa gta ctc gag ctc ccg aat
Val Arg Arg Ser Tyr Ser Arg Ile Asn Glu Val Leu Glu Leu Pro Asn
20 25 30

•																
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ttg Leu	aag Lys	gaa Glu 50	atg Met	ttt Phe	agt Ser	gat Asp	att Ile 55	tcc Ser	cca Pro	atc Ile	gat Asp	gat Asp 60	ttt Phe	tca Ser	ggc Gly	192
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act Thr 80	gtt Val	gaa Glu	gaa Glu	gct Ala	aga Arg 85	gag Glu	cat His	gat Asp	gcg Ala	aac Asn 90	tat Tyr	tct Ser	gcc Ala	ccc Pro	ctc Leu 95	288
tac Tyr	gtg Val	aag Lys	tta Leu	cgt Arg 100	ttg Leu	atc Ile	aac Asn	aag Lys	gaa Glu 105	act Thr	ggt Gly	gaa Glu	gtc Val	aag Lys 110	gaa Glu	336
caa Gln	gaa Glu	gtc Val	ttc Phe 115	ttc Phe	ggt Gly	gac Asp	ttt Phe	ccg Pro 120	tta Leu	atg Met	aca Thr	gaa Glu	caa Gln 125	GJA aaa	acc Thr	384
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gaa Glu	aca Thr	gat Asp	acc Thr	aaa Lys 180	: Gly	atc Ile	tcc Ser	aat Asn	gtt Val 185	Arg	att	gac Asp	cga Arg	acc Thr 190	Arg	576
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gco Ala 240	a Lei	g aaa u Lys	a gad s Asp	c tto p Le	tai 1 Ty: 24!	r Glı	cgo Arg	tt: J Le	g cgt ı Arç	cca p Pro 250	G13	gaa Glu	a cco	c aaa o Lys	a aca 5 Thr 255	768
gct	gaa	a tco	tc!	t cgi	t aa	c ttg	g ato	aa a	t acc	cgg	g tto	: tti	t gad	ca	c aag	816

Ala	Glu	Ser	Ser	Arg 260	Asn	Leu	Ile	Asn	Thr 265	Arg	Phe	Phe	Asp	His 270	Lys	
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gac Asp	ctt Leu	aaa Lys 290	acc Thr	cgc Arg	ttg Leu	atg Met	ggg Gly 295	act Thr	gtc Val	ctt Leu	gcc Ala	gaa Glu 300	aac Asn	ctg Leu	gtt Val	912
gat Asp	cct Pro 305	gaa Glu	gct Ala	ggc Gly	gag Glu	gtc Val 310	tta Leu	gct Ala	gaa Glu	gaa Glu	ggt Gly 315	agt Ser	gaa Glu	gtg Val	acc Thr	960
cgg Arg 320	tct Ser	gtg Val	atg Met	gac Asp	aag Lys 325	ctt Leu	Gly	cct Pro	tac Tyr	ctt Leu 330	gac Asp	ggt Gly	gac Asp	atg Met	aac Asn 335	1008
									gcg Ala 345							1056
-									aaa Lys	_						1104
									gac Asp							1152
									agt Ser							1200
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	_		Thr						aac Asn		_					1392
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480					485					490					495	
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gac Asp	gtc Val	cac His	tat Tyr 515	tcc Ser	cac His	tac Tyr	ggc Gly	cgg Arg 520	atg Met	tgc Cys	ccg Pro	atc Ile	gaa Glu 525	aca Thr	cct Pro	1584
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caa Gln	gtt Val 625	Val	tct Ser	gtg Val	gcc Ala	aca Thr 630	gct Ala	tcc Ser	att	cct Pro	ttc Phe 635	tta Leu	gaa Glu	aac Asn	gac Asp	1920
gac Asp 640	Ser	aac	cgg Arg	gct Ala	cta Leu 645	atg Met	Gly	gct Ala	aac Asn	atg Met 650	Gln	cgg Arg	caa Gln	gct Ala	gtt Val 655	1968
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att Ile	gca Ala	gcc	cgt Arg 675	Asp	tct Ser	gga	gct Ala	gcc Ala 680	val	att Ile	gcc Ala	aag Lys	gct Ala 685	Asp	Gly	2064
gtt Val	gtg Val	gag Glu	ı Tyr	gtt Val	gat . Asp	gcc Ala	aag Lys 695	Thi	g gtc Val	aaa Lys	gto Val	cgt Arg 700	Gln	gcc Ala	gat Asp	2112
ggt Gly	acc Thr	Lev	aac 1 Asi	aac Asr	tac Tyr	t aag Lys 710	Leu	gct Ala	aag A Lys	g tac Tyr	: aaa : Lys 715	s Arg	tco Ser	aac Asn	tcc Ser	2160

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gac Asp	aaa Lys	ggc Gly	gac Asp	atc Ile 740	cta Leu	gca Ala	gat Asp	ggt Gly	ccg Pro 745	tcc Ser	atg Met	gaa Glu	aat Asn	ggt Gly 750	gaa Glu	2256
atg Met	gcc Ala	ctt Leu	ggt Gly 755	aaa Lys	aac Asn	cca Pro	ttg Leu	att Ile 760	gcc Ala	ttt Phe	acc Thr	acc Thr	ttt Phe 765	gat Asp	ggc Gly	2304
tac Tyr	aac Asn	ttc Phe 770	gag Glu	gat Asp	gcc Ala	gtc Val	att Ile 775	atg Met	agt Ser	gaa Glu	cgt Arg	ttg Leu 780	gtc Val	aaa Lys	gat Asp	2352
gac Asp	gtt Val 785	tat Tyr	acc Thr	tcc Ser	atc Ile	cac His 790	att Ile	gaa Glu	gaa Glu	tat Tyr	gaa Glu 795	tct Ser	gaa Glu	gcc Ala	cgc Arg	2400
gat Asp 800	acc Thr	aag Lys	tta Leu	GJA aaa	cca Pro 805	gaa Glu	gaa Glu	atc Ile	acc Thr	cgg Arg 810	gaa Glu	att Ile	cca Pro	aac Asn	gtc Val 815	2448
GJÀ aaa	gaa Glu	agt Ser	gcc Ala	ctc Leu 820	aag Lys	aac Asn	ttg Leu	gat Asp	gaa Glu 825	aga Arg	ggc	att Ile	atc Ile	cgg Arg 830	atc Ile	2496
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aaa Lys	Gly ggg	gtt Val 850	Ser	gaa Glu	cta Leu	tca Ser	gct Ala 855	Glu	gaa Glu	aaa Lys	ctc Leu	ctc Leu 860	His	gct Ala	atc Ile	2592
ttc Phe	ggc Gly 865	Glu	aaa Lys	gcc Ala	cgg Arg	gaa Glu 870	Val	cgt Arg	gac Asp	acc Thr	tcc Ser 875	Leu	cgt Arg	gtc Val	cca Pro	2640
cac His 880	Gly	agt Ser	ggc Gly	gga Gly	att 11e 885	val	cac His	gat Asp	gtc Val	cag Gln 890	Ile	ttt Phe	acc Thr	Cgg	gaa Glu 895	2688
gcc Ala	. Gly	gac Asp	gaa Glu	ctg Lev 900	ı Ser	cca Pro	ggc Gly	gtt Val	aac Asr 905	TYT	ctt Leu	gto Val	cga Arg	gtt Val 910	ttc Phe	2736
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cac His	: G17 : aac	aac Asi 930	ı Lys	g ggt s Gly	gtt Val	gtt L Val	tco Ser 935	. Le	ato 1 Ile	tta Lev	cca Pro	a gaa o Glu 940	ı Glu	gad Asi	atg Met	2832

32/235

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- -

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<210> 14 <211> 1192 <212> PRT <213> Alloiococcus otitidis

Met Asn Lys Leu Val Gly Lys Lys Val Asn Phe Gly Lys His Arg Val

Ile Glu Ile Gln Thr Asp Ser Tyr Asp Trp Phe Leu Asp Glu Gly Leu 35 40 45

Lys Glu Met Phe Ser Asp Ile Ser Pro Ile Asp Asp Phe Ser Gly Asn 50 55 60

Leu Ser Leu Glu Phe Val Asp Tyr Lys Phe Tyr Glu Ser Lys Tyr Thr 65 70 75 80

Val Glu Glu Ala Arg Glu His Asp Ala Asn Tyr Ser Ala Pro Leu Tyr 85 90 95

Val Lys Leu Arg Leu Ile Asn Lys Glu Thr Gly Glu Val Lys Glu Gln 100 105 110

Glu Val Phe Phe Gly Asp Phe Pro Leu Met Thr Glu Gln Gly Thr Phe 115 120 125

Ile Ile Asn Gly Ala Glu Arg Val Ile Val Ser Gln Leu Val Arg Ser 130 135 140

Pro Gly Val Tyr Tyr Ser Pro Lys Val Glu Lys Asn Gly Arg Glu Gly 145 150 155

Phe Ser Thr Val Leu Ile Pro Asn Arg Gly Ala Trp Leu Glu Tyr Glu 165 170 175

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Thr Asp Thr Lys Gly Ile Ser Asn Val Arg Ile Asp Arg Thr Arg Lys 180 185 Ile Pro Ile Thr Val Leu Leu Arg Ala Leu Gly Ile Gly Ser Asp Asp Glu Ile Ile Asp Leu Ile Gly Leu Asn Asp Ser Leu Glu Ala Thr Leu Glu Lys Asp Val His Lys Ser Thr Ser Asp Ser Arg Val Glu Glu Ala 235 230 225 Leu Lys Asp Leu Tyr Glu Arg Leu Arg Pro Gly Glu Pro Lys Thr Ala 250 Glu Ser Ser Arg Asn Leu Ile Asn Thr Arg Phe Phe Asp His Lys Arg 265 260 Tyr Asp Leu Ala Tyr Val Gly Arg Tyr Lys Met Asn Lys Lys Leu Asp 280 275 Leu Lys Thr Arg Leu Met Gly Thr Val Leu Ala Glu Asn Leu Val Asp 290 295 Pro Glu Ala Gly Glu Val Leu Ala Glu Glu Gly Ser Glu Val Thr Arg Ser Val Met Asp Lys Leu Gly Pro Tyr Leu Asp Gly Asp Met Asn Gln 330 Val Thr Ile Asn Pro Ser Glu Glu Ala Val Ile Pro Glu Pro Ile Asp Leu Gln Ile Val Lys Val Tyr Ser Lys Glu Asp Pro Asp Arg Ile Val 360 365 355 Asn Met Ile Gly Asn Gly His Pro Asp Lys Lys Ala Lys Trp Ile Thr 370 Pro Ala Asp Met Ile Ala Ala Met Ser Tyr Phe Phe Asn Leu Gln Glu

390

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Gly Ile Gly Asp Val Asp Asp Ile Asp His Leu Gly Asn Arg Arg Ile 410 Arg Ser Val Gly Glu Leu Leu Gln Asn Gln Phe Arg Ile Gly Leu Ser 425 Arg Met Glu Arg Val Val Arg Glu Arg Met Ser Ile Gln Asp Ile Ser Ser Thr Thr Pro Gln Gln Leu Ile Asn Ile Arg Pro Val Val Ala Ser 450 455 460 Leu Lys Glu Phe Phe Gly Ser Ser Gln Leu Ser Gln Phe Met Asp Gln 470 Thr Asn Pro Leu Gly Glu Leu Thr His Lys Arg Arg Leu Ser Ala Leu-490 Gly Pro Gly Gly Leu Thr Arg Asp Arg Ala Gly Tyr Glu Val Arg Asp 505 Val His Tyr Ser His Tyr Gly Arg Met Cys Pro Ile Glu Thr Pro Glu 520 Gly Pro Asn Ile Gly Leu Ile Asn Ser Leu Ser Thr Tyr Ala Lys Ile 535 Asn Lys Phe Gly Phe Ile Glu Thr Pro Tyr Arg Arg Val Asp Arg Glu 555 Thr Gly Gln Val Thr Asp Lys Ile Asp Tyr Leu Thr Ala Asp Glu Glu 565 Asp Leu Tyr Val Val Ala Gln Ala Asn Ala Glu Leu Asp Glu Asp Gly 585 His Phe Ala Asn Asp Val Val Leu Ala Arg Arg Arg Asp Val Asn Glu 600

Glu Val Asp Ala Ser Glu Val Asp Tyr Met Asp Val Ser Pro Lys Gln
610 615 620

Val Val Ser Val Ala Thr Ala Ser Ile Pro Phe Leu Glu Asn Asp Asp

36/235

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Gly Val Ser Glu Leu Ser Ala Glu Glu Lys Leu Leu His Ala Ile Phe

850

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- Gly Glu Lys Ala Arg Glu Val Arg Asp Thr Ser Leu Arg Val Pro His 875
- Gly Ser Gly Gly Ile Val His Asp Val Gln Ile Phe Thr Arg Glu Ala 890
- Gly Asp Glu Leu Ser Pro Gly Val Asn Tyr Leu Val Arg Val Phe Ile
- Ala Gln Lys Arg Lys Ile Asp Val Gly Asp Lys Met Ala Gly Arg His 920
- Gly Asn Lys Gly Val Val Ser Leu Ile Leu Pro Glu Glu Asp Met Pro 935 930
- Phe Met Pro Asp Gly Thr Pro Ile Asp Ile Met Leu Asn Pro Leu Gly 950 945
- Val Pro Ser Arg Met Asn Val Gly Gln Val Ile Glu Leu His Met Gly 970 965
- Met Ala Ala Arg Gln Leu Gly Glu His Ile Ala Thr Pro Val Phe Asp 985
- Gly Ala Asn Glu Glu Asp Val Trp Glu Thr Ile Lys Glu Ala Gly Met 1000
- Asp Ala Asp Ala Lys Thr Val Leu Tyr Asp Gly Arg Thr Gly Glu Pro 1015 1020 1010
- Phe Asp Asn Lys Val Ser Val Gly Val Met Tyr Phe Ile Lys Leu Val 1030 . 1025 1035
  - His Met Val Asp Asp Lys Leu His Ala Arg Ser Thr Gly Pro Tyr Ser 1050 1055 1045
  - Leu Val Thr Gln Gln Pro Leu Gly Gly Lys Ala Gln Phe Gly Gly Gln 1065
  - Arg Phe Gly Glu Met Glu Val Trp Ala Leu Glu Ala Tyr Gly Ala Ser 1080 1075

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Arg Thr Leu Gln Glu Ile Leu Thr Tyr Lys Ser Asp Asp Val Ile Gly 1090 1095 1100	
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Pro Gly Val Pro Glu Ser Phe Arg Val Leu Val Lys Glu Leu Gln Ser 1125 1130 1135	
Leu Gly Leu Asp Leu Lys Val Leu Asp Lys Glu Gln Asn Glu Ile Asn 1140 1145 1150	
Leu Lys Ala Glu Asp Asp Glu Ser Glu Asp Gln Val Val Asp Ser Leu 1155 1160 1165	
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ctc tta aat gca gaa gcc ttg gtg gcg gcc atg gaa tat gtg gat gaa Leu Leu Asn Ala Glu Ala Leu Val Ala Ala Met Glu Tyr Val Asp Glu 30 35 40	147
gat gac ttt tac cgg cgg gcc cac cag ttg atc ttt aag gcc atg ata Asp Asp Phe Tyr Arg Arg Ala His Gln Leu Ile Phe Lys Ala Met Ile 45 50 55	195
gac ctc tat gaa gac aac cag gcc att gat gtc att acc att aaa gac	243

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Asp Leu Tyr Glu Asp Asn Gln Ala Ile Asp Val Ile Thr Ile Lys Asp aag ctg gaa gcc aat gac cag ttg gag gat atc ggg ggt gcc tct tac 291 Lys Leu Glu Ala Asn Asp Gln Leu Glu Asp Ile Gly Gly Ala Ser Tyr 80 339 cta gct gag att gct ggg gtc acc cca acc gca gct aac gtg tcc tat Leu Ala Glu Ile Ala Gly Val Thr Pro Thr Ala Ala Asn Val Ser Tyr 95 387 tac gct aag att gtg gaa gat cgg tct ctt ttg cgc aac ttg att gcg Tyr Ala Lys Ile Val Glu Asp Arg Ser Leu Leu Arg Asn Leu Ile Ala 115 aca gct aat gag att gcc cag tct ggc tac gaa gac cat gac gat gtg 435 Thr Ala Asn Glu Ile Ala Gln Ser Gly Tyr Glu Asp His Asp Asp Val 130 125 cca gaa gtt tta aac aat gct gag cag aag atc ttg cag gtt tct gaa 483 Pro Glu Val Leu Asn Asn Ala Glu Gln Lys Ile Leu Gln Val Ser Glu 145 140 aaa cga aac cgg acc ggc ttt gct agt att tca gaa atc ctc cac caa 531 Lys Arg Asn Arg Thr Gly Phe Ala Ser Ile Ser Glu Ile Leu His Gln 160 579 acc atc gag cat att gat gaa ctc cac caa agg gat gaa gag atc acc Thr Ile Glu His Ile Asp Glu Leu His Gln Arg Asp Glu Glu Ile Thr 175 180 ggg att tca act ggc tac ccc tac ctg gac agg atg act tca ggc ctt 627 Gly Ile Ser Thr Gly Tyr Pro Tyr Leu Asp Arg Met Thr Ser Gly Leu cat gaa gat gag ttg att att gtc gca gca aga ccg ggt gtg ggg aaa 675 His Glu Asp Glu Leu Ile Ile Val Ala Ala Arg Pro Gly Val Gly Lys 205 acg gct ttt gcc ttg aat gtc gcc caa aat atc ggg aca gcc aca gat 723 Thr Ala Phe Ala Leu Asn Val Ala Gln Asn Ile Gly Thr Ala Thr Asp 220 225 771 gaa act att gcg att ttt tcc ctt gag atg ggg gct gaa cag ctg gtc Glu Thr Ile Ala Ile Phe Ser Leu Glu Met Gly Ala Glu Gln Leu Val 240 819 aac cgg atg tta tgt tca gaa ggc agt att gat gcc act aac ctc cga Asn Arg Met Leu Cys Ser Glu Gly Ser Ile Asp Ala Thr Asn Leu Arg 260 aat ggc aag cta acg ccg gaa gaa tat gac cgt ttg ttt gtg gcc atg 867 Asn Gly Lys Leu Thr Pro Glu Glu Tyr Asp Arg Leu Phe Val Ala Met 275 270 915 ggg agc ttg tct gaa gct gat att tat att gat gac act ccc ggc atc Gly Ser Leu Ser Glu Ala Asp Ile Tyr Ile Asp Asp Thr Pro Gly Ile

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	40/235	

	·															
			285					290					295			
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gga Gly	agt Ser 315	ctg Leu	ggc	ttg Leu	att Ile	gtc Val 320	att Ile	gac Asp	tac Tyr	ctg Leu	caa Gln 325	ttg Leu	atc Ile	gaa Glu	gga Gly	1011
gct Ala 330	tca Ser	aac Asn	tat Tyr	gaa Glu	tcc Ser 335	aga Arg	cag Gln	cag Gln	cag Gln	gtg Val 340	tct Ser	gat Asp	ata Ile	tct Ser	cgg Arg 345	1059
cag Gln	ctg Leu	aag Lys	aag Lys	ctt Leu 350	tct Ser	aag Lys	gaa Glu	ctt Leu	tct Ser 355	gtc Val	cca Pro	gtt Val	att Ile	gcc Ala 360	ctg Leu	1107
tca Ser	caa Gln	ctg Leu	tcc Ser 365	cgg Arg	agt Ser	gtg Val	gaa Glu	cag Gln 370	aga Arg	caa Gln	gac Asp	Lys	cgg Arg 375	ccc Pro	atc Ile	1155
ctc Leu	agt Ser	gac Asp 380	ttg Leu	cgg Arg	gaa Glu	tca Ser	382 GJA āāā	Ser	att Ile	gaa Glu	cag Gln	gat Asp 390	Ala	gat Asp	att Ile	1203
gtg Val	gcc Ala 395	Phe	ctt Leu	tac Tyr	cgg Arg	gag Glu 400	Asp	tac Tyr	tac Tyr	caa Gln	aat Asn 405	Glu	gaa Glu	gat Asp	atc Ile	1251
gat Asp 410	Glu	gac Asp	ttt Phe	gtc Val	gat Asp 415	Asn	agc Ser	gtg Val	gaa Glu	gtc Val 420	Ile	atc :Ile	gaa Glu	aaa Lys	aac Asn 425	1299
cgg Arg	tca Ser	gga Gly	gct Ala	cga Arg 430	Gly	aca Thr	gto Val	aag Lys	ttg Leu 435	Asn	ttt Phe	aag Lys	aaa Lys	gag Glu 440	ttc Phe	1347
aac Asn	aaa Lys	ttt Phe	acc Thr 445	Ser	att : Ile	tct Ser	tac Tyr	cgg Arg 450	Ser	gaa Glu	gat Asp	gaa Glu	gto Val 455	. Pro	gcc Ala	1395
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<213> Alloiococcus otitidis

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- Ala Glu Gln Ser Val Leu Gly Ser Val Leu Leu Asn Ala Glu Ala Leu 25
- Val Ala Ala Met Glu Tyr Val Asp Glu Asp Asp Phe Tyr Arg Arg Ala
- His Gln Leu Ile Phe Lys Ala Met Ile Asp Leu Tyr Glu Asp Asn Gln
- Ala Ile Asp Val Ile Thr Ile Lys Asp Lys Leu Glu Ala Asn Asp Gln 70
- Leu Glu Asp Ile Gly Gly Ala Ser Tyr Leu Ala Glu Ile Ala Gly Val
- Thr Pro Thr Ala Ala Asn Val Ser Tyr Tyr Ala Lys Ile Val Glu Asp
- Arg Ser Leu Leu Arg Asn Leu Ile Ala Thr Ala Asn Glu Ile Ala Gln 120
- Ser Gly Tyr Glu Asp His Asp Asp Val Pro Glu Val Leu Asn Asn Ala 135
- Glu Gln Lys Ile Leu Gln Val Ser Glu Lys Arg Asn Arg Thr Gly Phe
- Ala Ser Ile Ser Glu Ile Leu His Gln Thr Ile Glu His Ile Asp Glu 165
- Leu His Gln Arg Asp Glu Glu Ile Thr Gly Ile Ser Thr Gly Tyr Pro 180
- Tyr Leu Asp Arg Met Thr Ser Gly Leu His Glu Asp Glu Leu Ile Ile 200
- Val Ala Ala Arg Pro Gly Val Gly Lys Thr Ala Phe Ala Leu Asn Val
- Ala Gln Asn Ile Gly Thr Ala Thr Asp Glu Thr Ile Ala Ile Phe Ser
- Leu Glu Met Gly Ala Glu Gln Leu Val Asn Arg Met Leu Cys Ser Glu

255 250 245 Gly Ser Ile Asp Ala Thr Asn Leu Arg Asn Gly Lys Leu Thr Pro Glu 265 260 Glu Tyr Asp Arg Leu Phe Val Ala Met Gly Ser Leu Ser Glu Ala Asp 280 Ile Tyr Ile Asp Asp Thr Pro Gly Ile Arg Thr Ala Glu Ile Arg Ala 300 Lys Cys Arg Arg Leu Val Gln Glu Lys Gly Ser Leu Gly Leu Ile Val 305 Ile Asp Tyr Leu Gln Leu Ile Glu Gly Ala Ser Asn Tyr Glu Ser Arg 325 Gln Gln Gln Val Ser Asp Ile Ser Arg Gln Leu Lys Lys Leu Ser Lys 345 Glu Leu Ser Val Pro Val Ile Ala Leu Ser Gln Leu Ser Arg Ser Val ·355 Glu Gln Arg Gln Asp Lys Arg Pro Ile Leu Ser Asp Leu Arg Glu Ser 370 Gly Ser Ile Glu Gln Asp Ala Asp Ile Val Ala Phe Leu Tyr Arg Glu 390 Asp Tyr Tyr Gln Asn Glu Glu Asp Ile Asp Glu Asp Phe Val Asp Asn 410 Ser Val Glu Val Ile Ile Glu Lys Asn Arg Ser Gly Ala Arg Gly Thr 430 Val Lys Leu Asn Phe Lys Lys Glu Phe Asn Lys Phe Thr Ser Ile Ser 435 Tyr Arg Ser Glu Asp Glu Val Pro Ala Asn Phe Gly 455 450

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ctg Leu 15	tcc Ser	aag Lys	gag Glu	atg Met	aaa Lys 20	aac Asn	tca Ser	ttc Phe	tta Leu	gac Asp 25	tat Tyr	gcc Ala	atg Met	agt Ser	gtc Val 30	99
atc Ile	gtc Val	tcc Ser	cgg Arg	gcc Ala 35	cta Leu	ccc Pro	gat Asp	gtc Val	cgg Arg 40	gac Asp	ggc	ttg Leu	aag Lys	ccg Pro 45	gtt Val	147
cac	cga Arg	aga Arg	atc Ile 50	ctg Leu	tac Tyr	gga Gly	atg Met	aat Asn 55	gaa Glu	ctg Leu	ggc Gly	tta Leu	acc Thr 60	ccg Pro	gac Asp	195
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tac Tyr	cac His 80	ccc Pro	cac His	ggt Gly	gac Asp	act Thr 85	gct Ala	att Ile	tat Tyr	gac Asp	tcc Ser 90	atg Met	gtc Val	aga Arg	atg Met	291
gcc Ala 95	cag Gln	gac Asp	ttt Phe	tct Ser	tac Tyr 100	cga Arg	gtt Val	ccc Pro	tta Leu	gtg Val 105	gac Asp	ggc Gly	cat His	Gly ggg	aac Asn 110	339
ttt Phe	ggg Gly	tcg Ser	gtt Val	gac Asp 115	Gly	gac Asp	gga Gly	gct Ala	gct Ala 120	Ala	atg Met	cgg	tat Tyr	acc Thr 125	gaa Glu	387
gcc Ala	cgg Arg	atg Met	tcc Ser 130	Lys	atg Met	gcc Ala	ttg Leu	gaa Glu 135	Leu	ctg Leu	cga Arg	gac Asp	atc Ile 140	ASII	aag Lys	435
gat Asp	acc Thr	att Ile 145	: Asp	tac Tyr	cac His	gat Asp	aac Asn 150	Tyr	gat Asr	Gly ggg	act Thr	gag Glu 155	. Ser	gaa Glu	ccc Pro	483
gat Asp	atc Ile 160	Leu	cct Pro	gcc Ala	cgc Arg	tto Phe 165	Pro	aac Asn	cto Leu	tta Lev	gtc Val 170	ASI	. Gly	gct Ala	tcg Ser	531
gg9 Gly 175	Ile	gct Ala	gtt Val	G17 : aaa	g ato Met 180	: Ala	a acc	aat Asr	ato 1 Ile	c cca Pro 185	Pro	cac His	aat Asi	ctt Lei	aag Lys 190	579

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gaa Gli	a gtc ı Val	att Ile	gat Asp	gcc Ala 195	tgc Cys	gtc Val	ctc Leu	ttg Leu	atg Met 200	gaa Glu	aat Asn	gag Glu	gat Asp	gtg Val 205	act Thr	627
gt Va	g gct l Ala	gac Asp	ctt Leu 210	atg Met	gaa Glu	gtc Val	tta Leu	cca Pro 215	gga Gly	cct Pro	gac Asp	ttt Phe	ccg Pro 220	act Thr	GJÀ aaa	675
gc Al	t tcc a Ser	ctt Leu 225	att Ile	ggt Gly	gtt Val	tct Ser	ggc Gly 230	gtc Val	cgc Arg	aag Lys	gct Ala	tat Tyr 235	gag Glu	acc Thr	ggt Gly	723
cg Ar	t ggg g Gly 240	tcc Ser	att Ile	aaa Lys	tta Leu	cgg Arg 245	gcc Ala	aag Lys	tcc Ser	cgg Arg	atc Ile 250	gat Asp	gtc Val	gac Asp	caa Gln	771
аа Ьу 25	a ggt s Gly 5	aag Lys	gaa Glu	aga Arg	att Ile 260	att Ile	atc Ile	gac Asp	gaa Glu	att Ile 265	cct Pro	tac Tyr	atg Met	gtc Val	aac Asn 270	819
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at Il	t gac e Asp	ggc	att Ile 290	acc Thr	gat Asp	tta Leu	aat Asn	gat Asp 295	gag Glu	tct Ser	gac Asp	cgg Arg	gaa Glu 300	ggc	ttg Leu	915
C9 Ar	g att g Ile	gtg Val 305	Ile	gat Asp	gta Val	cgc Arg	cgg Arg 310	gat Asp	act Thr	tct Ser	gct Ala	ggt Gly 315	ata Ile	tta Leu	ctt Leu	963
a.a A.s	c aag n Lys 320	Leu	tac Tyr	aaa Lys	atg Met	acc Thr 325	caa Gln	ttg Leu	cag Gln	gtt Val	tct Ser 330	Phe	ggc	ttt Phe	aac Asn	1011
at Me 33	g ctg t Lev	gct Ala	atc Ile	gtc Val	gat Asp 340	Gly	gtg Val	ccc	aaa Lys	acc Thr 345	Leu	ggc Gly	ctc Leu	aaa Lys	gac Asp 350	1059
at Il	c ctç e Lev	acc Thr	cac His	tac Tyr 355	Leu	gac Asp	cat His	caa Gln	aaa Lys 360	Thr	gtt Val	atc Ile	cgc Arg	agg Arg 365	Arg	1107
ac Th	a gag ır Glı	ttt Phe	gac Asp 370	Lys	aac Asn	aag Lys	gct Ala	gaa Glu 375	Ser	cgg Arg	gcc	cac His	atc Ile 380	Leu	gaa Glu	1155
G:	gg ctt y Lei	cgg Arg 385	Thr	gcc Ala	tta Leu	gac Asp	cat His	Ile	gat Asp	gcc Ala	att Ile	att Ile 395	Thr	att	atc Ile	1203
C(	gt cag rg Glr 400	Ser	cag Glr	caa Glr	gct Ala	gaa Glu 405	Glu	gcc Ala	aaa Lys	agt Ser	caa Glr 410	. Leu	atg Met	gct Ala	tct Ser	1251
ta	at gad	cto	tct	gac	: cgt	caa	gco	caç	gcg	att	tta	gac	atg	cgg	atg	1299

Tyr 415	Asp	Leu	Ser	Asp	Arg 420	Gln	Ala	Gln	Ala	Ile 425	Leu	Asp	Met	Arg	Met 430	
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gaa Glu	ctc Leu	tta Leu	gaa Glu 450	aaa Lys	atc Ile	gag Glu	gac Asp	ttg Leu 455	egt Arg	gac Asp	atc Ile	ttg Leu	gcc Ala 460	cgg Arg	cca Pro	1395
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aaa Lys	cac His 480	ggc Gly	caa Gln	gac Asp	cgc Arg	cta Leu 485	act Thr	gac Asp	atc Ile	cgg Arg	gtt Val 490	GIA	gaa Glu	gag Glu	ttg Leu	1491
agc Ser 495	att Ile	gaa Glu	gac Asp	gaa Glu	gac Asp 500	ttg Leu	att Ile	gaa Glu	gaa Glu	gaa Glu 505	gat Asp	atc Ile	atc Ile	att Ile	acc Thr 510	1539
ata.	tct Ser	cga Arg	aaa Lys	ggc Gly 515	tat Tyr	atc Ile	aaa Lys	cgg Arg	atg Met 520	Pro	gct Ala	gga Gly	gaa Glu	ttc Phe 525	ьys	1587
gcc Ala	caa Gln	aac Asn	cgc Arg 530	Gly	ggc	cgt Arg	Gly	gtt Val	aag Lys	GJA GGG	atg Met	act Thr	acc Thr 540	Asn	gat Asp	1635
GJÀ āāā	gac Asp	ttt Phe 545	· Val	gaa Glu	. cag . Gln	ctg Leu	act Thr 550	Phe	tgt Cys	tct Ser	agt Ser	cat His	Asp	cas Glr	atc Ile	1683
ctc Leu	tto Phe	Phe	acc Thr	aac Asr	caa Gln	ggc Gly 565	Lys	gtt Val	tat L Tyr	aag Lys	ato 110 570	5 TÀS	gcc Ala	tac Tyr	gaa Glu	1731
ato Ile 575	Pro	gaa Glu	a tat ı Tyı	GJ7	cgt Arg 580	Ası	gco n Ala	a ag	g gga s Gly	att 7 Ile 585	Pro	t gcc o Ala	ato a Ile	aac Ası	ttt Phe 590	1779
tta Lev	aat Asi	ata 110	a gat e Ası	t aas Dys 599	s Asp	) Gli	л Туз	r Ile	t cas e Gli 600	a Ala	a Me	t Va.	aac L Asi	ttg Lei 60!	g act 1 Thr	1827
gad As <u>ı</u>	caç Gli	g gca n Ala	a gat a Asj 61	o Ası	c caq o Gli	g gad n Asj	c caa o Gli	a tten Ph	e Phe	e tti	t gc e Al	g aca a Thi	a aga r Arg	a re	t ggc u Gly	1875
cg:	g gto g Vai	aaa l Ly: 62!	s Ar	g ac	g gco	c cas	n Se	t ga r Gl	a tt u Ph	t ca e Gl	a aa n As	t ate n Ile 63	e Ar	a ag g Se	t agc r Ser	1923
Gl;	g tt Y Le	7 22	ם מכי	g at a Il	c aa e As	t ct	a aa u As	t ga n Gl	a gg u Gl	c ga y As	t ga p Gl	a tte	g gt u Va	t aa l As	c gtg n Val	1971

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	46/235	

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tat Tyr	gcg Ala	gtc Val	tac Tyr	ttt Phe 675	gat Asp	gaa Glu	aaa Lys	gat Asp	atc Ile 680	cgt Arg	agc Ser	atg Met	ggt Gly	cga Arg 685	gjà aaa	2067
gct Ala	gca Ala	ggt Gly	gtc Val 690	cgt Arg	gga Gly	att Ile	cgc Arg	tta Leu 695	ggt Gly	gat Asp	Gly ggc	gac Asp	aca Thr 700	gtg Val	gtt Val	2115
gcc Ala	atg Met	gaa Glu 705	gtc Val	tta Leu	gag Glu	ccg Pro	ggc Gly 710	caa Gln	gac Asp	gta Val	tta Leu	gtc Val 715	att Ile	act Thr	gaa Glu	2163
aaa Lys	ggg Gly .720	tac Tyr	ggc	aaa Lys	cga Arg	acc Thr 725	tcc Ser	caa Gln	gaa Glu	gag Glu	tac Tyr 730	acc Thr	ctc Leu	cac His	aag Lys	2211
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ccc Pro	cta Leu	att Ile	gga Gly	ctg Leu 755	Lys	act Thr	gtc Val	tct Ser	ggt Gly 760	Gly	gag Glu	gac Asp	gtc Val	atg Met 765	IIe	2307
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cag Gln	acc Thr	tcc Ser 785	Arg	cta Leu	acc Thr	caa Gln	ggt Gly 790	Val	cgt Arg	tta Leu	att Ile	cga Arg 795	Leu	gaa Glu	gaa Glu	2403
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Arg Ile Leu Tyr Gly Met Asn Glu Leu Gly Leu Thr Pro Asp Lys Ser 50 60

Tyr Lys Lys Ser Ala Arg Ile Val Gly Asp Val Met Gly Lys Tyr His 65 70 75 80

Pro His Gly Asp Thr Ala Ile Tyr Asp Ser Met Val Arg Met Ala Gln 85 90 95

Asp Phe Ser Tyr Arg Val Pro Leu Val Asp Gly His Gly Asn Phe Gly 100 105 110

Ser Val Asp Gly Asp Gly Ala Ala Ala Met Arg Tyr Thr Glu Ala Arg 115 120 125

Met Ser Lys Met Ala Leu Glu Leu Leu Arg Asp Ile Asn Lys Asp Thr 130 135 140

Ile Asp Tyr His Asp Asn Tyr Asp Gly Thr Glu Ser Glu Pro Asp Ile 145 150 155 160

Leu Pro Ala Arg Phe Pro Asn Leu Leu Val Asn Gly Ala Ser Gly Ile 165 170 175

Ala Val Gly Met Ala Thr Asn Ile Pro Pro His Asn Leu Lys Glu Val 180 185 190

Ile Asp Ala Cys Val Leu Leu Met Glu Asn Glu Asp Val Thr Val Ala 195 200 205

Asp Leu Met Glu Val Leu Pro Gly Pro Asp Phe Pro Thr Gly Ala Ser 210 215 220

Leu Ile Gly Val Ser Gly Val Arg Lys Ala Tyr Glu Thr Gly Arg Gly 225 230 235 240

Ser Ile Lys Leu Arg Ala Lys Ser Arg Ile Asp Val Asp Gln Lys Gly

255 250 245 Lys Glu Arg Ile Ile Ile Asp Glu Ile Pro Tyr Met Val Asn Lys Ala 260 265 Lys Leu Val Glu Lys Ile Ala Glu Leu Ala Arg Asp Lys Lys Ile Asp 280 Gly Ile Thr Asp Leu Asn Asp Glu Ser Asp Arg Glu Gly Leu Arg Ile 300 Val Ile Asp Val Arg Arg Asp Thr Ser Ala Gly Ile Leu Leu Asn Lys 315 310 Leu Tyr Lys Met Thr Gln Leu Gln Val Ser Phe Gly Phe Asn Met Leu 325 Ala Ile Val Asp Gly Val Pro Lys Thr Leu Gly Leu Lys Asp Ile Leu 340 345 Thr His Tyr Leu Asp His Gln Lys Thr Val Ile Arg Arg Arg Thr Glu · 355 Phe Asp Lys Asn Lys Ala Glu Ser Arg Ala His Ile Leu Glu Gly Leu 370 Arg Thr Ala Leu Asp His Ile Asp Ala Ile Ile Thr Ile Ile Arg Gln 385 390 Ser Gln Gln Ala Glu Glu Ala Lys Ser Gln Leu Met Ala Ser Tyr Asp 410 Leu Ser Asp Arg Gln Ala Gln Ala Ile Leu Asp Met Arg Met Val Arg Leu Thr Gly Leu Glu Arg Glu Lys Ile Glu Asp Glu Tyr Ala Glu Leu 435 Leu Glu Lys Ile Glu Asp Leu Arg Asp Ile Leu Ala Arg Pro Glu Arg 455 450 Ile Lys Gln Ile Ile Lys Glu Glu Met Ile Glu Ile Ala Glu Lys His 470 475

Gly Gln Asp Arg Leu Thr Asp Ile Arg Val Gly Glu Glu Leu Ser Ile 485 490 495

Glu Asp Glu Asp Leu Ile Glu Glu Glu Asp Ile Ile Ile Thr Leu Ser 500 505 510

Arg Lys Gly Tyr Ile Lys Arg Met Pro Ala Gly Glu Phe Lys Ala Gln 515 520 525

Asn Arg Gly Gly Arg Gly Val Lys Gly Met Thr Thr Asn Asp Gly Asp 530 535 540

Phe Val Glu Gln Leu Thr Phe Cys Ser Ser His Asp Gln Ile Leu Phe 545 550 555 560

Phe Thr Asn Gln Gly Lys Val Tyr Lys Ile Lys Ala Tyr Glu Ile Pro 565 570 575

Glu Tyr Gly Arg Asn Ala Lys Gly Ile Pro Ala Ile Asn Phe Leu Asn 580 585 590

Ile Asp Lys Asp Glu Tyr Ile Gln Ala Met Val Asn Leu Thr Asp Gln 595 600 605

Ala Asp Asp Gln Asp Gln Phe Phe Phe Ala Thr Arg Leu Gly Arg Val 610 615 620

Lys Arg Thr Ala Gln Ser Glu Phe Gln Asn Ile Arg Ser Ser Gly Leu 625 630 635 640

Asn Ala Ile Asn Leu Asn Glu Gly Asp Glu Leu Val Asn Val Val Pro 645 650 655

Thr His Asn Asp Gln Ala Ile Ile Leu Ala Ser Gln Gln Gly Tyr Ala 660 665 670

Val Tyr Phe Asp Glu Lys Asp Ile Arg Ser Met Gly Arg Gly Ala Ala 675 680 685

Gly Val Arg Gly Ile Arg Leu Gly Asp Gly Asp Thr Val Val Ala Met 690 695 700

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Glu Val Leu Glu Pro Gly Gln Asp Val Leu Val Ile Thr Glu Lys Gly 705 710 Tyr Gly Lys Arg Thr Ser Gln Glu Glu Tyr Thr Leu His Lys Arg Gly Gly Lys Gly Val Lys Thr Leu His Ile Thr Asp Lys Asn Gly Pro Leu Ile Gly Leu Lys Thr Val Ser Gly Gly Glu Asp Val Met Ile Val Thr 760 Asp Gln Gly Ile Met Ile Arg Ile Glu Ala Asp Ser Ile Ser Gln Thr 775 Ser Arg Leu Thr Gln Gly Val Arg Leu Ile Arg Leu Glu Glu Asp Ser 795 790 Arg Val Ser Thr Val Ala Leu Ile Asp Ile Asp Gln Glu Leu Asp Asn 810 Gln Val Asn Gln Thr Val Glu Glu 820 <210> 19 <211> 1956 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (7)..(1956) <223> <400> 19 cgtgta atg gct gaa gat aga cca tta aca cca aat gag tta gca gaa 48 Met Ala Glu Asp Arg Pro Leu Thr Pro Asn Glu Leu Ala Glu 96 ctg aaa aaa aca tat gat gct agt caa atc caa gtc tta gaa ggc cta Leu Lys Lys Thr Tyr Asp Ala Ser Gln Ile Gln Val Leu Glu Gly Leu 25 20 15 gaa gca gtc aga gta cgg ccg ggt atg tac att ggg tcc acc agc aag 144 Glu Ala Val Arg Val Arg Pro Gly Met Tyr Ile Gly Ser Thr Ser Lys gaa ggc ctc cac cac ttg gta tgg gag atc gtg gac aat gct att gac 192

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gaa Glu	gaa Glu	tco Ser	gaa Glu	cto Lev 435	Phe	att Ile	gta Val	gaa Glu	440	/ Asp	tca Ser	a gct Ala	gga Gly	ggg Gly 445	tcg Ser	1344
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Leu His His Leu Val Trp Glu Ile Val Asp Asn Ala Ile Asp Glu Ala

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Met Ala Gly Tyr Ala Asp Lys Ile Ser Val Ser Ile Leu Glu Gly Asp 65 70 75 80

Val Ile Gln Val Ala Asp Asn Gly Arg Gly Ile Pro Val Asp Ile Gln 85 90 95

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His Gly Val Gly Ser Ser Val Val Asn Ala Leu Ser Glu Tyr Leu Gln 130 135 140

Val Gln Val His Arg Asp Gly Lys Ile Tyr Gln Gln Val Tyr Lys Arg 145 150 155 160

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Gly Thr Ile Val Thr Phe Lys Ala Asp Ser Leu Ile Phe Lys Asp Thr 180 185 190

Thr Ser Tyr Asp Phe Asn Thr Leu Ala Thr Arg Ile Arg Glu Leu Ala 195 200 205

Phe Leu Asn Arg Gly Leu Asn Ile Ser Ile Glu Asp Lys Arg Gln Ala 210 215 220

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Val Asp His Leu Asn Ser Ser Arg Glu Val Leu Tyr Glu Thr Pro Ile 245 250 255

Phe Leu Glu Gly Glu Glu Glu Gly Ile Ser Val Glu Ile Ala Leu Gln 260 265 270

His Thr Asp Ser Phe His Thr Asn Leu Met Ser Phe Ala Asn Asn Ile 275 280 285

His Thr Tyr Glu Gly Gly Met His Ile Ser Gly Phe Lys Thr Ala Leu 290 295 300

Thr Arg Ala Val Asn Asp Tyr Ala Arg Gln Asn Asn Leu Leu Arg Glu 305 310 315 320

Ser Glu Asp Asn Phe Thr Gly Asp Asp Val Arg Glu Gly Leu Thr Val 325 330 335

Val Leu Ser Ile Lys His Pro Asp Pro Gln Phe Glu Gly Gln Thr Lys 340 345

Thr Lys Leu Gly Asn Ser Glu Val Arg Gly Ile Ile Asp Arg Leu Phe 355 360 365

Ser Gln His Phe Glu Arg Tyr Leu Met Glu Asn Pro Lys Val Gly Lys 370 375 380

Arg Ile Val Asp Lys Ala Leu Leu Ala Ser Lys Ala Arg Gln Ala Ala 385 390 395 400

Lys Arg Ala Arg Glu Val Thr Arg Lys Lys Ser Gly Leu Glu Ile Ser 405 410 415

Asn Leu Pro Gly Lys Leu Ala Asp Cys Ser Ser Lys Asp Pro Glu Glu 420 425 430

Ser Glu Leu Phe Ile Val Glu Gly Asp Ser Ala Gly Gly Ser Ala Lys 435 440 445

Gln Gly Arg Ser Arg Val Phe Gln Ala Ile Leu Pro Ile Arg Gly Lys 450 455 460

Ile Leu Asn Val Glu Lys Ala Ser Ile Asp Arg Ile Leu Ala Asn Glu 465 470 475 480

Glu Ile Arg Ser Leu Phe Thr Ala Met Gly Thr Gly Phe Gly Glu Glu 485 490 495

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Ala Asp Val Asp Gly Ala His Ile Arg Thr Leu Leu Leu Thr Leu Leu 520 Tyr Arg Tyr Met Arg Pro Leu Ile Glu Ala Gly Phe Val Tyr Ile Ala 535 Gln Pro Pro Leu Tyr Gln Val Lys Gln Gly Lys Lys Val Lys Tyr Phe 545 Asp Ser Asp Arg Glu Leu Asp Ser Tyr Leu Lys Glu Ile Pro Asp Ser 565 Pro Lys Pro Ser Val Gln Arg Tyr Lys Gly Leu Gly Glu Met Asp Ala 585 Glu Gln Leu Trp Glu Thr Thr Met Asn Pro Glu His Arg Arg Leu Leu 605 600 Arg Val Asp Val Asp Asp Ala Ile Glu Ala Asp Thr Ile Phe Asp Met 610 Leu Met Gly Glu Asp Val Lys Pro Arg Arg Asp Phe Ile Lys Glu Asn 630 625 Ala Arg Tyr Val Glu Asn Ile Asp Ile 645 <210> 21 <211> 1218 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (16)..(1218) <223> agacctaatc atttt ttg aaa tgg aga aag aca aaa acc atc tat ggt ata 51 Met Lys Trp Arg Lys Thr Lys Thr Ile Tyr Gly Ile ctt aag aac aaa agg aag ttt gga ggg att ttt ttg aaa ttt tca gta 99 Leu Lys Asn Lys Arg Lys Phe Gly Gly Ile Phe Leu Lys Phe Ser Val aaa cgg acg gaa ttt cta aaa gta tta aaa aaa gta cag att gca gtg 147

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	58/235	

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Ser Asn His Glu Ser Arg Pro Ile Leu Thr Gly Val Asn Leu Ser Leu 180 185 190

Lys Glu Gly Arg Leu Lys Ala Val Ala Thr Asp Ser His Arg Leu Ser 195 200 205

Gln Arg Ser Ile Gln Leu Glu Ser Ala Pro Asp Ile Ser Phe Asp Ile 210 215 220

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gtg gad Val Asp	ttt Phe	Leu	gtt Val	tta Leu	gaa Glu	act Thr 145	Gly ggg	gta Val	GJA aaa	gga Gly	aaa Lys 150	att Ile	gat Asp	gcg Ala	483
acc aat Thr Asr 159	ı Val	gtg Val	ccc	gct Ala	cca Pro 160	Leu	gtc Val	tca Ser	gtc Val	att Ile 165	Ile	tct Ser	att Ile	Gly	531
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caa ac Gln Th 23	r Le	t gad u Ası	tao Tyi	c cag c Glr	tco Ser 240	r Phe	aaa Lys	a tao	ggt Gly	ggg Gly 245	ASI	tto Lev	gti Va	tta Leu	771
gag ac Glu Th 250	t ca r Gl:	a ato n Me	g ati t Ile	t ggt e Gly 25!	y Ası	c cad	c caq s Gli	g cte	g gta u Vai 26	l Ası	e act	gco Ala	c cta a Lem	a gct 1 Ala 265	819
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	•															
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caa Gln	aag Lys	cta Leu 300	tct Ser	gac Asp	cag Gln	cca Pro	gtg Val 305	gtt Val	gtt Val	ctt Leu	gat Asp	ggg Gly 310	gcc Ala	cac His	aac Asn	963
gaa Glu	atc Ile 315	GJA āāā	gtc Val	aag Lys	gct Ala	ctt Leu 320	aga Arg	cag Gln	tca Ser	att Ile	gac Asp 325	cac His	ttt Phe	ttc Phe	ccc Pro	1011
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gcc Ala	atc Ile 395	Leu	gac Asp	tac Tyr	ata Ile	aac Asn 400	Gln	caa Gln	gca Ala	aaa Lys	Ala	Asp	gaa Glu	att Ile	atc Ile	1251
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Gly Thr Gly Asp Lys Lys Asn Leu Asn Arg Met Arg Leu Leu Lys 20 25 30

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- Glu Leu Gly Asn Pro Glu Thr Asp Leu Pro Val Ile His Val Ala Gly 3.5
- Thr Asn Gly Lys Gly Thr Thr Cys Ala Tyr Ile Ala His Ser Leu Ala
- Arg Ala Gly Tyr Lys Thr Gly Leu Tyr Thr Ser Pro His Leu Glu Arg
- Val Asn Glu Arg Ile Arg Ile Asn Asp Arg Tyr Ile Ser Asp Gln Asp 90
- Leu Met Ala Leu Thr Gly Gln Ile Ala Pro Ile Ile Asp His Leu Glu 105
- Asp Cys Leu Gly Glu Lys Tyr Tyr Ser Phe Glu Ile Leu Thr Ala Leu
- Ala Phe Leu Tyr Phe Gln Gln Ala Gly Val Asp Phe Leu Val Leu Glu 135 130
- Thr Gly Val Gly Gly Lys Ile Asp Ala Thr Asn Val Val Pro Ala Pro 150
- Leu Val Ser Val Ile Ile Ser Ile Gly Tyr Asp His Thr His Val Leu 170
- Gly Asn Thr Leu Glu Asp Ile Thr Arg His Lys Ala Gly Ile Ile Lys
- Lys Gly Cys Pro Val Val Val Gly Pro Leu Ala Asp His Leu Leu Ala
- Ile Val Lys Glu Val Ser Lys Glu Met Asp Ser Asn Leu Thr Ile Val 215 - 220
- His Pro Asp Lys Phe Asp Ile Val His Gln Thr Leu Asp Tyr Gln Ser 240 230 235 225
- Phe Lys Tyr Gly Gly Asp Leu Val Leu Glu Thr Gln Met Ile Gly Asn 250

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His Gln Leu Val Asn Thr Ala Leu Ala Tyr Glu Ala Leu Lys Ile Val 265

Gln Gln Ser Tyr Pro Asp Leu Thr Asp Leu Asp Ile Leu Glu Gly Leu 280

Lys Thr Thr His Trp Pro Gly Arg Met Gln Lys Leu Ser Asp Gln Pro 295

Val Val Val Leu Asp Gly Ala His Asn Glu Ile Gly Val Lys Ala Leu 310 315

Arg Gln Ser Ile Asp His Phe Phe Pro Gly Lys Lys Ile Thr Tyr Phe 325

Ala Gly Met Met Val Glu Lys Asp Phe Ala Lys Met Phe Asp Leu Leu

Gly Glu Thr Ala Asp Lys Phe Tyr Leu Ile Ser Pro Asp Leu Thr Arg 360

Gly Phe Asp Val Asp Gln Ala Val Gln Ser Leu Thr Asp Lys Gly Tyr 380

Gln Ala Ser Ser Val Ala Ser Leu Gln Ala Ile Leu Asp Tyr Ile Asn 390

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<210> 25

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<212> DNA

<213> Alloiococcus otitidis

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<223>

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acc ctt	ttc Phe	ttt Phe 60	tgc Cys	aag Lys	ggt Gly	gct Ala	tcc Ser 65	ttt Phe	aaa Lys	aga Arg	gac Asp	tac Tyr 70	cta Leu	gcc Ala	306
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tcc ct Ser Le 17	u Thr	act Thr	cct Pro	gaa Glu	gcc Ala 175	Leu	gac Asp	cto Leu	tac Tyr	cag Gln 180	Met	att Ile	gcc	cgg Arg	642
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acc Thr 425	Glu	gat Asg	gac Asp	ccc Pro	aat Asn 430	Phe	gaa Glu	a gad 1 As <u>r</u>	gtt Val	caa L Glr 435	ı Ala	ato a Ile	tgo Cys	caa Glr	gaa Glu 440	1410

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	67/235	

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Lys Gly Lys Thr Thr Thr Ser Tyr Leu Leu Lys Ser Ile Leu Asp Gln

Asp Gln Ala Gly Lys Thr Ala Ile Ile Ser Thr Leu Gly Ile Ser Leu

Asp Gly Gln Thr Gln Glu Glu Ala Ser Leu Thr Thr Pro Glu Ala Leu 165 170

Asp Leu Tyr Gln Met Ile Ala Arg Ala Gln Asp Gln Gly Met Asp Gln 180

Leu Ile Met Glu Val Ser Ser Gln Ala Tyr Lys Met Asp Arg Val Tyr

Gly Leu Thr Phe Asp Phe Gly Ala Phe Leu Asn Ile Ser Pro Asp His 215

Ile Gly Pro Asn Glu His Pro Asp Met Glu Asp Tyr Phe Tyr Cys Lys 230 235

Ser Arg Leu Val Lys His Ser Lys Leu Ala Leu Leu Asn Ala Gly Leu

Asp Gln Leu Asp Tyr Leu Lys Asp Leu Ser Gln Lys Asn Gly Gly Gln 260

Val Gln Val Tyr Gly Gln Asp Pro Lys Thr Cys Asp Tyr Tyr Phe Glu 280 275

Val Asn Asn Gln Asp Ser Arg Arg Phe Ala Ile Lys Ser Gln Ser Pro 300

Asp Asp Leu Ala Ile Asp Gly Asp Tyr Gln Phe Glu Met Leu Gly Asp

Phe Asn Lys Glu Asn Ala Leu Cys Ala Ala Leu Ile Ala Gly His Leu 325

Glu Val Gly Gln Glu Ala Ile Tyr Gln Gly Ile Ala Gln Ala Gln Val

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340 345 350

Pro Gly Arg Met Gln His Tyr Thr Tyr Gly Asn Asn His Ile Tyr Val 355 360 365

Asp Phe Ala His Asn Tyr Ile Ser Leu Lys Asn Leu Phe Asp Phe Ala 370 380

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Gly Asn Lys Gly Val Ser Arg Arg Lys Asp Met Gly Tyr Leu Leu Ser 405 410 415

Gln Tyr Gln Gly Glu Val Ile Leu Thr Glu Asp Asp Pro Asn Phe Glu 420 425 430

Asp Val Gln Ala Ile Cys Gln Glu Ile Ala Gln Tyr Ile Asp Gly Pro 435 440 445

Ile Gln Val Thr Phe Asn Asp Asn Arg Ile Asn Ala Ile Gln Asp Leu 450 455 460

Leu Glu Ser Leu Thr Pro Glu Ser Gln Lys Val Ile Leu Leu Ala Gly 465 470 475 480

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ctc Leu	tct Ser	ggc	cac His 125	Arg	gat Asp	acc Thr	gtc Val	ttc Phe 130	Arg	gac Asp	ttt Phe	Gly	gaa Glu 135	Leu	gaa Glu	435
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gag Glu	att Ile 155	Gln	gac Asp	tat Tyr	gaa Glu	att Ile 160	Val	gac Asp	cgg Arg	gat Asp	gat Asp 165	Thr	tcc	gto Val	atc	531
cgg Arg 170	Pro	atg Met	: Gly	gaa Glu	gaa Glu 175	ı Val	tta Leu	gtg Val	gtt Val	tca Ser 180	Thr	tgc Cys	tac Tyr	e ccc	ttt Phe 185	579
gaa Glu	ttt Phe	tac Tyr	ggt Gly	ttt Phe	Ala	cct Pro	gac Asp	cgc Arg	ttt Phe 195	val	tto Phe	tate Tyr	tgt Cys	tac Tyr 200	ccc Pro )	627
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<210> 28

<211> 203

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Ala Glu Ala Gly Asp Gly Asp Gln Asp Gly Gln Asp Gly Ala Ser Asp

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Leu Asp Ile Pro Lys Leu Asp Arg Ser Ile Gly Ile Val Ala Gly Thr 85

Asp Ala Asp Ser Leu Lys Lys Gly Val Gly His Val Glu Asn Thr Val 105

Phe Pro Gly Gln Gly Glu Gln Ile Val Leu Ser Gly His Arg Asp Thr 120 115

Val Phe Arg Asp Phe Gly Glu Leu Glu Ile Gly Asp Asn Phe Ile Val 130

Gln Met Pro Tyr Gly Asp Tyr Glu Tyr Glu Ile Gln Asp Tyr Glu Ile

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gac Asp	cgg Arg 210	gta Val	gga Gly	gcc Ala	tct Ser	ctg Leu 215	acc Thr	cta Leu	att Ile	tct Ser	caa Gln 220	tcc Ser	gac Asp	ctg Leu	act Thr	672
tat Tyr 225	aac Asn	ctg Leu	act Thr	tcg Ser	gac Asp 230	ctc Leu	ttg Leu	caa Gln	gac Asp	ttt Phe 235	gaa Glu	tac Tyr	aag Lys	cag Gln	gtt Val 240	720
ccc Pro	tac Tyr	cgc Arg	att Ile	tca Ser 245	ctt Leu	tta Leu	gaa Glu	gat Asp	tat Tyr 250	caa Gln	att Ile	tac Tyr	aac Asn	gcc Ala 255	ctg Leu	768
gta Val	gca Ala	ctc Leu	gaa Glu 260	atc Ile	tct Ser	ttt Phe	gcc Ala	tta Leu 265	Gln	gat Asp	gct Ala	ggc	tgg Trp 270	cag Gln	att Ile	816
agc Ser	cct Pro	aaa Lys 275	Ala	att Ile	aaa Lys	caa Gln	ggt Gly 280	Leu	gtt Val	gag Glu	acc Thr	cgc Arg 285	TIP	ccc Pro	ggc Gly	864
cgt Arg	ttt Phe 290	Glu	ctt Leu	atc Ile	gcc Ala	tct Ser 295	His	ccg Pro	acc Thr	gtg Val	atc Ile 300	Val	gat Asp	ggg Gly	tct Ser	912
cat His 305	Asn	gaa Glu	gac Asp	ggc Gly	cto Lev	ı Gln	gct Ala	cto Lev	ttg Lev	gct Ala 315	ASD	cta Leu	gac Asp	cgc Arg	tac Tyr 320	960
ttt Phe	cca Pro	gaa Glu	caa Glr	aaa Lys 325	Arc	g att g Ile	ggg Gly	ato Ile	gta Val	F GT7	atg Met	ttg Lev	gcc Ala	gac Asp 335	aag Lys	1008
gat As <u>r</u>	gtt Val	gat L As <u>r</u>	gco Ala	a Ala	cta Le	a gct ı Ala	cet Pro	tta Lev	ı Thi	c aaa r Lys	a ago s Sei	ttt Phe	gad Ası 350	) AL	g ctt g Leu	1056
tat Ty:	ace r Thi	g gtg r Val	LThi	r Pro	Ası	t tcg p Sei	Pro	o Arg	g GL	y Mei	t Ala	a gco a Ala 36	a PIG	caa o Gli	a atg n Met	1104
aa: Ly:	a ga s Gl	u Ly:	a cto s Leo	g aco	gaa Gl	a atq u Mei 37	. Va	g tc: 1 Se:	g cc: r Pr	g tc o Se:	t act r Thi	r Arg	g gto g Vai	c ata	a gct e Ala	1152
tg Cy: 38	s Gl	a ag u Se:	t ta r Ty:	t aa r Ası	c ca n Gl: 39	n Ala	c tt a Le	a ga u As	c ct	g gc u Al 39	a GI	t caa	a gt n Va	a gc l Al	c ggc a Gly 400	1200
gg gg	a ga y As	t ga p As	c ct p Le	a at u Il	t gt e Va	c gt l Va	t tt 1 Ph	t gg e Gl	a ag y Se	t tt r Ph	t ta e Ty	t at r Il	t gt e Va	t gg l Gl	t aag y Lys	1248

74/235

405 410 415

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<211> 429

<212> PRT

<213> Alloiococcus otitidis

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Lys Met Asp Leu Gly Leu Ala Thr Met Thr Gln Val Met Asp Leu Leu 20 25 30

Gly Lys Pro Gln Asp Gln Val Pro Met Val His Ile Ala Gly Thr Asn 35 40 45

Gly Lys Gly Ser Ala Ala Ala Phe Thr Glu Arg Ile Leu Arg Glu Ala 50 55 60

Gly Tyr Lys Val Gly Leu Tyr Ile Ser Pro Ser Leu Val Glu Phe Asn 65 70 75 80

Glu Arg Ile Gln Ile Asn Gly Gln Ala Thr Ser Asp Asp Gln Leu Leu 85 90 95

Lys Ala Val Lys Thr Leu Ser Gln Ala Leu Glu Gly Thr Ser Leu Cys
100 105 110

Leu Thr Glu Phe Glu Leu Phe Thr Ala Leu Ala Phe Leu Thr Phe Gln 115  $\,$  120  $\,$  125

Asp Gln Ala Cys Asp Ile Ala Val Val Glu Val Gly Leu Gly Gly Arg 130 135 140

Leu Asp Ala Thr Asn Val Ile Ser Arg Pro Ala Val Thr Ala Ile Thr 145 150 155 160

Lys Ile Gly Met Asp His Thr Ala Phe Leu Gly Asp Ser Leu Pro Glu 165 170 175

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Ile Ala Gly Glu Lys Ala Ala Ile Ala Lys Ala Gly Ser Pro Met Val 180 185 190

Val Tyr Pro Gln Gly Pro Glu Val Thr Arg Val Ile Gln Asn Gln Ala 195 200 205

Asp Arg Val Gly Ala Ser Leu Thr Leu Ile Ser Gln Ser Asp Leu Thr 210 215 220

Tyr Asn Leu Thr Ser Asp Leu Leu Gln Asp Phe Glu Tyr Lys Gln Val 225 230 235 240

Pro Tyr Arg Ile Ser Leu Leu Glu Asp Tyr Gln Ile Tyr Asn Ala Leu 245 250 255

Val Ala Leu Glu Ile Ser Phe Ala Leu Gln Asp Ala Gly Trp Gln Ile 260 265 270

Ser Pro Lys Ala Ile Lys Gln Gly Leu Val Glu Thr Arg Trp Pro Gly 275 280 285

Arg Phe Glu Leu Ile Ala Ser His Pro Thr Val Ile Val Asp Gly Ser 290 295 300

His Asn Glu Asp Gly Leu Gln Ala Leu Leu Ala Asn Leu Asp Arg Tyr 305 310 315 320

Phe Pro Glu Gln Lys Arg Ile Gly Ile Val Gly Met Leu Ala Asp Lys 325 330 335

Asp Val Asp Ala Ala Leu Ala Pro Leu Thr Lys Ser Phe Asp Arg Leu 340 345 350

Tyr Thr Val Thr Pro Asp Ser Pro Arg Gly Met Ala Ala Pro Gln Met 355 360 365

Lys Glu Lys Leu Thr Glu Met Val Ser Pro Ser Thr Arg Val Ile Ala 370 375 380

Cys Glu Ser Tyr Asn Gln Ala Leu Asp Leu Ala Gly Gln Val Ala Gly 385 390 395

Gly Asp Asp Leu Ile Val Val Phe Gly Ser Phe Tyr Ile Val Gly Lys

76/235

405 410 415

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cag att ttt aaa gtc gac ctt gtc tta gta act gac ctc aag tta gcg Gln Ile Phe Lys Val Asp Leu Val Leu Val Thr Asp Leu Lys Leu Ala 30 35 40	147													
ggt caa aca gac aag atg ggg cac agt atc cac tac ggg gaa gtt tat Gly Gln Thr Asp Lys Met Gly His Ser Ile His Tyr Gly Glu Val Tyr 45 . 50 55 60	195													
gac ctg gtc aag tcc att gtg gaa ggt acc ccc ttt aag ctt ttg gag Asp Leu Val Lys Ser Ile Val Glu Gly Thr Pro Phe Lys Leu Glu 65 70 75	243													
tcc ttg gcg gaa acc cta gcc caa gaa gtt ctc aag act ttt gac cag Ser Leu Ala Glu Thr Leu Ala Gln Glu Val Leu Lys Thr Phe Asp Gln 80 85 90	291													
gtt gag gag gtc ttg gtc cgg gtc aac aaa ccc cag gcc ccg att cct Val Glu Val Leu Val Arg Val Asn Lys Pro Gln Ala Pro Ile Pro 95 100 105	339													
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Met Asp Lys Arg Asp Lys Ile Arg Leu Gln Gly Met Thr Phe His Gly

His His Gly Leu Met Glu Ala Glu Thr Lys Leu Gly Gln Ile Phe Lys
20 25 30

Val Asp Leu Val Leu Val Thr Asp Leu Lys Leu Ala Gly Gln Thr Asp 35 40 45

Lys Met Gly His Ser Ile His Tyr Gly Glu Val Tyr Asp Leu Val Lys 50 55 60

Ser Ile Val Glu Gly Thr Pro Phe Lys Leu Leu Glu Ser Leu Ala Glu 65 70 75 80

Thr Leu Ala Gln Glu Val Leu Lys Thr Phe Asp Gln Val Glu Glu Val
85 90 95

Leu Val Arg Val Asn Lys Pro Gln Ala Pro Ile Pro Gly Val Phe Asp 100 105 110

Asn Val Ala Val Glu Ile Thr Arg Ala Arg His

<210> 33

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<212> DNA

<213> Alloiococcus otitidis

<220>

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<400> 33

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1 5 10

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Asn Met Gly Thr Lys Leu Ala Tyr Leu Asn Arg Ala Leu Ala Lys Ile
15 20 25

aat agc cta gac cag gta gca gtc aag caa gtt tca aag gtt tac cag

Asn Ser Leu Asp Gln Val Ala Val Lys Gln Val Ser Lys Val Tyr Gln

30 35 40

act gaa ccg gtg ggc tac aag gac cag gac gat ttt tac aat atg gtt 195
Thr Glu Pro Val Gly Tyr Lys Asp Gln Asp Asp Phe Tyr Asn Met Val

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50 45 243 gct ggc ctt gaa att gaa cca ggc aag acc ccc ttg gac ctc tta gaa Ala Gly Leu Glu Ile Glu Pro Gly Lys Thr Pro Leu Asp Leu Leu Glu 60 gac ttg ctg gcg att gag gca gac ctg gac agg aag cgg acc att aaa 291 Asp Leu Leu Ala Ile Glu Ala Asp Leu Asp Arg Lys Arg Thr Ile Lys 339 aat ggc ccc cga acc att gac ttg gat gtc ttg ctg gtg gag ggt caa Asn Gly Pro Arg Thr Ile Asp Leu Asp Val Leu Leu Val Glu Gly Gln 100 387 gaa att gac cat ccc aag ctc caa gtt ccc cac cca agg ctc cag gac Glu Ile Asp His Pro Lys Leu Gln Val Pro His Pro Arg Leu Gln Asp 115 435 cgg gcc ttt gtc ttg gtc ccc ttg gct gag ttg gac ccc aac tac ctg Arg Ala Phe Val Leu Val Pro Leu Ala Glu Leu Asp Pro Asn Tyr Leu 130 483 gtt cct ggc ata gat aag aca gtt gcg gac ttg ttg gct tct tta aac Val Pro Gly Ile Asp Lys Thr Val Ala Asp Leu Leu Ala Ser Leu Asn 145 140 caa acc gac cta gca ggg gtg gag gct ttg ggt cag ttg acg aac cta 531 Gln Thr Asp Leu Ala Gly Val Glu Ala Leu Gly Gln Leu Thr Asn Leu . 160 165 155 552 tta gaa gac cgt gag gct tga Leu Glu Asp Arg Glu Ala 175

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<212> PRT

<213> Alloiococcus otitidis

<400> 34

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Ala Tyr Leu Asn Arg Ala Leu Ala Lys Ile Asn Ser Leu Asp Gln Val 20 25 30

Ala Val Lys Gln Val Ser Lys Val Tyr Gln Thr Glu Pro Val Gly Tyr 35 40 45

Lys Asp Gln Asp Asp Phe Tyr Asn Met Val Ala Gly Leu Glu Ile Glu 50 55 60

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Pro Gly Lys Thr Pro Leu Asp Leu Leu Glu Asp Leu Leu Ala Ile Glu

Ala Asp Leu Asp Arg Lys Arg Thr Ile Lys Asn Gly Pro Arg Thr Ile 85 90 95

Asp Leu Asp Val Leu Leu Val Glu Gly Gln Glu Ile Asp His Pro Lys 100 105 110

Leu Gln Val Pro His Pro Arg Leu Gln Asp Arg Ala Phe Val Leu Val 115 120 125

Pro Leu Ala Glu Leu Asp Pro Asn Tyr Leu Val Pro Gly Ile Asp Lys 130 135 140

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<211> 1242

<212> DNA

<213> Alloiococcus otitidis

<220>

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Phe Arg Arg Phe Asn Met Gln Ile Gly Ile Asp Lys Leu Ala Phe Ala
10 15 20

act cca acc agg tac ttg gaa atg gcg agt ctg gcc caa gcc cgg tcc

Thr Pro Thr Arg Tyr Leu Glu Met Ala Ser Leu Ala Gln Ala Arg Ser

25

30

35

caa gac cct aat aaa tat atc aag ggg cta ggc caa gaa gcc atg gct

Gln Asp Pro Asn Lys Tyr Ile Lys Gly Leu Gly Gln Glu Ala Met Ala

40

45

			gaa Glu													246
			tta Leu	_	_	_	-		-			-	_			294
			gaa Glu													342
		_	ctg Leu 105	_						_	_	_	_			390
			tgc Cys			_	_	-					_		_	438
			tta Leu													486
			tat Tyr													534
			gcc Ala			_	-	_	_		-		_		-	582
			agt Ser 185													630
			agc Ser													678
			tcc Ser													726
			tct Ser													774
			atg Met													822
gac Asp	cag Gln	aaa Lys	gag Glu 265	att Ile	gac Asp	cgc Arg	ttg Leu	ctg Leu 270	gcc Ala	tat Tyr	tac Tyr	gag Glu	cct Pro 275	ggt Gly	cgc Arg	870

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agt ttg att tcc ctc tta gac cag gta agt gac ctg gag gct ggc gac Ser Leu Ile Ser Leu Leu Asp Gln Val Ser Asp Leu Glu Ala Gly Asp 295 300 305	966
cgg att ggc ctc tat tct tat ggg tct ggt gcc gtt gga gag ttc ttt Arg Ile Gly Leu Tyr Ser Tyr Gly Ser Gly Ala Val Gly Glu Phe Phe 310 315 320 325	1014
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ttc gac cag gtt gtc aac cag cgt tca gca tta gag atg tac agc tat Phe Asp Gln Val Val Asn Gln Arg Ser Ala Leu Glu Met Tyr Ser Tyr 345 350 355	1110
cag gac ttg ctg acc ttt agc cta cct caa gac ggc caa act tac act Gln Asp Leu Leu Thr Phe Ser Leu Pro Gln Asp Gly Gln Thr Tyr Thr 360 365 370	1158
aca gat aaa agt cac cag gtc cca ggc cgt ttt gtc tta gac cgg gtg Thr Asp Lys Ser His Gln Val Pro Gly Arg Phe Val Leu Asp Arg Val 375 380 385	1206
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Ala Gln Ala Arg Ser Gln Asp Pro Asn Lys Tyr Ile Lys Gly Leu Gly 35 40 45	
Gln Glu Ala Met Ala Val Pro Glu Glu Ser Asp Asp Ala Val Ser Leu 50 55 60	
Ala Ala Asn Ala Gly Asn Leu Ile Leu Ser Glu Glu Asp Lys Ala Ala 65 70 75 80	

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Ile Asp Met Val Ile Val Gly Thr Glu Ser Gly Val Asp Gln Ser Lys

Ser Ala Ala Ser Trp Val His Asp Leu Leu Gly Ile Asn Pro His Ala 105 100

Arg Ser Leu Glu Ile Lys Gln Ala Cys Tyr Gly Ala Thr Ala Gly Leu . 115 120 125

Lys Leu Ala Val Ala His Leu Ala Leu Asn Pro Asp Ser Lys Val Leu 130 135

Val Ile Gly Ser Asp Ile Ala Lys Tyr Gly Leu Glu Thr Gly Gly Glu 150 145

Pro Thr Gln Gly Ala Gly Ala Val Ala Ile Leu Val Ser Arg Asp Pro 165 170

Ala Ile Ala Val Val Asn Asn Asp Ser Ala Met Leu Thr Lys Asn Ile 185

Ala Asp Phe Trp Arg Pro Asn Tyr Ser Asp Tyr Ala His Val Asp Gly 200 . 205

Lys Phe Ser Asn Gln Ala Tyr Leu Ser Asn Leu Ala Glu Val Trp Arg 215 210

Gln Tyr Lys Ile Lys Asn Gln Leu Ser Ala Lys Asp Phe Lys Ala Met 225 230 235

Val Phe His Ser Pro Tyr Thr Lys Met Gly Lys Lys Ala Leu Leu Lys

Leu Gly Asp Tyr Glu Asp Gln Lys Glu Ile Asp Arg Leu Leu Ala Tyr

Tyr Glu Pro Gly Arg Tyr Tyr Asn Lys Arg Val Gly Asn Ile Tyr Thr 280

Gly Ser Leu Tyr Leu Ser Leu Ile Ser Leu Leu Asp Gln Val Ser Asp

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Leu Glu Ala Gly Asp Arg Ile Gly Leu Tyr Ser Tyr Gly Ser Gly Ala 315 305 310 Val Gly Glu Phe Phe Ser Ile Arg Leu Gln Pro Gly Tyr Lys Glu Ser 325 Leu Gln Gln Val Asp Phe Asp Gln Val Val Asn Gln Arg Ser Ala Leu Glu Met Tyr Ser Tyr Gln Asp Leu Leu Thr Phe Ser Leu Pro Gln Asp 360 Gly Gln Thr Tyr Thr Thr Asp Lys Ser His Gln Val Pro Gly Arg Phe Val Leu Asp Arg Val Ala Asp His Ile Arg Tyr Tyr Arg Arg Leu Ala 400 390 <210> 37 <211> 1323 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (31)..(1323) <223> <400> 37 ttctggtata gattaaggaa ggaggagacc atg tta ccc tta ttc aag caa ttt Met Leu Pro Leu Phe Lys Gln Phe tac aag caa agc ctc agc cag cgc ctc aaa gct cta gaa aag gcc ggc 102 Tyr Lys Gln Ser Leu Ser Gln Arg Leu Lys Ala Leu Glu Lys Ala Gly 150 tat ctt gat cct gac cag gcg ggt aaa ctc cag tca ggg gaa ctg ggt Tyr Leu Asp Pro Asp Gln Ala Gly Lys Leu Gln Ser Gly Glu Leu Gly 35 ttg acc cat gaa gcc ggc gac cac atg att gaa aac tac atc ggc tcc 198

Leu Thr His Glu Ala Gly Asp His Met Ile Glu Asn Tyr Ile Gly Ser

tat acc ctc cct ctg gga ctg gcc ctc cac ttt tta ctc gat ggc aag

Tyr Thr Leu Pro Leu Gly Leu Ala Leu His Phe Leu Leu Asp Gly Lys

246

			60					65					70			
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gcc Ala	agc Ser 90	aac Asn	ggt Gly	gcc Ala	aag Lys	atg Met 95	gta Val	gcc Ala	caa Gln	agc Ser	ggt Gly 100	ggt Gly	ttc Phe	cat His	aca Thr	342
gtc Val 105	aag Lys	gaa Glu	aac Asn	cgg Arg	ctg Leu 110	atg Met	atc Ile	ggt Gly	caa Gln	gtg Val 115	gtc Val	ata Ile	gcc Ala	gga Gly	agc Ser 120	390
aca Thr	aaa Lys	cct Pro	agc Ser	cag Gln 125	gac Asp	cgg Arg	gga Gly	aaa Lys	atc Ile 130	ctg Leu	agc Ser	cac His	cag Gln	caa Gln 135	gac Asp	438
tta Leu	atc Ile	gac Asp	cta Leu 140	gcc Ala	aat Asn	gct Ala	agc Ser	tat Tyr 145	ccc Pro	tca Ser	att Ile	ggt Gly	aaa Lys 150	aga Arg	GJA aaa	486
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gaa Glu 185	Ala	atg Met	ggg	gct Ala	aac Asn 190	att Ile	atc Ile	aac Asn	acc Thr	atg Met 195	Leu	gaa Glu	gcc Ala	ctg Leu	gct Ala 200	630 <sup>-</sup>
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tct Ser	aac Asn	ctg Leu	gco Ala 220	thr	gaa Glu	tcc Ser	ctt	gto Val 225	. Thr	gtt Val	tcc Ser	tgt Cys	caa Gln 230	vaı	aaa Lys	726
ccc Pro	aga Arg	ttt Phe 235	e Let	a gto 1 Val	aaa Lys	aat Asn	gac Asr 240	) Met	gca Ala	GJ7 Gg3	gaa Glu	a gct ı Ala 245	f Agr	cgg Arg	g Asp	774
caa Glr	a ato 1 Ile 250	e Ile	c caq e Gli	g gco n Ala	tac Tyl	cag Glr 255	t Ty	gco Ala	tgo a Cys	c cto s Lev	g gad 1 Asp 260	o Pro	tac Tyr	cgg Arg	g gca g Ala	822
gco Ala 26	a Thi	c cac	c aac s Asi	c aag	g ggg s Gly 270	y Ile	ato Me	g aad C Asi	e ggg	g gta y Val 27:	l As	c ggo p Gly	ttq y Le	g gto ı Va	c cta l Leu 280	870
gc	t ag	t ggg	gaa yAs	t ga n As; 28	рТП	g cgg	g gca g Ala	a ato a Ilo	c gaa e Gli 29	ı Ali	g gg a Gl	g gc	c cat	t gct s Ala 29	t tac a Tyr 5	918

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caa Gln	gac Asp	gga Gly 315	cag Gln	tta Leu	aaa Lys	GJÀ āāā	acc Thr 320	att Ile	acc Thr	ctt Leu	ccc Pro	ttg Leu 325	cca Pro	att Ile	gcc Ala	1014	
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<211> 430

<212> PRT <213> Alloiococcus otitidis

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Lys Leu Gln Ser Gly Glu Leu Gly Leu Thr His Glu Ala Gly Asp His

Met Ile Glu Asn Tyr Ile Gly Ser Tyr Thr Leu Pro Leu Gly Leu Ala

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55 60 50 Leu His Phe Leu Leu Asp Gly Lys Ser Tyr Leu Val Pro Met Ala Ile 70 Glu Glu Pro Ser Val Ile Ala Ala Ala Ser Asn Gly Ala Lys Met Val Ala Gln Ser Gly Gly Phe His Thr Val Lys Glu Asn Arg Leu Met Ile 105 100 Gly Gln Val Val Ile Ala Gly Ser Thr Lys Pro Ser Gln Asp Arg Gly 125 Lys Ile Leu Ser His Gln Gln Asp Leu Ile Asp Leu Ala Asn Ala Ser Tyr Pro Ser Ile Gly Lys Arg Gly Gly Gly Ala Arg Gly Ile Gln Val 150 145 Lys Gln Phe Asp Ser Asp Leu Gly Gln Asp Met Gly Ser Tyr Leu Ala 165 170 Val Tyr Leu Thr Val Asp Cys Gln Glu Ala Met Gly Ala Asn Ile Ile Asn Thr Met Leu Glu Ala Leu Ala Pro Glu Ile Asp Arg Leu Thr Ser 200 Gly Gln Val Leu Met Ser Ile Leu Ser Asn Leu Ala Thr Glu Ser Leu 215 Val Thr Val Ser Cys Gln Val Lys Pro Arg Phe Leu Val Lys Asn Asp 230 235 Met Ala Gly Glu Ala Val Arg Asp Gln Ile Ile Gln Ala Tyr Gln Tyr Ala Cys Leu Asp Pro Tyr Arg Ala Ala Thr His Asn Lys Gly Ile Met 260 Asn Gly Val Asp Gly Leu Val Leu Ala Ser Gly Asn Asp Trp Arg Ala

280

285

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Ile Glu Ala Gly Ala His Ala Tyr Ala Ser Leu Thr Gly His Tyr Arg

Pro Leu Ser Lys Trp Glu Lys Thr Gln Asp Gly Gln Leu Lys Gly Thr 310

Ile Thr Leu Pro Leu Pro Ile Ala Thr Val Gly Gly Ala Ile Ala Ser 330

His Pro Val Ala Gln Val Ser Gln Gln Ile Leu Gly Gln Pro Thr Ala 345

Lys Gln Leu Ala Arg Leu Val Ala Ala Val Gly Leu Ala Gln Asn Leu 355

Ser Ala Leu Arg Ala Leu Val Thr Thr Gly Ile Gln Gln Gly His Met 375

Ala Leu Gln Ala Arg Ser Leu Ala Met Asn Ala Gly Ala Arg Gly Asp 390 395

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<222> (13)..(930)

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gtg Val	acg Thr	att Ile	atc Ile	aag Lys 50	tgg Trp	atc Ile	aac Asn	caa Gln	agt Ser 55	ccg Pro	gaa Glu	tac Tyr	caa Gln	gct Ala 60	ggc Gly	1 <b>9</b> 5
gat Asp	ctc Leu	ccc Pro	ctc Leu 65	act Thr	atc Ile	tta Leu	Gjà gàc	aat Asn 70	gct Ala	agc Ser	aac Asn	ctg Leu	atc Ile 75	gta Val	aaa Lys	243
gat Asp	ggt Gly	80 GJA āāā	ata Ile	aga Arg	GJÀ āāā	att Ile	acc Thr 85	atc Ile	att Ile	acc Thr	acc Thr	ggc 90	att Ile	aaa Lys	acc Thr	291
att Ile	tgt Cys 95	cac His	gaa Glu	gag Glu	aac Asn	cgg Arg 100	atc Ile	act Thr	gcg Ala	ggc Gly	gct Ala 105	gga Gly	gca Ala	gct Ala	att Ile	339
atc Ile 110	gat Asp	gtt Val	agc Ser	cag Gln	gct Ala 115	gcc Ala	ttg Leu	gac Asp	cat His	agc Ser 120	tta Leu	act Thr	Gly	ttg Leu	gaa Glu 125	387
ttc Phe	gct Ala	tgt Cys	ggc	ata Ile 130	Pro	ggt Gly	agt Ser	aca Thr	ggc Gly 135	GJA aaa	gct Ala	gtt Val	tac Tyr	atg Met 140	Asn	435
gct Ala	GJA aaa	gct Ala	tac Tyr 145	Gly	Gly	gaa Glu	gtc Val	cag Gln 150	His	tgt Cys	gtt Val	gaa Glu	agt Ser 155	gtc Val	caa Gln	483
gtc Val	ttg Leu	acc Thr 160	Arg	cat His	ggc	cag Gln	ttg Leu 165	aag Lys	acc	tat Tyr	agt Ser	aat Asn 170	Ala	gaa Glu	atg Met	531
aac Asn	ttc Phe 175	Ser	tac Tyr	cgc Arg	cac His	agt Ser 180	Tyr	ttg Leu	atg Met	gaa Glu	gaa Glu 185	Asp	gat Asp	ata Ile	gta Val	579
gto Val 190	Ser	gtg Val	acc Thr	ttt Phe	aaa Lys 195	Lev	gag Glu	tcg	ggc Gly	gac Asp 200	туг	ato	act Thr	ato	aag Lys 205	627
gaa Glu	aaç Lys	ato Met	gat Asp	gaa Glu 210	ı Lev	a acc	tac Tyr	ctt Leu	aga Arg 215	Glu	tco Ser	aaa Lys	a caa s Glr	ccg Pro 220	g ctg Leu )	675
gaa Glu	tac Tyr	Pro	tct Ser 225	: Суз	ggg Gly	g tca / Ser	gtc Val	ttt Phe	Lys	aga Arg	cct Pro	gaa Glu	ggd Gly 235	, His	ttt Phe	723
acç Thi	. GJ7	aaa Lys 240	Leu	a ato ı Ile	c cag e Glr	g gat n Asp	gct Ala 245	Gly	ctt Leu	caa 1 Glr	a gga a Gly	a ttg / Let 250	ı Val	cat L His	ggt Gly	771
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Gly Ala Gln Val Ser Glu Lys His Ala Gly Phe Ile Ile Asn Ile Gly
255 260 265

aat gct acc gcc agc gac tac caa gag ttg atc caa cat atc caa gaa
Asn Ala Thr Ala Ser Asp Tyr Gln Glu Leu Ile Gln His Ile Gln Glu
270 280 285

gaa gtc tac cgg att tac aag gtt aag ctg gaa cgt gaa gtt cgc att

Glu Val Tyr Arg Ile Tyr Lys Val Lys Leu Glu Arg Glu Val Arg Ile

290

295

300

ata ggg gag gat tag 930
Ile Gly Glu Asp

<210> 40 <211> 305 <212> PRT <213> Alloiococcus otitidis

305

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Ala Asp Ile Leu Ile Phe Pro Glu Ser Ile Asp Glu Ile Val Thr Ile 35 40 45

Ile Lys Trp Ile Asn Gln Ser Pro Glu Tyr Gln Ala Gly Asp Leu Pro 50 60

Leu Thr Ile Leu Gly Asn Ala Ser Asn Leu Ile Val Lys Asp Gly Gly 65 70 75 80

Ile Arg Gly Ile Thr Ile Ile Thr Thr Gly Ile Lys Thr Ile Cys His 90 95

Glu Glu Asn Arg Ile Thr Ala Gly Ala Gly Ala Ile Ile Asp Val 100 105 110

Ser Gln Ala Ala Leu Asp His Ser Leu Thr Gly Leu Glu Phe Ala Cys 115 120 125

Gly Ile Pro Gly Ser Thr Gly Gly Ala Val Tyr Met Asn Ala Gly Ala 130 135 140

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Tyr Gly Gly Glu Val Gln His Cys Val Glu Ser Val Gln Val Leu Thr 150 145

Arg His Gly Gln Leu Lys Thr Tyr Ser Asn Ala Glu Met Asn Phe Ser 170

Tyr Arg His Ser Tyr Leu Met Glu Glu Asp Asp Ile Val Val Ser Val 185

Thr Phe Lys Leu Glu Ser Gly Asp Tyr Ile Thr Ile Lys Glu Lys Met 195

Asp Glu Leu Thr Tyr Leu Arg Glu Ser Lys Gln Pro Leu Glu Tyr Pro 215

Ser Cys Gly Ser Val Phe Lys Arg Pro Glu Gly His Phe Thr Gly Lys 235 230

Leu Ile Gln Asp Ala Gly Leu Gln Gly Leu Val His Gly Gly Ala Gln 245

Val Ser Glu Lys His Ala Gly Phe Ile Ile Asn Ile Gly Asn Ala Thr 265 260

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Arg Ile Tyr Lys Val Lys Leu Glu Arg Glu Val Arg Ile Ile Gly Glu 295

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<222> (16)..(1104)

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1 5 10

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ctg Leu	aca Thr 30	gtc Val	agc Ser	cgg Arg	tat Tyr	ttg Leu 35	act Thr	tta Leu	gac Asp	att Ile	tgg Trp 40	gaa Glu	aca Thr	tct Ser	ccc Pro	14	7
gac Asp 45	caa Gln	gct Ala	tca Ser	gtc Val	agg Arg 50	tct Ser	caa Gln	aca Thr	tat Tyr	ggc Gly 55	aac Asn	cag Gln	gcc Ala	tat Tyr	gct Ala 60	19	5
tgg Trp	gag Glu	cgg Arg	tta Leu	gat Asp 65	ggt Gly	atc Ile	ttt Phe	agc Ser	ttt Phe 70	aag Lys	gac Asp	tgg Trp	tcc Ser	cac His 75	ccc Pro	24	3
ttc Phe	cac His	cta Leu	gtc Val 80	gaa Glu	acg Thr	gtg Val	atc Ilė	caa Gln 85	aca Thr	gtg Val	gaa Glu	gcc Ala	tac Tyr 90	ata Ile	gaa Glu	29	1
tcc Ser	ttg Leu	tcc Ser 95	ctg Leu	cct Pro	tta Leu	aaa Lys	agt Ser 100	tac Tyr	Gly aaa	att Ile	cag Gln	atc Ile 105	aag Lys	agc Ser	cag Gln	33	9
ttg Leu	gac Asp 110	tac Tyr	cag Gln	Gly	aaa Lys	aaa Lys 115	att Ile	ggc Gly	ctg Leu	GJÀ aaa	tct Ser 120	agt Ser	Gly ggg	gcc Ala	gtt Val	38	; <b>7</b>
acc Thr 125	Ile	gct Ala	gtt Val	atc Ile	cga Arg 130	ggc	ctg Leu	agc Ser	ctt Leu	ctt Leu 135	tac Tyr	gac Asp	ctc Leu	cac His	tta Leu 140	43	15
aaa Lys	gac Asp	ata Ile	gac Asp	att Ile 145	Phe	aaa Lys	cta Leu	gct Ala	gcc Ala 150	Ile	gcc Ala	cat His	atc Ile	cag Gln 155	cta Leu	48	}3
aag Lys	agc Ser	aag Lys	ggg Gly 160	Ser	ttt Phe	ggc	gat Asp	ttg Leu 165	gca Ala	gcc Ala	tgc Cys	act Thr	tat Tyr 170	Thr	ggt Gly	53	31
gtg Val	atc Ile	cgc Arg 175	Tyr	cag Gln	tcc Ser	ctg Leu	gat Asp 180	Arg	gaa Glu	tgg Trp	tta Leu	caa Gln 185	Glu	caa Gln	atc Ile	57	79
tcc Ser	aac Asn 190	His	tcc Ser	ato	aag Lys	gac Asp 195	Leu	ctg Leu	gcc	atg Met	gat Asp 200	Trp	cct Pro	agc Ser	cta Leu	62	27
ggt Gly 205	Leu	gac Asp	cgg Arg	cto Leu	ago Ser 210	Leu	ccc Pro	cat His	gac	cto Leu 215	Arg	ctt Leu	tta Leu	ato Ile	gga Gly 220	61	75
tgg Trp	acc Thr	ggc	cag Gln	cct	gcc Ala	tcc Ser	aca Thr	gaa Glu	aaa Lys	ttg Lev	gtt Val	cag Glr	gct Ala	gto Val	tac Tyr	72	23

92/235

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	225	230	235
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gac tcc cag gca Asp Ser Gln Ala 270	agc tta gct tgg Ser Leu Ala Trp 275	atc caa aag aac cga acc Ile Gln Lys Asn Arg Thr 280	ctc ctc 867 Leu Leu
aag gca atg ggc Lys Ala Met Gly 285	caa agc cgg ggg Gln Ser Arg Gly 290	aaa gtc atc gaa acc aaa Lys Val Ile Glu Thr Lys 295	gcc ttg 915 Ala Leu 300
acc tac ttg tgc Thr Tyr Leu Cys	gat att gtc gcg Asp Ile Val Ala 305	aaa tac gga ggc caa gcc Lys Tyr Gly Gly Gln Ala 310	aag tct 963 Lys Ser 315
tcc ggt gcc ggc Ser Gly Ala Gly 320	Gly Gly Asp Cys	ggc att ggc cta atc aca Gly Ile Gly Leu Ile Thr 325 330	Arg Giu
agc cca ata gaa Ser Pro Ile Glu 335	n gcc atc tac cgg n Ala Ile Tyr Arg 340	gaa tgg atg gat gca ggt Glu Trp Met Asp Ala Gly 345	atc ttg 1059 Ile Leu
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Arg Tyr Leu Th 35	r Leu Asp Ile Trp 40	Glu Thr Ser Pro Asp Gli 45	n Ala Ser

Val Arg Ser Gln Thr Tyr Gly Asn Gln Ala Tyr Ala Trp Glu Arg Leu 50 55 60

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Asp Gly Ile Phe Ser Phe Lys Asp Trp Ser His Pro Phe His Leu Val 70 Glu Thr Val Ile Gln Thr Val Glu Ala Tyr Ile Glu Ser Leu Ser Leu 90 Pro Leu Lys Ser Tyr Gly Ile Gln Ile Lys Ser Gln Leu Asp Tyr Gln 105 Gly Lys Lys Ile Gly Leu Gly Ser Ser Gly Ala Val Thr Ile Ala Val 120 115 Ile Arg Gly Leu Ser Leu Leu Tyr Asp Leu His Leu Lys Asp Ile Asp 135 140 Ile Phe Lys Leu Ala Ala Ile Ala His Ile Gln Leu Lys Ser Lys Gly Ser Phe Gly Asp Leu Ala Ala Cys Thr Tyr Thr Gly Val Ile Arg Tyr Gln Ser Leu Asp Arg Glu Trp Leu Gln Glu Gln Ile Ser Asn His Ser 185 Ile Lys Asp Leu Leu Ala Met Asp Trp Pro Ser Leu Gly Leu Asp Arg 200 205 Leu Ser Leu Pro His Asp Leu Arg Leu Leu Ile Gly Trp Thr Gly Gln 210 215 Pro Ala Ser Thr Glu Lys Leu Val Gln Ala Val Tyr Pro Gln Lys Ile

Thr Arg Thr Pro Leu Asp Phe Gln Ser Phe Leu Asp Gln Ser Gln Glu 250 245

Cys Val Asp Gly Leu Val Glu Ser Leu Ser Gln Ala Asp Ser Gln Ala 265

Ser Leu Ala Trp Ile Gln Lys Asn Arg Thr Leu Leu Lys Ala Met Gly 280

Gln Ser Arg Gly Lys Val Ile Glu Thr Lys Ala Leu Thr Tyr Leu Cys

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95

300 295 290 Asp Ile Val Ala Lys Tyr Gly Gly Gln Ala Lys Ser Ser Gly Ala Gly 305 310 Gly Gly Asp Cys Gly Ile Gly Leu Ile Thr Arg Glu Ser Pro Ile Glu 325 Ala Ile Tyr Arg Glu Trp Met Asp Ala Gly Ile Leu Pro Leu Arg Leu 345 Asp Ile Val Glu Asn Gly Ala Cys Tyr Asp <210> 43 <211> 1023 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (13)..(1023) <223> <400> 43 gagaagccaa cc atg act aag cag gcc ttt gaa aag aaa aag tta ggc cgg 51 Met Thr Lys Gln Ala Phe Glu Lys Lys Lys Leu Gly Arg att tgc cgg gcc cat acc aac att gcc ttg atc aag tac tgg ggt aag 99 Ile Cys Arg Ala His Thr Asn Ile Ala Leu Ile Lys Tyr Trp Gly Lys 25 gct gat agg gac ttg att atc ccc aat aac aac tcc cta tct tta acc 147 Ala Asp Arg Asp Leu Ile Ile Pro Asn Asn Asn Ser Leu Ser Leu Thr 35 195 ttg gac gct ttt tat acc gat acc cag gta gtt ttt gac cca gac ttg Leu Asp Ala Phe Tyr Thr Asp Thr Gln Val Val Phe Asp Pro Asp Leu 55 gac cag gac caa tta tgg cta gac ggg aaa cag gaa aaa ggg tcc gcc 243 Asp Gln Asp Gln Leu Trp Leu Asp Gly Lys Gln Glu Lys Gly Ser Ala 70 tta acc aag gcc cag gtc atc ctg gac ttg gtt cgg gac caa gcc cag 291 Leu Thr Lys Ala Gln Val Ile Leu Asp Leu Val Arg Asp Gln Ala Gln 80 ctt gac tgg ccg gcc aaa att acc agc cac aac caa gtt gcc act gca 339 Leu Asp Trp Pro Ala Lys Ile Thr Ser His Asn Gln Val Ala Thr Ala

105

100 .

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387 Ala Gly Leu Ala Ser Ser Ala Ser Gly Leu Ala Ala Leu Ala Gly Ala 120 115 tca get gat get tta gac ett gge eta tee eea aet gae ete tee ega 435 Ser Ala Asp Ala Leu Asp Leu Gly Leu Ser Pro Thr Asp Leu Ser Arg 130 ttg gcc cgc agg gga tct ggg tct gcc tca cga agt att ttt ggt ggt 483 Leu Ala Arg Arg Gly Ser Gly Ser Ala Ser Arg Ser Ile Phe Gly Gly 150 145 ttt gtc gag tgg gaa aag ggt cat gat gat agc tct tcc ttt gcc aag 531 Phe Val Glu Trp Glu Lys Gly His Asp Asp Ser Ser Phe Ala Lys 165 579 ccc atc gac ttg gcc cag tgg gat att gcc atg ctc ttt gtc att gta Pro Ile Asp Leu Ala Gln Trp Asp Ile Ala Met Leu Phe Val Ile Val 180 175 627 age gae ega eca aag gea att tee tee age caa gge atg caa ttg ace Ser Asp Arg Pro Lys Ala Ile Ser Ser Ser Gln Gly Met Gln Leu Thr 195 cag gag acg tcg gac ttt tac cag gcc tgg tta gac agc ctg gac caa 675 Gln Glu Thr Ser Asp Phe Tyr Gln Ala Trp Leu Asp Ser Leu Asp Gln 215 210 gac cta gca gac atc aag tcc gct atc caa gcc caa gac ctc gac cag 723 Asp Leu Ala Asp Ile Lys Ser Ala Ile Gln Ala Gln Asp Leu Asp Gln 230 225 gtt ggg tcc att gca gaa aga aat gcc ctg aaa atg cat gcc acc aac 771 Val Gly Ser Ile Ala Glu Arg Asn Ala Leu Lys Met His Ala Thr Asn 245 240 ctg gca gcc aag ccc ccc ttc acc tat tgg act aaa gaa agt tta gcc 819 Leu Ala Ala Lys Pro Pro Phe Thr Tyr Trp Thr Lys Glu Ser Leu Ala 260 255 ctg atg cag gaa gta tgg gac cgg cgc aag gct ggc cag tcc ctc tac 867 Leu Met Gln Glu Val Trp Asp Arg Arg Lys Ala Gly Gln Ser Leu Tyr ttc acc atg gac gcc ggc ccc aat gtc aag gtt att ggc agg gaa gct 915 Phe Thr Met Asp Ala Gly Pro Asn Val Lys Val Ile Gly Arg Glu Ala 295 gac ctt aaa gcc ttc aaa gca gac ctc agc caa gac tgg ccc gac aag 963 Asp Leu Lys Ala Phe Lys Ala Asp Leu Ser Gln Asp Trp Pro Asp Lys 310 1011 cat ctt gtc tta gct aaa ccg ggt cca ggc ctg gcc ttt att gat gga His Leu Val Leu Ala Lys Pro Gly Pro Gly Leu Ala Phe Ile Asp Gly 330 325

1023

cct ttg aac tag Pro Leu Asn 335

<210> 44 <211> 336 <212> PRT <213> Alloiococcus otitidis

ZZISZ AITOTOCOCCUS COLUL

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Asp Leu Ile Ile Pro Asn Asn Asn Ser Leu Ser Leu Thr Leu Asp Ala 35 40

Phe Tyr Thr Asp Thr Gln Val Val Phe Asp Pro Asp Leu Asp Gln Asp 50 55 60

Gln Leu Trp Leu Asp Gly Lys Gln Glu Lys Gly Ser Ala Leu Thr Lys
70 75 80

Ala Gln Val Ile Leu Asp Leu Val Arg Asp Gln Ala Gln Leu Asp Trp 85 90 95

Pro Ala Lys Ile Thr Ser His Asn Gln Val Ala Thr Ala Ala Gly Leu 100 105 110

Ala Ser Ser Ala Ser Gly Leu Ala Ala Leu Ala Gly Ala Ser Ala Asp

Ala Leu Asp Leu Gly Leu Ser Pro Thr Asp Leu Ser Arg Leu Ala Arg 130 135 140

Arg Gly Ser Gly Ser Ala Ser Arg Ser Ile Phe Gly Gly Phe Val Glu 145 150 155 160

Trp Glu Lys Gly His Asp Asp Ser Ser Ser Phe Ala Lys Pro Ile Asp 165 170 175

Leu Ala Gln Trp Asp Ile Ala Met Leu Phe Val Ile Val Ser Asp Arg 180 185 190 97/235

Pro Lys Ala Ile Ser Ser Ser Gln Gly Met Gln Leu Thr Gln Glu Thr 200

Ser Asp Phe Tyr Gln Ala Trp Leu Asp Ser Leu Asp Gln Asp Leu Ala 215 220 210

Asp Ile Lys Ser Ala Ile Gln Ala Gln Asp Leu Asp Gln Val Gly Ser 230 235 240

Ile Ala Glu Arg Asn Ala Leu Lys Met His Ala Thr Asn Leu Ala Ala 250 245

Lys Pro Pro Phe Thr Tyr Trp Thr Lys Glu Ser Leu Ala Leu Met Gln

Glu Val Trp Asp Arg Arg Lys Ala Gly Gln Ser Leu Tyr Phe Thr Met 285 275 280

Asp Ala Gly Pro Asn Val Lys Val Ile Gly Arg Glu Ala Asp Leu Lys 295

Ala Phe Lys Ala Asp Leu Ser Gln Asp Trp Pro Asp Lys His Leu Val 315

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102

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tat Tyr	aac Asn	atg Met	ccg Pro	gcc Ala 30	atc Ile	gcc Ala	ctc Leu	cct Pro	ttt Phe 35	acc Thr	aca Thr	gcc Ala	acc Thr	atc Ile 40	acc Thr	150
gtt Val	gaa Glu	gtt Val	agt Ser 45	cct Pro	tac Tyr	caa Gln	ggc Gly	aaa Lys 50	agc Ser	tat Tyr	cta Leu	gaa Glu	agt Ser 55	gct Ala	tgc Cys	198
tac Tyr	tgc Cys	gga Gly 60	tct Ser	tta Leu	gac Asp	caa Gln	gcg Ala 65	ccc Pro	Gly ggg	gac Asp	ttg Leu	gca Ala 70	GJA āāā	ctt Leu	caa Gln	246
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ttg Leu 90	tat Tyr	atc Ile	aag Lys	gtc Val	gac Asp 95	agc Ser	atg Met	atc Ile	cct Pro	gct Ala 100	gaa Glu	aga Arg	gga Gly	atg Met	ggg Gly 105	342
tcc Ser	agt Ser	gct Ala	gct Ala	gtg Val 110	gcc Ala	acc Thr	gcc Ala	tta Leu	gtc Val 115	aag Lys	gcc Ala	ctc Leu	ttt Phe	cac His 120	tac Tyr	390
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gcc Ala	gaa Glu	aaa Lys 140	att Ile	acc Thr	cat His	ggc Gly	aag Lys 145	cca Pro	tcg Ser	ggt Gly	ctg Leu	gat Asp 150	gct Ala	aca Thr	gtc Val	486
gtc Val	aac Asn 155	Ser	att Ile	gcc Ala	ccc Pro	gtt Val 160	tat Tyr	ttt Phe	aaa Lys	cgc Arg	aac Asn 165	Gln	ctt Leu	ccc Pro	aag Lys	534
gcc Ala 170	Ile	cct Pro	tta Leu	aat Asn	gtt Val 175	gac Asp	ggc	tat Tyr	tta Leu	att Ile 180	gca Ala	gcc Ala	gat Asp	act Thr	ggg Gly 185	582
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gaa Glu	act Thr	gcc Ala	aag Lys 205	Val	caa Gln	acc Thr	atg Met	gac Asp 210	Ile	gtc Val	cac	cac His	ctc Leu 215	Gly	cag Gln	678
ctt Leu	acc Thr	cac His	Gln	gct Ala	aaa Lys	aaa Lys	gca Ala 225	Ile	atg Met	acc Thr	aat Asn	aac Asn 230	Leu	cct Pro	ggc	726
tta Leu	ggg	gag Glu	att Ile	ttg Leu	aac Asn	cag Gln	tcc Ser	cac His	caa Gln	ctc Leu	tta Lev	aag Lys	gat Asp	tta Leu	act Thr	774

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	235					240					245					
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gga Gly	gct Ala	tgc Cys	gga Gly	gct Ala 270	aag Lys	tta Leu	acc Thr	ggt Gly	ggg Gly 275	ggc Gly	cgg Arg	ggt Gly	ggt Gly	tgc Cys 280	atg Met	870
att Ile	gcc Ala	cta Leu	gcc Ala 285	caa Gln	agc Ser	aac Asn	cag Gln	gat Asp 290	gcc Ala	tcc Ser	aat Asn	att Ile	gcc Ala 295	caa Gln	aaa Lys	918
ttg Leu	gaa Glu	aaa Lys 300	gcg Ala	gga Gly	gcc Ala	att Ile	gaa Glu 305	acc Thr	tgg Trp	atc Ile	cac His	ccc Pro 310	tta Leu	gga Gly	gaa Glu	966
_	aac Asn 315			taa												981
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Ala Leu Val Lys Ala Leu Phe His Tyr Phe Gln Val Asp Leu Ser Ser 120

Glu Ala Leu Ser Ala Tyr Val Glu Ile Ala Glu Lys Ile Thr His Gly

Lys Pro Ser Gly Leu Asp Ala Thr Val Val Asn Ser Ile Ala Pro Val 150

Tyr Phe Lys Arg Asn Gln Leu Pro Lys. Ala Ile Pro Leu Asn Val Asp 170 165

Gly Tyr Leu Ile Ala Ala Asp Thr Gly Ile Lys Gly His Thr Lys Glu 185

Ala Val Gly Asp Val Ala Lys Leu Val Glu Thr Ala Lys Val Gln Thr

Met Asp Ile Val His His Leu Gly Gln Leu Thr His Gln Ala Lys Lys

Ala Ile Met Thr Asn Asn Leu Pro Gly Leu Gly Glu Ile Leu Asn Gln 235 230

Ser His Gln Leu Leu Lys Asp Leu Thr Val Ser Asn Pro Lys Leu Asp 250

Gln Leu Val Gln Ala Ala Gln Asp Ala Gly Ala Cys Gly Ala Lys Leu 260 265

Thr Gly Gly Gly Arg Gly Gly Cys Met Ile Ala Leu Ala Gln Ser Asn

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<213> Alloiococcus otitidis

<223	> CD > (4 > > 47	6)	(975														•
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cgt Arg 5	att Ile	ttt Phe	tta Leu	gtc Val	tat Tyr 10	gac Asp	cgt Arg	aaa Lys	gac Asp	tgg Trp 15	cag Gln	tct Ser	ctt Leu	aga Arg	gaa Glu 20	105	;
aat Asn	gcc Ala	agc Ser	ctt ·Leu	tct Ser 25	tta Leu	acg Thr	gaa Glu	aaa Lys	aac Asn 30	cta Leu	aat Asn	aac Asn	ttg Leu	cgt Arg 35	gca Ala	153	3
gtg Val	aat Asn	gac Asp	gtc Val 40	ata Ile	tcg Ser	atg Met	gaa Glu	gat Asp 45	gtc Val	cga Arg	gaa Glu	gtt Val	tac Tyr 50	gtc Val	ccc Pro	201	L
att Ile	atc Ile	caa Gln 55	tta Leu	ctg Leu	gat Asp	gtc Val	tac Tyr 60	ata Ile	aaa Lys	agt Ser	tac Tyr	tac Tyr 65	cgc Arg	cac His	cag Gln	249	)
gct Ala	tcc Ser 70	ttg Leu	atc Ile	aat Asn	tac Tyr	ttg Leu 75	aac Asn	ctg Leu	gac Asp	cag Gln	cct Pro 80	aaa Lys	aag Lys	tac Tyr	caa Gln	29	7
ccc Pro 85	tat Tyr	gtg Val	att Ile	Gly ggg	att Ile 90	gca Ala	GJA aaa	agc Ser	gtg Val	gct Ala 95	gtg Val	ggc Gly	aag Lys	tct Ser	acg Thr 100	34	5
gtt Val	gcc Ala	agg Arg	ctt Leu	čtt Leu 105	aag Lys	tcc Ser	ctc Leu	ttg Leu	agc Ser 110	gac Asp	tac Tyr	tat Tyr	ccg Pro	gaa Glu 115	aaa Lys	39	3
aag Lys	gta Val	gac Asp	ctc Leu 120	ctc Leu	aca Thr	aca Thr	gat Asp	ggc Gly 125	ttc Phe	ctt Leu	tat Tyr	ccg Pro	aat Asn 130	aag Lys	att Ile	44	1
tta Leu	aaa Lys	gag Glu 135	cga Arg	gat Asp	atc Ile	atg Met	gac Asp 140	cgc Arg	aag Lys	ggt Gly	ttt Phe	ccc Pro 145	gaa Glu	agc Ser	tat Tyr	48	9
gat Asp	atg Met 150	Lys	cgt Arg	ttg Leu	att Ile	aac Asn 155	ttt Phe	atg Met	acc Thr	gat Asp	gtc Val 160	aaa Lys	aat Asn	aat Asn	gtt Val	53	7
ccc Pro 165	Asn	atc Ile	cag Gln	gtg Val	ccc Pro 170	aag Lys	tat Tyr	tcc Ser	cac His	caa Gln 175	gtt Val	tac Tyr	gac Asp	ata Ile	gta Val 180	58	5
gaa Glu	GJA aaa	gaa Glu	agg Arg	ttg Leu 185	Thr	att Ile	aac Asn	cag Gln	cca Pro 190	gac Asp	atc Ile	ttg Leu	att Ile	gtc Val 195	Glu	63	3

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ggg Gly	atc Ile	aat Asn	gtg Val 200	ctc Leu	caa Gln	ctt Leu	cct Pro	tct Ser 205	aat Asn	gag Glu	aag Lys	att Ile	ttt Phe 210	gtt Val	agc Ser	681	
gat Asp	ttt Phe	ttc Phe 215	gac Asp	ttc Phe	tcc Ser	ttt Phe	tat Tyr 220	gtg Val	gat Asp	gcc Ala	tca Ser	gaa Glu 225	aat Asn	ctg Leu	att Ile	729	
gaa Glu	aaa Lys 230	tgg Trp	tac Tyr	atg Met	caa Gln	cgc Arg 235	ttt Phe	ggc	acc Thr	ttt Phe	atg Met 240	gat Asp	acc Thr	gcc Ala	ttc Phe	777	
caa Gln 245	gac Asp	ccc Pro	aac Asn	aac Asn	tat Tyr 250	tac Tyr	tac Tyr	aag Lys	ttt Phe	aat Asn 255	gac Asp	tgg Trp	gac Asp	cgc Arg	aag Lys 260	825	
gaa Glu	gct Ala	ttt Phe	gcc Ala	tat Tyr 265	gcc Ala	aac Asn	caa Gln	gtt Val	tgg Trp 270	gaa Glu	acg Thr	gtt Val	aac Asn	cta Leu 275	gaa Glu	873	
aac Asn	ctc Leu	agg Arg	gaa Glu 280	tat Tyr	att Ile	cta Leu	ccc Pro	acc Thr 285	cga Arg	ctc Leu	cgg Arg	gct Ala	aac Asn 290	ctc Leu	atc Ile	921	-
ctc Leu	cat His	aaa Lys 295	Thr	cat His	aac Asn	cac His	tac Tyr 300	atc Ile	gac Asp	aag Lys	att Ile	tta Leu 305	ctc Leu	aaa Lys	aaa Lys	969	ı
cac His	tga															975	;
<21 <21	0> 4 1> 3 2> P 3> A	09 RT	ococ	cus	otit	idis											
	0> 4 Asn		Ser	Arg 5	Ile	Phe	Leu	Val	Тут 10	Asp	Arg	Lys	Asp	Trp 15	Gln		
Ser	Leu	ı`Arg	Glu 20	Asn	Ala	. Ser	Leu	Ser 25	Leu	Thr	Glu	Lys	Asn 30	Leu	Asn		
Asn	Leu	ı Arg	, Ala	Val	Așn	Asp	Val	Ile	e Ser	Met	Glu	Asp 45	Val	Arg	Glu		

Val Tyr Val Pro Ile Ile Gln Leu Leu Asp Val Tyr Ile Lys Ser Tyr 50 55 60

Tyr Arg His Gln Ala Ser Leu Ile Asn Tyr Leu Asn Leu Asp Gln Pro 65 70 75 80

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- Lys Lys Tyr Gln Pro Tyr Val Ile Gly Ile Ala Gly Ser Val Ala Val
- Gly Lys Ser Thr Val Ala Arg Leu Leu Lys Ser Leu Leu Ser Asp Tyr 100
- Tyr Pro Glu Lys Lys Val Asp Leu Leu Thr Thr Asp Gly Phe Leu Tyr 125 120
- Pro Asn Lys Ile Leu Lys Glu Arg Asp Ile Met Asp Arg Lys Gly Phe
- Pro Glu Ser Tyr Asp Met Lys Arg Leu Ile Asn Phe Met Thr Asp Val 145
- Lys Asn Asn Val Pro Asn Ile Gln Val Pro Lys Tyr Ser His Gln Val 170 165
- Tyr Asp Ile Val Glu Gly Glu Arg Leu Thr Ile Asn Gln Pro Asp Ile 185
- Leu Ile Val Glu Gly Ile Asn Val Leu Gln Leu Pro Ser Asn Glu Lys 200
- Ile Phe Val Ser Asp Phe Phe Asp Phe Ser Phe Tyr Val Asp Ala Ser 215 210
- Glu Asn Leu Ile Glu Lys Trp Tyr Met Gln Arg Phe Gly Thr Phe Met 230 235 225
- Asp Thr Ala Phe Gln Asp Pro Asn Asn Tyr Tyr Tyr Lys Phe Asn Asp
- Trp Asp Arg Lys Glu Ala Phe Ala Tyr Ala Asn Gln Val Trp Glu Thr
- Val Asn Leu Glu Asn Leu Arg Glu Tyr Ile Leu Pro Thr Arg Leu Arg 280
- Ala Asn Leu Ile Leu His Lys Thr His Asn His Tyr Ile Asp Lys Ile 295 300

Leu Leu Lys Lys His 305	
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gtc caa gcc caa att aac ccc aat gag gaa att cgc cgg acc att gac Val Gln Ala Gln Ile Asn Pro Asn Glu Glu Ile Arg Arg Thr Ile Asp 15 20 25 30	96
ttt atc aag gac tat ctc cag gcc cac ccc ttc ttt gaa tcc tta atc Phe Ile Lys Asp Tyr Leu Gln Ala His Pro Phe Phe Glu Ser Leu Ile 35 40 45	144
ttg ggc atc tcc ggt ggc cag gat tcc acc ctc ctg ggt aag cta gcc Leu Gly Ile Ser Gly Gly Gln Asp Ser Thr Leu Leu Gly Lys Leu Ala 50 55 60	192
cag atg gcc tgc ctt gaa ctg agg gaa gag gag ggg tct gac aag cca Gln Met Ala Cys Leu Glu Leu Arg Glu Glu Glu Gly Ser Asp Lys Pro 65 70 75	240
att ttt att ggt atc cgc cta cct tat ggg gat caa ttt gat gaa gca Ile Phe Ile Gly Ile Arg Leu Pro Tyr Gly Asp Gln Phe Asp Glu Ala 80 85 90	288
gaa gcc cag caa gcc ctc aat tgg atc cag cct gac cag gct ctg acc Glu Ala Gln Gln Ala Leu Asn Trp Ile Gln Pro Asp Gln Ala Leu Thr 95 100 105 110	336
att aat atc aaa gag tcc gtt gat ggc ctg gtt gac act ttg gcc ggc Ile Asn Ile Lys Glu Ser Val Asp Gly Leu Val Asp Thr Leu Ala Gly 115 120 125	384
caa ggc att gaa gtt tct gac ttt aac aag ggc aat atc aaa gct cgg Gln Gly Ile Glu Val Ser Asp Phe Asn Lys Gly Asn Ile Lys Ala Arg 130 135 140	432
atc cga atg gtg gcc caa tat ggc gta gcg ggt cac ttc cac ggg gcg Ile Arg Met Val Ala Gln Tyr Gly Val Ala Gly His Phe His Gly Ala 145 150 155	480
gtg tta gga tct gac cat tca gcc gaa aat gta act ggc ttt ttc acc	528

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Val	Leu 160	Gly	Ser	Asp	His	Ser 165	Ala	Glu	Asn	Val	Thr 170	Gly	Phe	Phe	Thr		
aag Lys 175	cat His	Gly aaa	gac Asp	ggc Gly	gct Ala 180	agt Ser	gac Asp	ctc Leu	aac Asn	cct Pro 185	ctt Leu	ttc Phe	cgc Arg	cta Leu	aat Asn 190	576	
									gaa Glu 200							624	
ttg Leu	tac Tyr	caa Gln	aag Lys 210	acc Thr	ccc Pro	aca Thr	gct Ala	gat Asp 215	ttg Leu	gaa Glu	gaa Glu	gac Asp	cag Gln 220	ccc Pro	ggc	672	
									tct Ser							720	
									gag Glu							768	
aaa Lys 255	tgg Trp	tat Tyr	caa Gln	caa Gln	acg Thr 260	gcc Ala	cac His	aag Lys	cgc Arg	cac His 265	ttg Leu	ccg Pro	gtg Val	act Thr	atc Ile 270	816	
	_	_				-	aaa Lys									846	
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	0> 5( Gly		Asp	Leu 5	Arg	Glu	Glu	Ile	Leu 10	Asp	Arg	Met	Lys	Val 15	Gln		•
Ala	Gln	Ile	Asn 20	Pro	Asn	Glu	Glu	Ile 25	Arg	Arg	Thr	Ile	Asp 30	Phe	Ile		
Lys	Asp	Tyr 35	Leu	Gln	Ala	His	Pro 40	Phe	Phe	Glu	Ser	Leu 45	Ile	Leu	Gly		
Ile	Ser 50	Gly	Gly	Gln	Asp	Ser 55	Thr	Leu	Leu	Gly	Lys 60	Leu	Ala	Gln	Met		
Ala 65	Суѕ	Leu	Glu	Leu	Arg 70	Glu	Glu	Glu	Gly	Ser 75	Asp	Lys	Pro	Ile	Phe 80		

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Ile Gly Ile Arg Leu Pro Tyr Gly Asp Gln Phe Asp Glu Ala Glu Ala 90 85

Gln Gln Ala Leu Asn Trp Ile Gln Pro Asp Gln Ala Leu Thr Ile Asn 105 100

Ile Lys Glu Ser Val Asp Gly Leu Val Asp Thr Leu Ala Gly Gln Gly 120

Ile Glu Val Ser Asp Phe Asn Lys Gly Asn Ile Lys Ala Arg Ile Arg

Met Val Ala Gln Tyr Gly Val Ala Gly His Phe His Gly Ala Val Leu

Gly Ser Asp His Ser Ala Glu Asn Val Thr Gly Phe Phe Thr Lys His 165 . 170

Gly Asp Gly Ala Ser Asp Leu Asn Pro Leu Phe Arg Leu Asn Lys Arg 185

Gln Gly Arg Ala Leu Leu Glu Glu Leu Gly Ser Pro Lys Asn Leu Tyr

Gln Lys Thr Pro Thr Ala Asp Leu Glu Glu Asp Gln Pro Gly Leu Ser 210

Asp Glu Asp Lys Leu Gly Val Ser Tyr Glu Ala Ile Asp Asp Tyr Leu 235 230

Glu Gly Lys Pro Val Ser Gln Glu Asp Gln Ala Thr Ile Glu Lys Trp 255 250

Tyr Gln Gln Thr Ala His Lys Arg His Leu Pro Val Thr Ile Phe Asp 265

Asp Phe Trp Lys Glu Lys Asn 275

<210> 51

<211> 843

<212> DNA

<213> Alloiococcus otitidis

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									gcc Ala							96
_					_		-		gcc Ala 40							144
									gcc Ala							192
									ttg Leu							240
									aaa Lys							288
									cga Arg							336
									acc Thr 120							384
_					-				aaa Lys	_	_	_		-		432
									ccc Pro							480
							Ser		gtg Val							528
cta Leu 175	gaa Glu	gac Asp	caa Gln	gat Asp	ttg Leu 180	gat Asp	acc Thr	ctt Leu	att Ile	ttg Leu 185	ggt Gly	tgc Cys	acc Thr	cac His	tat Tyr 190	576
									tct Ser							624

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	195			200					205		
ttg gtt gat cca Leu Val Asp Pro	Gly Ala	gaa gct Glu Ala	gtg Val 215	aat Asn	gac Asp	ttg Leu	agt Ser	gtc Val 220	tta Leu	tta Leu	672
gac tat tat gad Asp Tyr Tyr Asp 225	ttg act Leu Thr	aat gad Asn Asp 230	Arg	ttt Phe	aat Asn	ccc Pro	aac Asn 235	ctg Leu	acc Thr	cac His	720
cat ttt tac acc His Phe Tyr The 240	e acg gga Thr Gly	gat aaa Asp Lys 245	gcc Ala	GJA aaa	ttt Phe	aag Lys 250	aaa Lys	atc Ile	gcg Ala	gat Asp	768
gac tgg ctt gad Asp Trp Leu Asp 255	c cac cac His His 260	Asn Ty	cgg Arg	gtt Val	gac Asp 265	cat His	tta Leu	gat Asp	tta Leu	gag Glu 270	816
gag ttg caa ga Glu Leu Gln Gl											843
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<213> ATTOTOCO	ccus otit	idis									
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<pre>&lt;400&gt; 52 Met Ile Met Ty 1  Gly Phe Thr Le 20  Phe Tyr Tyr Le 35  Met Ala Thr Va</pre>	r Thr Asg 5 u Val Lys u Gly Asg	Gly Il G Glu Al Thr Al 40 A Tyr Al 55	a Met 25 a Arg	Lys Ser Glu	Gln Pro Leu	Leu Tyr Ala	Pro Gly 45 Asn	Asn 30 Pro	Glu Lys	Gln Asp Val	
<pre>&lt;400&gt; 52 Met Ile Met Ty 1  Gly Phe Thr Le 20  Phe Tyr Tyr Le 35  Met Ala Thr Va 50  Lys Asn His Gl</pre>	r Thr Asp 5 u Val Lys u Gly Asp 1 Lys Ala n Ile Lys 70	Gly II Glu Al Thr Al 40 A Tyr Al 55	a Met 25 a Arg a Phe	Lys Ser Glu	Gln Pro Leu Ala 75	Leu Tyr Ala 60 Cys	Pro Gly 45 Asn	Asn 30 Pro Tyr	Glu Lys Leu	Gln Asp Val Thr 80	

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His Gln Ile Gly Val Ile Ala Thr His Gly Thr Ile Gln Ser Gly Arg 115 120 125

Tyr Glu Leu Glu Leu Lys Arg Lys Arg Pro Asp Ile Glu Val Thr Ser 130 135 140

Leu Ala Cys Pro Glu Phe Ala Pro Met Val Glu Ala Gly Asp Tyr Arg 145 150 155 160

Ser Val Gln Ala Ser Ser Val Val Arg Thr Ser Leu Gln Ala Leu Glu 165 170 175

Asp Gln Asp Leu Asp Thr Leu Ile Leu Gly Cys Thr His Tyr Pro Ile 180 185 190

Ile Lys Asp Leu Ile Gln Asp Ser Ile Gly Pro Gly Ile Ser Leu Val 195 200 205

Asp Pro Gly Ala Glu Ala Val Asn Asp Leu Ser Val Leu Leu Asp Tyr 210 215 220

Tyr Asp Leu Thr Asn Asp Arg Phe Asn Pro Asn Leu Thr His His Phe 225 230 235 240

Tyr Thr Thr Gly Asp Lys Ala Gly Phe Lys Lys Ile Ala Asp Asp Trp 245 250 255

Leu Asp His His Asn Tyr Arg Val Asp His Leu Asp Leu Glu Glu Leu 260 265 270

Gln Glu Val Asn Gly Arg 275

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<211> 957

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<213> Alloiococcus otitidis

<220>

<221> CDS

<222> (7)..(957)

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						-	-				gat Asp		_			96
											gac Asp					144
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											att Ile					240
											tca Ser 90					288
											Gly ggg					336
	_							_	_	_	atg Met	-	_	-		384
											aac Asn					432
											gat Asp					480
_				-	-	_	-	_		-	tcc Ser 170					528
-		_	_	_				-		_	ttg Leu					576
											gac Asp					624
		_		_	_	_			_	_	att Ile		_	_	_	672
											gga Gly					720

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	ttc Phe 240															768
gag Glu 255	acc Thr	caa Gln	gca Ala	gga Gly	ttt Phe 260	tta Leu	aac Asn	cga Arg	gac Asp	ctg Leu 265	gct Ala	tca Ser	gct Ala	att Ile	att Ile 270	816
	gct Ala		_	_				_	_		-					864
	gat Asp															912
	gat Asp													taa		957
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Leu	Asn	Ile	Thr 20	Pro	Asp	Ser	Phe	Ser 25	Asp	Gly	Gly	Arg	Asn 30	Tyr	Gln	
Ala	Asp	Gln 35	Ala	Ile	Ala	His	Gly 40	Leu	Asp	Leu	Val	Asp 45	Lys	Gly	Ala	
Asp	Met 50	Leu	Asp	Ile	Gly	Gly 55	Glu	Ser	Thr	Arg	Pro 60	Gly	Ser	Ser	Pro	
Val 65	Asp	Leu	Gln	Asp	Glu 70	Ile	Asp	Arg	Ile	Val 75	Pro	Val	Ile	Lys	Gly 80	
65	Asp Arg			_	70					75			•		80	

Ile Thr Gly Leu Thr Gly Asp Val Asp Met Ala Asp Leu Leu Ala Gln

112/235

115 120 125

Glu Gly Val Lys Ala Ile Val Met Phe Asn Pro Val Ile Ala Arg Pro 130 135 140

Asp His Pro Ser Ser Gln Lys Phe Arg Asp Phe Gly Gly Arg Asp Phe 145 150 155

Phe Thr Asp Glu Glu Arg Asp Lys Met Ser Gln Ala Pro Ile Glu Glu 165 170 175

Ala Met Met Val Tyr Phe Asp Lys Val Leu Asn Lys Ala His Gln Ala 180 185 190

Gly Ile Asp Arg Asp Lys Ile Leu Leu Asp Pro Gly Ile Gly Phe Gly 195 200 205

Leu Thr Lys Lys Glu Asn Tyr Lys Leu Ile His Ser Val Ala Ser Ile 210 215 220

His Asp Lys Gly Tyr Pro Val Phe Leu Gly Val Ser Arg Lys Arg Phe 225 230 235 240

Leu Val Gly Glu Val Ser Lys Leu Gly Ile Glu Ala Asp Pro Glu Thr 245 250 255

Gln Ala Gly Phe Leu Asn Arg Asp Leu Ala Ser Ala Ile Ile Thr Ala 260 265 270

Tyr Ala Ser His Ile Gly Val Asp Tyr Val Arg Val His Ser Leu Asp 275 280 285

Glu His Lys Ile Ala Thr Thr Ile Thr His Asn Ile Leu Asn Ser Asp 290 295 300

Ser Leu Asp Asp Gln Ser Phe Asp Gln Tyr Lys Asn 305 310 315

<210> 55

<211> 561

<212> DNA

<213> Alloiococcus otitidis

<220>

113/235

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atc Ile	att Ile	ggt Gly	aaa Lys	gac Asp 30	aag Lys	gtt Val	ttg Leu	cct Pro	tgg Trp 35	gaa Glu	ttg Leu	tcc Ser	aat Asn	gac Asp 40	tta Leu	150
aag Lys	cat His	ttt Phe	aaa Lys 45	aaa Lys	gtt Val	aca Thr	gaa Glu	ggt Gly 50	cac His	acc Thr	atc Ile	ctg Leu	atg Met 55	ggc	cgg Arg	198
aag Lys	acc Thr	ttt Phe 60	gaa Glu	gga Gly	atg Met	gat Asp	aaa Lys 65	aag Lys	ccc Pro	ctc Leu	cct Pro	aac Asn 70	cga Arg	aaa Lys	acc Thr	246
ttg Leu	gta Val 75	ttg Leu	acc Thr	cgc Arg	caa Gln	gat Asp 80	gac Asp	tac Tyr	caa Gln	gct Ala	ggg Gly 85	gac Asp	gac Asp	cag Gln	gtt Val	294
gaa Glu 90	gtc Val	gtc Val	cac His	tcc Ser	aaa Lys 95	gac Asp	cag Gln	gcc Ala	ttg Leu	act Thr 100	tat Tyr	gcg Ala	tca Ser	ggt Gly	cat His 105	342
Gly ggg	gtg Val	gac Asp	ctc Leu	tat Tyr 110	gtg Val	att Ile	ggt Gly	GJÀ aaa	gcc Ala 115	ggc Gly	att Ile	ttc Phe	gac Asp	ttg Leu 120	ttt Phe	3,90
ctg Leu	gac Asp	caa Gln	gtt Val 125	gat Asp	gtt Val	ctc Leu	cac His	caa Gln 130	aca Thr	gtt Val	atc Ile	cac His	gag Glu 135	agc Ser	ttt Phe	438
gat Asp	ggt Gly	gac Asp 140	Thr	acc Thr	atg Met	cca Pro	gac Asp 145	Ile	gac Asp	tgg Trp	gac Asp	agc Ser 150	ttt Phe	aat Asn	cag Gln	486
gtg Val	tct Ser 155	Lys	gct Ala	tat Tyr	tat Tyr	gac Asp 160	Gln	gct Ala	gac Asp	ggt Gly	cac His 165	Asn	cac His	tcc Ser	cac His	534
			-		aga Arg 175	Arg										561

<210> 56

<211> 177 <212> PRT

114/235

<213> Alloiococcus otitidis

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Leu Pro Trp Glu Leu Ser Asn Asp Leu Lys His Phe Lys Lys Val Thr 35 40

Glu Gly His Thr Ile Leu Met Gly Arg Lys Thr Phe Glu Gly Met Asp 50 55 60

Lys Lys Pro Leu Pro Asn Arg Lys Thr Leu Val Leu Thr Arg Gln Asp 65 70 75 80

Asp Tyr Gln Ala Gly Asp Asp Gln Val Glu Val Val His Ser Lys Asp 85 90 95

Gln Ala Leu Thr Tyr Ala Ser Gly His Gly Val Asp Leu Tyr Val Ile 100 105 110

Gly Gly Ala Gly Ile Phe Asp Leu Phe Leu Asp Gln Val Asp Val Leu 115 120 125

His Gln Thr Val Ile His Glu Ser Phe Asp Gly Asp Thr Thr Met Pro 130 135 140

Asp Ile Asp Trp Asp Ser Phe Asn Gln Val Ser Lys Ala Tyr Tyr Asp 145 150 155 160

Gln Ala Asp Gly His Asn His Ser His Thr Ile Tyr Glu Tyr Arg Arg 165 170 175

Lys

<210> 57

<211> 1968

<212> DNA

<213> Alloiococcus otitidis

<220>

<221> CDS

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acc Thr	gat Asp	gcc Ala	agg Arg	ggt Gly 35	ttg Leu	cac His	cac His	ctg Leu	gtt Val 40	tat Tyr	gaa Glu	att Ile	acc Thr	gat Asp 45	aat Asn	144
gct Ala	att Ile	gat Asp	gag Glu 50	gtt Val	ttg Leu	gct Ala	ggc Gly	tac Tyr 55	gct Ala	gat Asp	gaa Glu	att Ile	gaa Glu 60	gtc Val	aag Lys	192
atc Ile	cac His	acg Thr 65	gac Asp	ggc Gly	tcg Ser	gtt Val	tcg Ser 70	gtc Val	aaa Lys	gac Asp	aat Asn	gga Gly 75	cgg Arg	ggc Gly	atg Met	240
cca Pro	acc Thr 80	GJA aaa	atg Met	cat His	gag Glu	tca Ser 85	ggc Gly	cta Leu	ccc Pro	acc Thr	atc Ile 90	cag Gln	gtt Val	atc Ile	ttt Phe	288
acc Thr 95	gtc Val	ctc Leu	cat His	gcc Ala	ggg Gly 100	gga Gly	aaa Lys	ttt Phe	ggc	caa Gln 105	gag Glu	Gly ggg	gcc Ala	tac Tyr	aag Lys 110	336
tca Ser	gcc Ala	ggt Gly	gga Gly	ctc Leu 115	cat His	GJÀ âââ	gtt Val	Gly	gcc Ala 120	tcg Ser	gtc Val	gtc Val	aac Asn	gcc Ala 125	ttg Leu	384
tct Ser	gat Asp	tgg Trp	ctc Leu 130	acg Thr	gtg Val	ata Ile	gtg Val	acc Thr 135	aag Lys	gac Asp	ggc	tat Tyr	gaa Glu 140	tac Tyr	cgg Arg	432
caa Gln	gac Asp	ttt Phe 145	Ser	caa Gln	gga Gly	ggc	cag Gln 150	gct Ala	aaa Lys	gga Gly	Gly ggc	atc Ile 155	cag Gln	aag Lys	aga Arg	480
aaa Lys	att Ile 160	Asn	cag Gln	caa Gln	aaa Lys	tcc Ser 165	Ser	acc Thr	ctg Leu	gtc Val	cac His 170	Phe	aaa Lys	ccc Pro	tca Ser	528
ggc Gly 175	Gln	gtc Val	ttt Phe	tcg Ser	acc Thr	Thr	gaa Glu	ttt Phe	aac Asr	ttt Phe 185	: Asn	acc Thr	ato	tgt Cys	gag Glu 190	576
cgg Arg	atg Met	cgg : Arg	gag g Glu	tcg Ser 195	Ala	ttc Phe	ctt Leu	gto Val	aaa Lys 200	s Gly	acc Thr	aag Lys	att Ile	acc Thr 205	gta Val	624
gag	gac	cto	g cgc	: caç	g gaa	a gaa	ago	caç	gto	ttc	caa	ttt	aat	gaa	a gga	672

Glu	Asp	Leu	Arg 210	Gln	Glu	Glu	Ser	Gln 215	Val	Phe	Gln	Phe	Asn 220	Glu	Gly	
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cca Pro	gta Val 240	acc Thr	tat Tyr	ttt Phe	gaa Glu	ggt Gly 245	tct Ser	gaa Glu	gat Asp	gaa Glu	att Ile 250	gaa Glu	gtt Val	gaa Glu	ttt Phe	768
gcc Ala 255	ttc Phe	caa Gln	tac Tyr	aat Asn	gac Asp 260	ggc Gly	tat Tyr	tcg Ser	gag Glu	acg Thr 265	gtt Val	ctg Leu	agt Ser	ttt Phe	gtc Val 270	816
aac Asn	aat Asn	gtc Val	cgt Arg	acc Thr 275	cgg Arg	gat Asp	Gly ggg	ggc Gly	agc Ser 280	cac His	gaa Glu	act Thr	gga Gly	gct Ala 285	aag Lys	864
tca Ser	gct Ala	att Ile	acc Thr 290	aag Lys	gct Ala	ttc Phe	aac Asn	gac Asp 295	tat Tyr	gct Ala	agg Arg	aaa Lys	agt Ser 300	ggc Gly	tta Leu	912
ctc Leu	aaa Lys	gag Glu 305	Lys	gac Asp	agt Ser	aac Asn	ttg Leu 310	gaa Glu	gga Gly	tct Ser	gac Asp	gtc Val 315	cgg Arg	gaa Glu	GJA aaa	960
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gaa Glu 335	. Gly	cag	acc Thr	aag Lys	agc Ser 340	Lys	tta Leu	gga Gly	act Thr	Pro	GIN	gco Ala	cgg Arg	acc	gcc Ala 350	1056
act Thr	gac Asp	cag Glr	g gtt n Val	ato Ile	Ser	gaa Glu	tcc Ser	tta Lev	act Thr 360	Тух	tto Phe	ctg Lev	g gcc 1 Ala	gaa Glu 365	aat Asn	1104
Gly ggg	gac Asp	tto Lei	tct Ser 370	Lys	caa Gln	ctt Lev	att Ile	cgc Arg	l PAs	g gcc s Ala	ato a Ile	c cga	a gcc g Ala 380	LAIC	tct Ser	1152
gco Ala	agg Arg	g gaa g Glv 385	ı Ala	a gct a Ala	cgc Arg	aag Lys	g gco s Ala 390	а Гу:	g gad s Asp	c caç o Gli	ı Ser	c cgg c Arg	g Asi	tct Sei	gct Ala	1200
tc: Se:	c aag Lys 400	s Ly	a aaa s Ly:	a gtt s Val	gaa l Glu	a act 1 Thi 405	: Lei	c cto	g tci u Sei	t gg: r Gl:	t aag y Ly: 41	s Le	g aco u Thi	c cca	a gct o Ala	1248
caa Gl: 41!	n Se	c aa r Ly	g aa s Asi	c gco n Ala	c cag a Glr 420	ı Ly:	a aat s Asi	t ga n Gl	a ct <sup>.</sup>	t ta u Ty: 42	r Le	a gt u Va	g gaq 1 Gl	g gg u Gl	g gat y Asp 430	*
tc: Se:	g gc	t gg a Gl	t gg y Gl	g tc y Se	a gco r Ala	c aag a Ly	g ca s Gl:	a gg n Gl	t ag y Ar	g ga g As	c cg p Ar	g aa g Ly	a tt	c ca e Gl	a gca n Ala	1344

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	117/235	

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				435									L_L		<b>.</b>	1392
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gat Asp	gat Asp	att Ile 465	tta Leu	aaa Lys	aat Asn	gaa Glu	gaa Glu 470	att Ile	tct Ser	acc Thr	atg Met	att Ile 475	tat Tyr	acc Thr	atc Ile	1440
ggt Gly	gca Ala 480	ggt Gly	gct Ala	GJA āāā	cct Pro	gag Glu 485	ttt Phe	gat Asp	att Ile	gaa Glu	gct Ala 490	gtt Val	aat Asn	tac Tyr	gat Asp	1488
aag Lys 495	ata Ile	gtc Val	att Ile	atg Met	act Thr 500	gat Asp	gcc Ala	gac Asp	aca Thr	gac Asp 505	ggc Gly	gcc Ala	cac His	atc Ile	cag Gln 510	1536
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gca Ala	Gly	aag Lys	gtc Val 530	tat Tyr	att Ile	gcc Ala	cta Leu	ccg Pro 535	ccc Pro	ttg Leu	tat Tyr	aag Lys	ttg Leu 540	acc Thr	aaa Lys	1632
aag Lys	caa Gln	gga Gly 545	aag Lys	caa Gln	gaa Glu	aaa Lys	aca Thr 550	gcc Ala	tat Tyr	gct Ala	tgg Trp	act Thr 555	gat Asp	gag Glu	gag Glu	1680
ttg Leu	gaa Glu 560	gac Asp	ctg Leu	gtt Val	aaa Lys	gat Asp 565	ttt Phe	Gly	aaa Lys	cac His	tac Tyr 570	Thr	ctc Leu	cag Gln	cgc Arg	1728
tac Tyr 575	aag Lys	ggt	tta Leu	ggc	gag Glu 580	atg Met	aat Asn	gct Ala	gac Asp	cag Gln 585	Leu	tgg Trp	gag Glu	acc Thr	acc Thr 590	1776
atg Met	gac Asp	cca Pro	gag Glu	acc Thr 595	Arg	acc Thr	ttg Leu	ato	cgg Arg 600	Val	acc	att Ile	gaa Glu	gac Asp 605	agt Ser	1824
gaa Glu	aag Lys	gct Ala	gaa Glu 610	Arg	cgg Arg	gtt Val	tcc Ser	acc Thr 615	Leu	atg Met	ggg Gly	acc Thr	aag Lys 620	Val	gat Asp	1872
cct Pro	aga Arg	cgg Arg 625	l Pàs	tgg Trp	att Ile	gaa Glu	gac Asp 630	His	att : Ile	gaa Glu	tto Phe	agt Ser 635	Leu	gca Ala	gaa Glu	1920
gat As <b>r</b>	ggc Gly 640	sez	att	tta Lev	gag Glu	aac Asr 645	Lys	gto Val	cta Lev	n gaa n Glu	gga Gly 650	r Glu	gcc Ala	aaç Lys	taa ;	1968

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<212> PRT <213> Alloiococcus otitidis

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Leu Asp Ala Val Lys Lys Arg Pro Gly Met Tyr Ile Gly Ser Thr Asp 20 25 30

Ala Arg Gly Leu His His Leu Val Tyr Glu Ile Thr Asp Asn Ala Ile 35 40 45

Asp Glu Val Leu Ala Gly Tyr Ala Asp Glu Ile Glu Val Lys Ile His 50 55

Thr Asp Gly Ser Val Ser Val Lys Asp Asn Gly Arg Gly Met Pro Thr 65 70 75 80

Gly Met His Glu Ser Gly Leu Pro Thr Ile Gln Val Ile Phe Thr Val 85 90 95

Leu His Ala Gly Gly Lys Phe Gly Gln Glu Gly Ala Tyr Lys Ser Ala 100 105 110

Gly Gly Leu His Gly Val Gly Ala Ser Val Val Asn Ala Leu Ser Asp 115 120 125

Trp Leu Thr Val Ile Val Thr Lys Asp Gly Tyr Glu Tyr Arg Gln Asp 130 135 140

Phe Ser Gln Gly Gly Gln Ala Lys Gly Gly Ile Gln Lys Arg Lys Ile 145 150 155 160

Asn Gln Gln Lys Ser Ser Thr Leu Val His Phe Lys Pro Ser Gly Gln 165 170 175

Val Phe Ser Thr Thr Glu Phe Asn Phe Asn Thr Ile Cys Glu Arg Met 180 185 190

Arg Glu Ser Ala Phe Leu Val Lys Gly Thr Lys Ile Thr Val Glu Asp 195 200 205

Leu Arg Gln Glu Glu Ser Gln Val Phe Gln Phe Asn Glu Gly Ile Lys

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220 215 210 Ala Phe Val Asp Tyr Leu Asn Glu Gly Lys Asp Thr Leu Ser Pro Val 235 230 225 Thr Tyr Phe Glu Gly Ser Glu Asp Glu Ile Glu Val Glu Phe Ala Phe 250 Gln Tyr Asn Asp Gly Tyr Ser Glu Thr Val Leu Ser Phe Val Asn Asn 265 Val Arg Thr Arg Asp Gly Gly Ser His Glu Thr Gly Ala Lys Ser Ala Ile Thr Lys Ala Phe Asn Asp Tyr Ala Arg Lys Ser Gly Leu Leu Lys 300 295 Glu Lys Asp Ser Asn Leu Glu Gly Ser Asp Val Arg Glu Gly Ile Ala 315 310 305 Val Val Leu Ser Val Arg Ile Pro Glu Glu Ile Leu Gln Phe Glu Gly 330 Gln Thr Lys Ser Lys Leu Gly Thr Pro Gln Ala Arg Thr Ala Thr Asp Gln Val Ile Ser Glu Ser Leu Thr Tyr Phe Leu Ala Glu Asn Gly Asp 360 355 Leu Ser Lys Gln Leu Ile Arg Lys Ala Ile Arg Ala Arg Ser Ala Arg 375 Glu Ala, Ala Arg Lys Ala Lys Asp Gln Ser Arg Asn Ser Ala Ser Lys 395 400 390 Lys Lys Val Glu Thr Leu Leu Ser Gly Lys Leu Thr Pro Ala Gln Ser 405 Lys Asn Ala Gln Lys Asn Glu Leu Tyr Leu Val Glu Gly Asp Ser Ala 425 420

Gly Gly Ser Ala Lys Gln Gly Arg Asp Arg Lys Phe Gln Ala Ile Leu

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Pro Leu Arg Gly Lys Val Ile Asn Thr Glu Lys Ser Ser Leu Asp Asp 450 455 460

Ile Leu Lys Asn Glu Glu Ile Ser Thr Met Ile Tyr Thr Ile Gly Ala 465 470 475 480

Gly Ala Gly Pro Glu Phe Asp Ile Glu Ala Val Asn Tyr Asp Lys Ile 485 490 495

Val Ile Met Thr Asp Ala Asp Thr Asp Gly Ala His Ile Gln Val Leu 500 505 510

Leu Leu Thr Phe Phe Tyr Arg Tyr Met Lys Pro Leu Ile Glu Ala Gly 515 520 525

Lys Val Tyr Ile Ala Leu Pro Pro Leu Tyr Lys Leu Thr Lys Lys Gln 530 535

Gly Lys Gln Glu Lys Thr Ala Tyr Ala Trp Thr Asp Glu Glu Leu Glu 545 550 555 560

Asp Leu Val Lys Asp Phe Gly Lys His Tyr Thr Leu Gln Arg Tyr Lys 565 570 575

Gly Leu Gly Glu Met Asn Ala Asp Gln Leu Trp Glu Thr Thr Met Asp 580 585 590

Pro Glu Thr Arg Thr Leu Ile Arg Val Thr Ile Glu Asp Ser Glu Lys 595 600 605

Ala Glu Arg Arg Val Ser Thr Leu Met Gly Thr Lys Val Asp Pro Arg 610 615 620

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gaa Glu	gat Asp	gtc Val	atg Met	ggg Gly 20	gac Asp	cgg Arg	ttc Phe	ggc	cgg Arg 25	tat Tyr	tcc Ser	aag Lys	tac Tyr	att Ile 30	ata Ile		96
cag Gln	gaa Glu	agg Arg	gcc Ala 35	cta Leu	ccg Pro	gac Asp	ttg Leu	cgg Arg 40	gac Asp	ggt Gly	tta Leu	aaa Lys	ccg Pro 45	gtc Val	caa Gln	1	L44
aga Arg	cgg Arg	atc Ile 50	ctc Leu	tat Tyr	gcc Ala	atg Met	cac His 55	cag Gln	gac Asp	aaa Lys	aac Asn	acc Thr 60	tat Tyr	gac Asp	aag Lys	1	L92
gct Ala	tac Tyr 65	cgg Arg	aag Lys	tcg Ser	gcc Ala	aag Lys 70	acg Thr	gtg Val	gga Gly	aat Asn	gtc Val 75	ata Ile	GJA aaa	aac Asn	tac Tyr	;	240
cac His 80	ccc Pro	cat His	ggc Gly	gac Asp	aca Thr 85	tcc Ser	gtt Val	tac Tyr	gat Asp	gcc Ala 90	atg Met	gtt Val	agg Arg	ctc Leu	agt Ser 95	:	288
cag Gln	cct Pro	tgg Trp	aag Lys	atg Met 100	cgc Arg	cat His	cct Pro	ttg Leu	gtt Val 105	gat Asp	atg Met	cac His	Gly ggg	aac Asn 110	aag Lys	:	336
ggg	agc Ser	atg Met	gac Asp 115	ggg	gac Asp	cca Pro	cca Pro	gct Ala 120	gcc Ala	atg Met	cgg Arg	tac Tyr	acc Thr 125	gaa Glu	gcc Ala	;	384
cgt Arg	ctg Leu	tct Ser 130	aaa Lys	att Ile	gct Ala	tcc Ser	gac Asp 135	Leu	ctg Leu	gct Ala	gat Asp	att Ile 140	gat Asp	aag Lys	gag Glu		432
acg Thr	gtg Val 145	Asp	cat His	gtc Val	tta Leu	aac Asn 150	ttt Phe	gat Asp	gac Asp	acg Thr	acc Thr 155	Glu	gag Glu	ccc Pro	acc Thr		480
gto Val 160	. Leu	ccc Pro	gcc Ala	cgt Arg	ttt Phe 165	Pro	aac Asn	ctc Leu	ttg Leu	gtc Val 170	Asn	ggg Gly	gct Ala	agc Ser	ggg Gly 175		528
att Ile	tca Ser	gcc Ala	ggt Gly	tat Tyr 180	Ala	act Thr	gac Asp	ata Ile	ccg Pro 185	Pro	cat His	aat Asn	ttg Leu	ago Ser 190	gag Glu		576
gto	att	gat	gcc	acc	ato	cac	tta	atc	aac	cac	ccc	aat	gca	agg	ctg		624

Val	Ile	Asp	Ala 195	Thr	Ile	His	Leu	Ile 200	Asn	His	Pro	Asn	Ala 205	Arg	Leu	
gag Glu	act Thr	ttg Leu 210	atg Met	gac Asp	tat Tyr	att Ile	caa Gln 215	gga Gly	cca Pro	gac Asp	ttt Phe	ccg Pro 220	act Thr	GJA aaa	GJÀ aaa	672
att Ile	atc Ile 225	caa Gln	ggt Gly	aaa Lys	agt Ser	ggc Gly 230	ctg Leu	aag Lys	aaa Lys	gcc Ala	tac Tyr 235	caa Gln	acg Thr	ggc Gly	aag Lys	720
gga Gly 240	aaa Lys	att Ile	atc Ile	atc Ile	cgg Arg 245	gcc Ala	aaa Lys	gca Ala	gat Asp	att Ile 250	gag Glu	gcc Ala	atc Ile	cgg Arg	ggt Gly 255	768
ggc Gly	aaa Lys	tcc Ser	caa Gln	att Ile 260	gtc Val	atc Ile	agt Ser	caa Gln	att Ile 265	cct Pro	tat Tyr	gag Glu	gtc Val	aac Asn 270	aag Lys	816
gca Ala	agg Arg	ttg Leu	gtc Val 275	caa Gln	aaa Lys	att Ile	gac Asp	gac Asp 280	atc Ile	Arg Arg	att Ile	aac Asn	aaa Lys 285	aaa Lys	atc Ile	864
gaċ Asp	ggc Gly	att Ile 290	gcc Ala	gat Asp	gtc Val	cgg Arg	gat Asp 295	gaa Glu	agt Ser	gac Asp	cgg Arg	tct Ser 300	Gly	ttg Leu	cgg Arg	912
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tta Leu	tct Ser	tct Ser	tac Tyr 355	Leu	gac Asp	cac His	aag Lys	cgg Arg 360	Thr	gtg Val	gtt Val	caa Gln	aac Asn 365	Arg	acc Thr	1104
cgt Arg	tac	ctc Leu 370	Leu	Ala	Lys	Ala	Lys	Asp	) Arg	cag Gln	. His	att Ile 380	Val	caa Gln	ggc Gly	1152
ctt Leu	ato 11e 385	Lys	gcc Ala	att Ile	tca Ser	ato 11e 390	Leu	gat Asp	gac Asp	ttg Leu	ato Ile 395	Gln	acc Thr	ato Ile	cgg Arg	1200
gcc Ala 400	Ser	gaa Glu	aac Asr	aag Lys	gcc Ala 405	Asr	gco Ala	aag Lys	ggaa Glu	aat Asn 410	Ile	ato E Ile	cag Gln	gct Ala	tat Tyr 415	1248
ggt Gly	ttt Phe	ago Ser	caa Glr	gac Asp	caa Glr	gco Ala	gaa Glu	gco Ala	att a Ile	gto Val	tco Sei	c cto	cag Glr	ctt Leu	tac ı Tyr	1296

				420					425					430		
cgc Arg	ttg Leu	acc Thr	aat Asn 435	aca Thr	gat Asp	ata Ile	aag Lys	gac Asp 440	tta Leu	caa Gln	gca Ala	gaa Glu	gcc Ala 445	aaa Lys	gac Asp	1344
tta Leu	gcc Ala	caa Gln 450	gcc Ala	atc Ile	ctg Leu	acc Thr	tac Tyr 455	cag Gln	gac Asp	ctc Leu	tta Leu	acc Thr 460	aac Asn	aag Lys	gcc Ala	1392
agc Ser	ctg Leu 465	gat Asp	gct Ala	ttg Leu	atg Met	aaa Lys 470	gaa Glu	gaa Glu	ttg Leu	aaa Lys	gaa Glu 475	gtc Val	aaa Lys	caa Gln	gca Ala	1440
tat Tyr 480	GJA āāā	gag Glu	gac Asp	cgg Arg	cta Leu 485	acc Thr	cag Gln	gtc Val	caa Gln	gac Asp 490	aag Lys	atc Ile	gaa Glu	aaa Lys	cta Leu 495	1488
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gtc Val	acc Thr	cag Gln	gga Gly 515	ggt Gly	tac Tyr	ttg Leu	aag Lys	cgg Arg 520	acc Thr	tcc Ser	atc Ile	cgg Arg	tct Ser 525	tac Tyr	aag Lys	1584
gct Ala	tcc Ser	caa Gln 530	gtg Val	gag Glu	gaa Glu	ttg Leu	ggc Gly 535	cgg Arg	cga Arg	gaa Glu	gac Asp	gac Asp 540	ttg Leu	gtc Val	atc Ile	1632
ttt Phe	atg Met 545	caa Gln	gag Glu	ttg Leu	tca Ser	acc Thr 550	cta Leu	gac Asp	caa Gln	ctc Leu	ctt Leu 555	Ile	ttc Phe	acc Thr	tcg Ser	1680
aaa Lys 560	Gly	aat Asn	gtg Val	gtc Val	aac Asn 565	cga Arg	cca Pro	gtc Val	cat His	gaa Glu 570	Leu	ccg Pro	gac Asp	atc Ile	aag Lys 575	1728
tgg Trp	aag Lys	gat Asp	att Ile	gga Gly 580	gag Glu	cac	ttg Leu	tca Ser	agg Arg 585	Thr	ato Ile	ccc Pro	ctt Leu	gga Gly 590	GIu	1776
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aag Lys	cgc Arg	tat Tyr 610	· Val	ttt Phe	atc Ile	act Thr	cga Arg 615	Asp	ggc Gly	tat Tyr	ato Ile	aaa Lys 620	Gln	agt Ser	cca Pro	1872
gag Glu	acg Thr 625	Glu	ttt i Phe	gag Glu	g ccc 1 Pro	aaa Lys 630	Arg	act Thi	tac Tyr	aaç Lys	tct Ser 635	Arg	gct Ala	tca Ser	act Thr	1920
gcc Ala 640	ı Ile	aaa Lys	a tta 5 Lei	a aaa 1 Lys	tca Ser 645	Ası	caa Glr	gat Asp	aga Arg	Lev 650	ı Glr	g gca n Ala	gto Val	tac Tyr	tat Tyr 655	1968

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tac Tyr	GJÀ ããã	ctc Leu	aag Lys 675	tat Tyr	gga Gly	cta Leu	gaa Glu	gaa Glu 680	gtg Val	tca Ser	gaa Glu	gta Val	ggg Gly 685	gcc Ala	cag Gln	2064
gct Ala	gca Ala	ggc Gly 690	gtc Val	aag Lys	tcc Ser	atg Met	aac Asn 695	ctg Leu	aaa Lys	gag Glu	GJA āāā	gac Asp 700	cat His	gtc Val	caa Gln	2112
gat Asp	ggt Gly 705	ttg Leu	gtc Val	ttt Phe	aag Lys	cgt Arg 710	aag Lys	cag Gln	ttc Phe	caa Gln	gaa Glu 715	gcc Ala	ttg Leu	ttc Phe	att Ile	2160
acc Thr 720	cag Gln	cga Arg	gcc Ala	agt Ser	gtt Val 725	aag Lys	aaa Lys	atg Met	gcc Ala	ctc Leu 730	cat His	gac Asp	ttt Phe	gac Asp	cgg Arg 735	2208
act Thr	tca Ser	cgg Arg	gcc Ala	aag Lys 740	cgg Arg	ggt Gly	tta Leu	caa Gln	atc Ile 745	ctc Leu	aga Arg	gaa Glu	ctg Leu	aag Lys 750	cga Arg	2256
aac Asn	ccc Pro	cac His	cga Arg 755	atc Ile	cag Gln	ttt Phe	atg Met	atc Ile 760	Gly	att Ile	tca Ser	caa Gln	aat Asn 765	aaa Lys	ttc Phe	2304
ctg Leu	gtc Val	aat Asn 770	Leu	cta Leu	act Thr	gat Asp	aca Thr 775	Lys	aaa Lys	cta Leu	gta Val	cag Gln 780	ata Ile	aac Asn	cca Pro	2352
gat Asp	gac Asp 785	Tyr	aca Thr	gtt Val	tca Ser	aac Asn 790	Arg	cat His	aac Asn	aat Asn	ggg Gly 795	tct Ser	ttt Phe	gtc Val	ctg Leu	2400
gac Asp 800	Thr	ago Ser	cga Arg	gat Asp	ggc Gly 805	. Pàs	cct Pro	gtt Val	tct Ser	tac Tyr 810	Tyr	tta Leu	agt Ser	gat Asp	aac Asn 815	2448
_			ttg Lev		ı											2463
	Λ- /	- 0														

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<212> PRT

<213> Alloiococcus otitidis

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Asp Val Met Gly Asp Arg Phe Gly Arg Tyr Ser Lys Tyr Ile Ile Gln 20 25 30

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- Glu Arg Ala Leu Pro Asp Leu Arg Asp Gly Leu Lys Pro Val Gln Arg 35 40 45
- Arg Ile Leu Tyr Ala Met His Gln Asp Lys Asn Thr Tyr Asp Lys Ala 50 55 60
- Tyr Arg Lys Ser Ala Lys Thr Val Gly Asn Val Ile Gly Asn Tyr His 65 70 75 80
- Pro His Gly Asp Thr Ser Val Tyr Asp Ala Met Val Arg Leu Ser Gln 85 90 95
- Pro Trp Lys Met Arg His Pro Leu Val Asp Met His Gly Asn Lys Gly 100 105 110
- Ser Met Asp Gly Asp Pro Pro Ala Ala Met Arg Tyr Thr Glu Ala Arg 115 120 125
- Leu Ser Lys Ile Ala Ser Asp Leu Leu Ala Asp Ile Asp Lys Glu Thr 130 135 140
- Val Asp His Val Leu Asn Phe Asp Asp Thr Thr Glu Glu Pro Thr Val 145 150 155
- Leu Pro Ala Arg Phe Pro Asn Leu Leu Val Asn Gly Ala Ser Gly Ile 165 170 175
- Ser Ala Gly Tyr Ala Thr Asp Ile Pro Pro His Asn Leu Ser Glu Val 180 185 190
- Ile Asp Ala Thr Ile His Leu Ile Asn His Pro Asn Ala Arg Leu Glu 195 200 205
- Thr Leu Met Asp Tyr Ile Gln Gly Pro Asp Phe Pro Thr Gly Gly Ile 210 215 220
- Ile Gln Gly Lys Ser Gly Leu Lys Lys Ala Tyr Gln Thr Gly Lys Gly 225 230 235 240
- Lys Ile Ile Ile Arg Ala Lys Ala Asp Ile Glu Ala Ile Arg Gly Gly 245 250 255

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- Lys Ser Gln Ile Val Ile Ser Gln Ile Pro Tyr Glu Val Asn Lys Ala 260 265 270
- Arg Leu Val Gln Lys Ile Asp Asp Ile Arg Ile Asn Lys Lys Ile Asp 275 280 285
- Gly Ile Ala Asp Val Arg Asp Glu Ser Asp Arg Ser Gly Leu Arg Ile 290 295 300
- Val Val Glu Thr Lys Lys Asp Gly Asp Gly Glu Gly Ile Leu Thr Tyr 305 310 315 320
- Leu Leu Lys Asn Thr Asp Leu Gln Val Thr Tyr Asn Leu Asn Met Val
- Ala Ile Asp Lys Lys Arg Pro Gln Gln Val Ser Leu Lys Gln Ile Leu 340 345 350
- Ser Ser Tyr Leu Asp His Lys Arg Thr Val Val Gln Asn Arg Thr Arg 355 360 365
- Tyr Leu Leu Ala Lys Ala Lys Asp Arg Gln His Ile Val Gln Gly Leu 370 375 380
- Ile Lys Ala Ile Ser Ile Leu Asp Asp Leu Ile Gln Thr Ile Arg Ala 385 390 395 400
- Ser Glu Asn Lys Ala Asn Ala Lys Glu Asn Ile Ile Gln Ala Tyr Gly
  405 410 415
- Phe Ser Gln Asp Gln Ala Glu Ala Ile Val Ser Leu Gln Leu Tyr Arg 420 425 430
- Leu Thr Asn Thr Asp Ile Lys Asp Leu Gln Ala Glu Ala Lys Asp Leu 435 440 445
- Ala Gln Ala Ile Leu Thr Tyr Gln Asp Leu Leu Thr Asn Lys Ala Ser 450 455 460
- Leu Asp Ala Leu Met Lys Glu Glu Leu Lys Glu Val Lys Gln Ala Tyr 465 470 475 480

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Gly Glu Asp Arg Leu Thr Gln Val Gln Asp Lys Ile Glu Lys Leu Glu 485 490 495

Ile Glu Thr Gln Val Leu Val Ser Glu Glu Asp Val Met Val Thr Val 500 505 510

Thr Gln Gly Gly Tyr Leu Lys Arg Thr Ser Ile Arg Ser Tyr Lys Ala 515 520 525

Ser Gln Val Glu Glu Leu Gly Arg Arg Glu Asp Asp Leu Val Ile Phe 530 535 540

Met Gln Glu Leu Ser Thr Leu Asp Gln Leu Leu Ile Phe Thr Ser Lys 545 550 555 560

Gly Asn Val Val Asn Arg Pro Val His Glu Leu Pro Asp Ile Lys Trp 565 570 575

Lys Asp Ile Gly Glu His Leu Ser Arg Thr Ile Pro Leu Gly Glu Asp 580 585 590

Glu Glu Leu Ile Lys Val Tyr Pro Tyr Arg Glu Leu Asp Ala Gly Lys
595 600 605

Arg Tyr Val Phe Ile Thr Arg Asp Gly Tyr Ile Lys Gln Ser Pro Glu 610 615 620

Thr Glu Phe Glu Pro Lys Arg Thr Tyr Lys Ser Arg Ala Ser Thr Ala 625 630 635 640

Ile Lys Leu Lys Ser Asp Gln Asp Arg Leu Gln Ala Val Tyr Tyr Ile 645 650 655

Pro Asp Gln Glu Asp Tyr Asp Val Phe Leu Ala Ser Tyr Lys Gly Tyr 660 665 670

Gly Leu Lys Tyr Gly Leu Glu Glu Val Ser Glu Val Gly Ala Gln Ala 675 680 685

Ala Gly Val Lys Ser Met Asn Leu Lys Glu Gly Asp His Val Gln Asp 690 695 700

Gly Leu Val Phe Lys Arg Lys Gln Phe Gln Glu Ala Leu Phe Ile Thr

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720 705 710 715 Gln Arg Ala Ser Val Lys Lys Met Ala Leu His Asp Phe Asp Arg Thr 730 725 Ser Arg Ala Lys Arg Gly Leu Gln Ile Leu Arg Glu Leu Lys Arg Asn Pro His Arg Ile Gln Phe Met Ile Gly Ile Ser Gln Asn Lys Phe Leu Val Asn Leu Leu Thr Asp Thr Lys Lys Leu Val Gln Ile Asn Pro Asp 775 Asp Tyr Thr Val Ser Asn Arg His Asn Asn Gly Ser Phe Val Leu Asp 790 795 Thr Ser Arg Asp Gly Lys Pro Val Ser Tyr Tyr Leu Ser Asp Asn Asp 810 805 Ser His Leu <210> 61 <211> 1113 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (4)..(1113) <223> <400> 61 48 tta gtg gtt gag aca aaa tca aaa cta gaa aat gca gta aac acc ctc Met Val Glu Thr Lys Ser Lys Leu Glu Asn Ala Val Asn Thr Leu 96 att aaa gac ttg aaa aat aaa aaa gag tcg acc att tct tat att gac Ile Lys Asp Leu Lys Asn Lys Lys Glu Ser Thr Ile Ser Tyr Ile Asp 25 ctc agc aac aaa att gct gaa ccc ttc gaa ctt gaa agt gaa gcc atg Leu Ser Asn Lys Ile Ala Glu Pro Phe Glu Leu Glu Ser Glu Ala Met 192 gac aag tta atc cag caa tta gaa gat gat ggg att ggt gta gtt gac Asp Lys Leu Ile Gln Gln Leu Glu Asp Asp Gly Ile Gly Val Val Asp 50

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caa Gln	gac Asp 65	ggt Gly	aat Asn	ccc Pro	ttg Leu	gcc Ala 70	aag Lys	caa Gln	cta Leu	gcc Ala	aag Lys 75	cag Gln	gaa Glu	gaa Glu	gaa Glu	240
gca Ala 80	gaa Glu	aaa Lys	gcc Ala	aag Lys	gat Asp 85	gaa Glu	gaa Glu	atg Met	ata Ile	gcc Ala 90	cca Pro	cct Pro	Gly ggg	gtt Val	aaa Lys 95	288
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ctt Leu	tta Leu	gat Asp	gct Ala 115	gaa Glu	gaa Glu	gaa Glu	gtg Val	gcc Ala 120	cta Leu	gcc Ala	aag Lys	cgg Arg	att Ile 125	gaa Glu	gaa Glu	384
ggc Gly	gat Asp	gaa Glu 130	atc Ile	gct Ala	aaa Lys	caa Gln	gaa Glu 135	cta Leu	gct Ala	gag Glu	gct Ala	aac Asn 140	ttg Leu	aga Arg	ctg Leu	432
gtt Val	gtc Val 145	tct Ser	att Ile	gct Ala	aaa Lys	cgg Arg 150	tac Tyr	gtt Val	ggc Gly	cgg Arg	ggc Gly 155	atg Met	agc Ser	ttt Phe	ttg Leu	480
gac Asp 160	ttg Leu	atc Ile	cag Gln	gaa Glu	ggg Gly 165	aat Asn	atg Met	Gly	cta Leu	atg Met 170	aag Lys	gca Ala	gtt Val	gaa Glu	aaa Lys 175	528
ttt Phe	gac Asp	tac Tyr	gaa Glu	aaa Lys 180	ggt Gly	ttc Phe	aaa Lys	ttt Phe	tca Ser 185	Thr	tat Tyr	gcc Ala	acc Thr	tgg Trp 190	tgg Trp	576
atc Ile	cgt Arg	caa Gln	gcc Ala 195	Ile	act Thr	cgg Arg	gcc Ala	att Ile 200	gcc Ala	gac Asp	caa Gln	gcc Ala	cga Arg 205	acc Thr	atc Ile	624
cgg Arg	att Ile	ccg Pro 210	Val	cac His	atg Met	gtc Val	gaa Glu 215	act Thr	att Ile	aac Asn	aag Lys	ctg Leu 220	Val	cga Arg	atc Ile	672
cag Gln	egg Arg 225	cag Gln	ctc Leu	cta Leu	caa Gln	gaa Glu 230	cta Leu	ggc	cgg Arg	gaa Glu	cca Pro 235	Thr	cca Pro	gaa Glu	gaa Glu	720
att Ile 240	Gly	gca Ala	gag Glu	atg Met	gat Asp 245	ttg Leu	cca Pro	acc Thr	gaa Glu	aaa Lys 250	Val	aga Arg	gat Asp	att Ile	ttg Leu 255	768
aaa Lys	att	tcc Ser	caa Gln	gaa Glu 260	Pro	gtc Val	tcc Ser	ctt Leu	gaa Glu 265	Thr	cca Pro	att Ile	Gly	gaa Glu 270	gaa Glu	816
gaa Glu	gat Asp	tcc Ser	cac His 275	Leu	gga Gly	gac Asp	ttt Phe	att Ile 280	Glu	gat Asp	gat Asp	ggg Gly	gcc Ala 285	Lev	tcg Ser	864
cca	tct	gat	aat	gca	gct	tat	gag	ctg	ttg	g aaa	ggg	gaa	cto	aaa	gga	912

Pro	Ser	Asp 290	Asn	Ala	Ala	Tyr	Glu 295	Ļeu	Leu	Lys	Gly	Glu 300	Leu	Lys	Gly	
gtc Val	tta Leu 305	gac Asp	acc Thr	cta Leu	act Thr	gac Asp 310	cgg Arg	gaa Glu	gaa Glu	aat Asn	gtc Val 315	ttg Leu	cgc Arg	ctc Leu	cgt Arg	960
ttt Phe 320	ggc Gly	cta Leu	gat Asp	gat Asp	ggc Gly 325	cgt Arg	caa Gln	cgt Arg	act Thr	tta Leu 330	gaa Glu	gat Asp	gtc Val	ggt Gly	aag Lys 335	1008
gtc Val	ttt Phe	Gly ggg	gtc Val	acc Thr 340	cgg Arg	gag Glu	cgg Arg	atc Ile	egt Arg 345	caa Gln	att Ile	gaa Glu	gcg Ala	aag Lys 350	gcc Ala	1056
ctc Leu	cgc Arg	aaa Lys	ctc Leu 355	cgc Arg	cac His	cct Pro	agc Ser	cgg Arg 360	tcc Ser	aaa Lys	caa Gln	tta Leu	aaa Lys 365	gac Asp	ttt Phe	1104
	gaa Glu	_														1113
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Met 1	Val	Glu							10					15		
Met 1 Lys	Val Asp	Glu	Lys 20	5 Asn	Lys	Lys	Glu	Ser 25	10 Thr	Ile	Ser	Tyr	Ile 30	15 Asp		
Met 1 Lys Ser	Val Asp Asn	Leu Lys 35	Lys 20	5 Asn Ala	Lys	Lys Pro	Glu Phe 40	Ser 25 Glu	Thr	Ile Glu	Ser	Tyr Glu 45	Ile 30 Ala	15 Asp Met	Leu	
Met 1 Lys Ser	Asp Asn Leu 50	Leu Lys 35	Lys 20 Ile	Asn Ala	Lys Glu Leu	Lys Pro Glu 55	Glu Phe 40	Ser 25 Glu Asp	Thr Leu Gly	Ile Glu Ile	Ser Ser Gly	Tyr Glu 45 Val	Ile 30 Ala Val	Asp Met	Leu Asp	
Lys Lys Lys Asp	Asp Asn Leu 50	Leu Lys 35	Lys 20 Ile Gln	Asn Ala Gln	Lys Glu Leu Ala 70	Pro Glu 55 Lys	Phe 40 Asp	Ser 25 Glu Asp	Thr Leu Gly	Ile Glu Ile	Ser Ser Gly 60	Tyr Glu 45 Val	Ile 30 Ala Val	Asp Met Asp	Leu Asp Gln Ala	

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Leu Asp Ala Glu	Glu Glu Val	l Ala Leu Ala	Lys Arg Ile	e Glu Glu Gly
115		120	12:	

- Asp Glu Ile Ala Lys Gln Glu Leu Ala Glu Ala Asn Leu Arg Leu Val 130 135 140
- Val Ser Ile Ala Lys Arg Tyr Val Gly Arg Gly Met Ser Phe Leu Asp 145 150 155 160
- Leu Ile Gln Glu Gly Asn Met Gly Leu Met Lys Ala Val Glu Lys Phe 165 170 175
- Asp Tyr Glu Lys Gly Phe Lys Phe Ser Thr Tyr Ala Thr Trp Trp Ile 180 185 190
- Arg Gln Ala Ile Thr Arg Ala Ile Ala Asp Gln Ala Arg Thr Ile Arg 195 200 205
- Ile Pro Val His Met Val Glu Thr Ile Asn Lys Leu Val Arg Ile Gln 210 220
- Arg Gln Leu Gln Glu Leu Gly Arg Glu Pro Thr Pro Glu Glu Ile 225 230 235 240
- Gly Ala Glu Met Asp Leu Pro Thr Glu Lys Val Arg Asp Ile Leu Lys 245 250 255
- Ile Ser Gln Glu Pro Val Ser Leu Glu Thr Pro Ile Gly Glu Glu Glu 260 265 270
- Asp Ser His Leu Gly Asp Phe Ile Glu Asp Asp Gly Ala Leu Ser Pro 275 280 285
- Ser Asp Asn Ala Ala Tyr Glu Leu Leu Lys Gly Glu Leu Lys Gly Val 290 295 300
- Leu Asp Thr Leu Thr Asp Arg Glu Glu Asn Val Leu Arg Leu Arg Phe 305 310 315 320
- Gly Leu Asp Asp Gly Arg Gln Arg Thr Leu Glu Asp Val Gly Lys Val 325 330 335
- Phe Gly Val Thr Arg Glu Arg Ile Arg Gln Ile Glu Ala Lys Ala Leu

350

340

Arg Lys Leu Arg His Pro Ser Arg Ser Lys Gln Leu Lys Asp Phe Leu 355 360 365

345

333

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tat Tyr 145	ctc Leu	cag Gln	aat Asn	cgt Arg	ggg Gly 150	att Ile	tcc Ser	aga Arg	gag Glu	gtg Val 155	atg Met	gaa Glu	gag Glu	ttc Phe	gaa Glu 160	480
ctg Leu	ggt Gly	tat Tyr	tct Ser	ccc Pro 165	agc Ser	caa Gln	agg Arg	gag Glu	tcg Ser 170	ctc Leu	cac His	ctt Leu	tat Tyr	ttg Leu 175	cag Gln	528
					gac Asp											576
ctt Leu	ttt Phe	tcc Ser 195	aaa Lys	aga Arg	gaa Glu	gtg Val	gaa Glu 200	agt Ser	gat Asp	agt Ser	ttt Phe	aaa Lys 205	gac Asp	cgc Arg	ttt Phe	624
					ttc Phe											672
ggc Gly 225					tat Tyr 230											720
					aac Asn											768
					tac Tyr											816
					ttc Phe											864
					aat Asn											912
					acc Thr 310										ttg Leu 320	960
					gaa Glu										ctg Leu	1008
					acc Thr											1056
					ccg Pro											1104
gcc	ttt	caa	aat	ctc	atc	caa	cat	ggt	agg	atg	act	gtc	tac	caa	ttc	1152

Ala	Phe 370	Gln	Asn	Leu	Ile	Gln 375	His	Gly	Arg	Met	Thr 380	Val	Tyr	Gln	Phe	
tta Leu 385	aaa Lys	gaa Glu	tac Tyr	ttt Phe	aaa Lys 390	aaa Lys	tcc Ser	tac Tyr	aat Asn	cta Leu 395	gat Asp	aac Asn	gac Asp	tcg Ser	gac Asp 400	1200
cgg Arg	ttg Leu	aaa Lys	ttt Phe	atc Ile 405	caa Gln	acc Thr	atg Met	acc Thr	aat Asn 410	aaa Lys	att Ile	ggc Gly	aag Lys	cta Leu 415	gct Ala	1248
tcc Ser	ccc Pro	ttg Leu	gaa Glu 420	agg Arg	gaa Glu	gtc Val	tat Tyr	gcc Ala 425	aag Lys	gat Asp	ttg Leu	gca Ala	gaa Glu 430	gaa Glu	ttt Phe	1296
aac Asn	ctg Leu	tct Ser 435	tat Tyr	gat Asp	acg Thr	att Ile	ata Ile 440	agc Ser	caa Gln	gtt Val	caa Gln	agt Ser 445	gaa Glu	gcc Ala	act Thr	1344
cta Leu	aac Asn 450	cag Gln	caa Gln	gag Glu	gct Ala	ttg Leu 455	aaa Lys	aag Lys	gac Asp	cgg Arg	cat His 460	aag Lys	gaa Glu	ttt Phe	tct Ser	1392
caa Gln 465	Ala	aga Arg	gtg Val	gaa Glu	gtc Val 470	aaa Lys	gcc Ala	cca Pro	agt Ser	agt Ser 475	caa Gln	aag Lys	act Thr	aag Lys	att Ile 480	1440
gac Asp	cgg Arg	gcc Ala	cag Gln	gaa Glu 485	aaa Lys	ctt Leu	'tta Leu	aac Asn	cga Arg 490	ctc Leu	ttt Phe	tac Tyr	tat Tyr	ccc Pro 495	caa Gln	1488
									ccg Pro							1536
gaa Glu	gtc Val	cac His 515	cag Gln	cgg Arg	att Ile	tac Tyr	ctc Leu 520	Leu	ttt Phe	tta Leu	gaa Glu	tac Tyr 525	agc Ser	cag Gln	gaa Glu	1584
		Ser							ttt Phe			Asp				1632
	Glu								aca Thr							1680
					Leu				gac Asp 570	Tyr						1728
ccc Pro	ctg Leu	gag Glu	caa Gln 580	Lys	cgc Arg	caa Gln	gac Asp	tgc Cys 585	ttg Leu	gag Glu	gaa Glu	gtc Val	aaa Lys 590	gca Ala	gct Ala	1776
aaa Lys	cag Gln	tcc Ser	ggt Gly	aat Asn	aag Lys	aag Lys	cga Arg	gag Glu	ctg Leu	gaa Glu	tta Leu	acc Thr	aat Asn	caa Gln	tta Leu	1824

1854

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595 600 605

att gaa ata aac cgt atg cta aaa caa taa Ile Glu Ile Asn Arg Met Leu Lys Gln  $\,$ 

610 615

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<211> 617

<212> PRT <213> Alloiococcus otitidis

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Ala Asn Tyr Phe Ala His Cys Pro Phe His Glu Asp Ser Thr Pro Ser 35 40 45

Phe Ser Val Asn Arg Asp Lys Gln Ile Tyr Lys Cys Phe Ser Cys Lys 50 60

Arg Gly Gly Ser Val Phe Ser Phe Ile Gln Glu Lys Glu Gly Leu Ser 65 70 75 80

Phe Pro Glu Ser Val Leu Lys Val Ala Asp Leu Ala Asn Val Asp Leu 85 90 95

Asp Pro Ala Leu Lys Glu Ala Val Gln Gly Gln Pro Asp Lys Ala Asp 100 105 110

Ser Pro Tyr Arg Asp Leu Tyr Thr Ile His Asp Gln Ala Lys Asp Tyr 115 120 125

Tyr Gln Tyr Ile Leu Leu Lys Ala Gln Val Gly Glu Val Ala Tyr Asp 130 135 140

Tyr Leu Gln Asn Arg Gly Ile Ser Arg Glu Val Met Glu Glu Phe Glu 145 150 155 160

Leu Gly Tyr Ser Pro Ser Gln Arg Glu Ser Leu His Leu Tyr Leu Gln 165 170 175

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Ser Gln Asp Gln Ala Asp Leu Thr Asp Asp Leu Leu Glu Glu Thr Gly 185 Leu Phe Ser Lys Arg Glu Val Glu Ser Asp Ser Phe Lys Asp Arg Phe 200 Ala Lys Arg Ile Ile Phe Pro Leu Lys Asn Leu Gln Gly Gln Thr Val 210 215 Gly Phe Ser Gly Arg Tyr Phe Gln Asp Glu Pro Asn Gln Asp Phe His 225 230 His Ala Lys Tyr Leu Asn Ser Pro Glu Thr Lys Ile Phe Asn Lys Arg Arg Thr Leu Phe Asn Tyr His Gln Ala Lys Ala Tyr Ile Arg Arg Ala 260 265 Lys Glu Val Val Leu Phe Glu Gly Tyr Met Asp Val Ile Ala Ala Trp Gln Ala Gly Val Lys Asn Gly Leu Ala Ser Met Gly Thr Ser Ile Thr 295 Ala Asp Gln Val Gln Thr Met Gln Arg Ile Ala Asp Thr Leu Val Leu 305 310 315 Ala Phe Asp Gly Asp Glu Ala Gly Leu Glu Ser Ser Lys Lys Ile Leu 325 330 Asp Asp Leu Ser Leu Thr Ser Lys Leu Gln Ile Glu Val Val Ile Phe Pro Lys Lys Met Asp Pro Asp Glu Tyr Ile Arg Glu Asn Gly Pro Glu 360 Ala Phe Gln Asn Leu Ile Gln His Gly Arg Met Thr Val Tyr Gln Phe 375 Leu Lys Glu Tyr Phe Lys Lys Ser Tyr Asn Leu Asp Asn Asp Ser Asp 390 395

Arg Leu Lys Phe Ile Gln Thr Met Thr Asn Lys Ile Gly Lys Leu Ala

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405 410 415

Ser Pro Leu Glu Arg Glu Val Tyr Ala Lys Asp Leu Ala Glu Glu Phe 420 425 430

Asn Leu Ser Tyr Asp Thr Ile Ile Ser Gln Val Gln Ser Glu Ala Thr 435 440 445

Leu Asn Gln Glu Ala Leu Lys Lys Asp Arg His Lys Glu Phe Ser 450 455 460

Gln Ala Arg Val Glu Val Lys Ala Pro Ser Ser Gln Lys Thr Lys Ile 465 470 475 480

Asp Arg Ala Gln Glu Lys Leu Leu Asn Arg Leu Phe Tyr Tyr Pro Gln 485 490 495

Val Gln Glu Ile Ile Asp Ala Tyr Asn Pro Asp Phe Glu Phe Lys Thr 500 505 510

Glu Val His Gln Arg Ile Tyr Leu Leu Phe Leu Glu Tyr Ser Gln Glu 515 520 525

Asn Asp Ser Ile Asp Ser Phe Ile Asp Phe Val Lys Asp Lys Glu Thr 530 540

Lys Glu Val Ile Ser Asp Ile Met Trp Thr Ser Ile Glu Val Glu Pro 545 550 555

Ser Asp Glu Glu Ile Leu Asp Tyr Leu Asp Tyr Ile Asp Gln Thr Tyr 565 570 575

Pro Leu Glu Gln Lys Arg Gln Asp Cys Leu Glu Glu Val Lys Ala Ala 580 585 590

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ctc aac caa ata Leu Asn Gln Ile 35	tta ggc ca Leu Gly Gl 40	g aag att a n Lys Ile 1	acc att atc a Thr Ile Ile So 45	gt gac aaa er Asp Lys	ccc 201 Pro									
caa aca acc cgg Gln Thr Thr Arg 50	r aat aaa at r Asn Lys Il 55	c cag ggt a e Gln Gly :	att tac acc ga Ile Tyr Thr As 60	ac caa gcg sp Gln Ala	ggg 249 Gly 65									
caa att gtc ttt Gln Ile Val Phe	atc gac ac lle Asp Th 70	r Pro Gly I	ata cat aaa co Ile His Lys Pi 75	cc aag cac ro Lys His 80	cgc 297 Arg									
ctg ggc cgg ttt Leu Gly Arg Phe 85	atg gtg ga Met Val As	tcg gct a Ser Ala N 90	atg tcg acc at Met Ser Thr I	cc aat gag le Asn Glu 95	gtg 345 Val									
gac ctg gtc tta Asp Leu Val Leu 100	ttt gtg gt Phe Val Va	e aat gtc a l Asn Val <i>I</i> 105	agg gaa aag at Arg Glu Lys II 11	le Gly Pro	Gly aga 393									
gac cgg ttc att Asp Arg Phe Ile 115	atc gac aa Ile Asp Lys 12	Leu Arg 1	acc atc gat ac Thr Ile Asp Th 125	eg cca gtt ur Pro Val	ttt 441 Phe									
tta att att aac Leu Ile Ile Asn 130	cag att gad Gln Ile Asp 135	cag gtc g Gln Val A	gat cca aca ga Asp Pro Thr As 140	sp Leu Leu	ccg 489 Pro 145									
gtt att agc gac Val Ile Ser Asp	tac caa gag Tyr Gln Glu 150	Glu Phe A	gac ttt gcc ga Asp Phe Ala Gl .55	a gtg gtt u Val Val 160	cca 537 Pro									
act tca ggc ttg Thr Ser Gly Leu 165	gaa ggg gaa Glu Gly Glu	aat atc c Asn Ile G 170	ag gag ctc at In Glu Leu Il	e Gln Thr	atc 585 Ile									

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tcg gac cac ccc gaa tac ttt att att tca gaa ctc atc cgg gag aag Ser Asp His Pro Glu Tyr Phe Ile Ile Ser Glu Leu Ile Arg Glu Lys 195 200 205	681
gtt tta gac ttg gct aga gaa gag att cct cat tca gta gca gta gta Val Leu Asp Leu Ala Arg Glu Glu Ile Pro His Ser Val Ala Val Val 210 225	729
act gag aag gta gac cga aac caa gat ggt aaa gtc caa acc tat gcc Thr Glu Lys Val Asp Arg Asn Gln Asp Gly Lys Val Gln Thr Tyr Ala 230 235 240	777
acc att att gtc gaa cgc aag agc caa aag ggg att att atc ggc aag Thr Ile Ile Val Glu Arg Lys Ser Gln Lys Gly Ile Ile Ile Gly Lys 245 250 255	825
caa ggg tcc atg att aaa aaa att ggt agc cta gct cgg cga gat att Gln Gly Ser Met Ile Lys Lys Ile Gly Ser Leu Ala Arg Arg Asp Ile 260 265 270	873
gag aaa cta ctg gga gat aag att tac ttg gaa ctc tgg gtt aaa gtc Glu Lys Leu Leu Gly Asp Lys Ile Tyr Leu Glu Leu Trp Val Lys Val 275 280 285	921
caa aga gac tgg cgg gac aag ccc agt cgc tta gaa gac ttt ggc tac Gln Arg Asp Trp Arg Asp Lys Pro Ser Arg Leu Glu Asp Phe Gly Tyr 290 295 300 305	969
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Pro Gln Thr Thr Arg Asn Lys Ile Gln Gly Ile Tyr Thr Asp Gln Ala

40

35

Gly Gln Ile Val Phe Ile Asp Thr Pro Gly Ile His Lys Pro Lys His 65 70 75 80

Arg Leu Gly Arg Phe Met Val Asp Ser Ala Met Ser Thr Ile Asn Glu 85 90 95

Val Asp Leu Val Leu Phe Val Val Asn Val Arg Glu Lys Ile Gly Pro 100 105 110

Gly Asp Arg Phe Ile Ile Asp Lys Leu Arg Thr Ile Asp Thr Pro Val 115 120 125

Phe Leu Ile Ile Asn Gln Ile Asp Gln Val Asp Pro Thr Asp Leu Leu 130 135 140

Pro Val Ile Ser Asp Tyr Gln Glu Glu Phe Asp Phe Ala Glu Val Val 145 150 155 160

Pro Thr Ser Gly Leu Glu Gly Glu Asn Ile Gln Glu Leu Ile Gln Thr 165 170 175

Ile Lys Ser Tyr Leu Pro Val Gly Pro Gln Phe Tyr Pro Asp Asp Gln 180 185 190

Val Ser Asp His Pro Glu Tyr Phe Ile Ile Ser Glu Leu Ile Arg Glu 195 200 205

Lys Val Leu Asp Leu Ala Arg Glu Glu Ile Pro His Ser Val Ala Val 210 215 220

Val Thr Glu Lys Val Asp Arg Asn Gln Asp Gly Lys Val Gln Thr Tyr 225 230 235 240

Ala Thr Ile Ile Val Glu Arg Lys Ser Gln Lys Gly Ile Ile Gly 245 250 255

Lys Gln Gly Ser Met Ile Lys Lys Ile Gly Ser Leu Ala Arg Arg Asp 260 265 270

Ile Glu Lys Leu Leu Gly Asp Lys Ile Tyr Leu Glu Leu Trp Val Lys 275 280 285

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Val Gln Arg Asp Trp Arg Asp Lys Pro Ser Arg Leu Glu Asp Phe Gly 290 295 300

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Val	Met	Gly 135	Thr	Val	Ile	Gln	Gly 140	Ile	Ala	Ala	Gly	Met 145	Ile	Ile	Pro	
									atg Met							537
		-		-	_	_		_	att Ile					_	_	585
-			_			_		_	gat Asp 190	_				_		633
									ctt Leu							681
	_				_	_	-		aat Asn	_	-					729
									ggt Gly							777
									ttt Phe							825
							-		gtc Val 270	_	_					873
				_					aat Asn		_					921
			_		_	-	_	_	gcc Ala		_			_		969
		Cys		Ser		Ile	Ile	Pro	att Ile	Tyr						1017
	_		_			_			atc Ile		_		_			1065
									ggc Gly 350							1113
gga Gly	gtg Val	gct Ala	cgg Arg	att Ile	ggc Gly	ctt Leu	atc Ile	ggt Gly	ggt Gly	atc Ile	tta Leu	ctt Leu	tta Leu	gtt Val	Gly ggg	1161

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Asp	Asn	Leu	Gln	Ile	Pro	Arg	Trp	Gln	Ile	Phe	Ala	Val	Leu	Phe	Thr	

Gly Ala Val Ile Val Val Leu Asn Gln Thr Ala Met Ser Thr Ala Leu

60

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55

50

Pro Asn Met Ile Glu Ser Leu Gly Ile Asp Pro Ser Leu Gly Gln Trp 70 75 Ile Val Ser Gly Tyr Thr Leu Val Lys Gly Ile Met Val Pro Ile Thr Ala Phe Ala Met Thr Lys Tyr Arg Thr Arg Asn Phe Phe Ile Leu Met Leu Ala Leu Phe Cys Thr Gly Ser Phe Leu Thr Gly Leu Gly Phe Asn 115 120 125 Phe Pro Val Val Wat Gly Thr Val Ile Gln Gly Ile Ala Ala Gly 130 Met Ile Ile Pro Leu Met Gln Thr Val Leu Leu Thr Leu Met Pro Val 155 Glu Ser Arg Gly Thr Ala Met Gly Val Met Ser Gly Val Ile Gly Ile Gly Pro Ala Leu Gly Pro Leu Val Gly Gly Val Ile Val Asp Ala Phe 185 Thr Trp Glu Ile Leu Phe Tyr Ile Trp Ala Leu Ile Thr Leu Leu Leu 195 Val Pro Leu Thr Trp Leu Val Leu Pro Asp Val Leu Pro Asn Ala Asp 210 215

Leu Thr Ile Asn Trp Ala Asn Ile Arg Asp Ser Leu Ile Gly Phe Gly 225 230 235 240

Leu Leu Phe Ser Leu Ser Val Phe Gly Ser Ser Gly Phe Ser Ser 245 250 255

Val Ile Ala Trp Val Ser Leu Leu Ile Gly Leu Val Phe Val Ala Lys 260 265 270

Phe Ile His Phe Asn Leu Lys Ala Asp Gln Pro Ile Leu Asn Leu Arg 275 280 285

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Leu	Phe 290	Lys	Lys	Thr	Tyr	Tyr 295	Arg	Arg	Ala	Val	Leu 300	Val	Ala	Thr	Leu
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Gln	Thr	Val	Arg	Gly 325	Leu	Gly	Ala	Ser	Ile 330	Ala	Gly	Leu	Ile	Leu 335	Met
Pro	Ala	Gly	Ile 340	Ile	Lys	Thr	Ile	Leu 345	Ala	Pro	Ile	Ser	Gly 350	Lys	Leu
Tyr	Asp	Lys 355	Val	Gly	Val	Ala	Arg 360	Ile	Gly	Leu	Ile	Gly 365	Gly	Ile	Leu
Leu	Leu 370	Val	Gly	Ser	Leu	Leu 375	Leu	Val	Thr	Leu	Asn 380	Glu	Ala	Ser	Ser
Leu 385	Tyr	Leu	Leu	Met	Ile 390	Tyr	Tyr	Gly	Ile	Leu 395	Ser	Ala	Gly	Phe	Gly 400
Leu	Phe	Asn	Ile	Pro 405	Ile	Thr	Thr	Ala	Gly 410	Met	Asn	Ile	Met	Ala 415	Lys
Glu	Asp	Met	Gly 420	His	Ala	Thr	Ser	Ala 425	Arg	Gln	Thr	Val	Arg 430	Gln	Ile
Ser	Ser	Ser 435	Phe	Ala	Val	Ser	Leu 440	Ser	Phe	Ile	Ile	Met 445	Thr	Leu	Val
Thr	Ile 450	Ala	Thr	Ser	Gly	Gln 455	Ser	Val	Gly	Val	Phe 460	Gln	Asp	Gly	Gly
Pro 465	Thr	Asp	Leu	Asn	Met 470	Ala	Gly	Val	Arg	Gly 475	Ala	Phe	Ile	Leu	Val 480
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Pro Lys Glu Lys Pro Asp Gln 500

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cac His	tta Leu	gaa Glu	gaa Glu 20	cac His	cta Leu	caa Gln	gac Asp	cga Arg 25	ccc Pro	ctt Leu	ctt Leu	aaa Lys	gcc Ala 30	ggc	agt Ser		153
ttg Leu	aag Lys	caa Gln 35	att Ile	gtt Val	gtt Val	tac Tyr	aag Lys 40	gct Ala	caa Gln	caa Gln	gcc Ala	tgg Trp 45	gac Asp	ctg Leu	acc Thr		201
ctc Leu	caa Gln 50	ttt Phe	cct Pro	cag Gln	atc Ile	ctc Leu 55	cct Pro	ttt Phe	aag Lys	gac Asp	ttc Phe 60	caa Gln	gtt Val	ttg Leu	gag Glu		249
tct Ser 65	gcc Ala	ctc Leu	ttg Leu	cag Gln	cat His 70	atc Ile	cca Pro	gaa Glu	gtc Val	aac Asn 75	cag Gln	atc Ile	cat His	tta Leu	agg Arg 80		297
gtt Val	gat Asp	gcc Ala	caa Gln	gat Asp 85	gac Asp	agt Ser	ttt Phe	gac Asp	cag Gln 90	gac Asp	ctc Leu	ctc Leu	cag Gln	gac Asp 95	tat Tyr		345
tgg Trp	cct Pro	aag Lys	gcg Ala 100	gtg Val	aag Lys	ttt Phe	agc Ser	gga Gly 105	gtc Val	gat Asp	tct Ser	ccc Pro	ctt Leu 110	tgc Cys	aat Asn		393
gac Asp	tta Leu	cta Leu 115	Asp	Lys	acc Thr	Leu	Pro	Tyr	cta Leu	gat Asp	Gly gag	aag Lys 125	caa Gln	gtt Val	tac Tyr	٠	441
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cta Leu 145	cct Pro	cgg	atc Ile	caa Gln	gct Ala 150	ggc	tac Tyr	cag Gln	caa Gln	gtg Val 155	Gly	ttt 'Phe	ccc Pro	aac Asn	cac His 160		537
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Phe	Lys	Ile	Lys	Ala	Arg	Val	Asp	Ala	Gln	Lys	Asn	Ser	Asp	Gln	Ile	
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gag Glu	cta Leu	acc Thr 195	aac Asn	caa Gln	ttt Phe	atc Ile	aag Lys 200	gcc Ala	agc Ser	caa Gln	aag Lys	aaa Lys 205	gaa Glu	gaa Glu	Gly ggg	681
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gac Asp 225	cac His	gaa Glu	acg Thr	att Ile	acc Thr 230	cag Gln	atg Met	gtt Val	gat Asp	gtg Val 235	gaa Glu	gaa Glu	gaa Glu	gag Glu	agc Ser 240	777
cgt Arg	ctg Leu	acc Thr	ttt Phe	gaa Glu 245	gga Gly	tac Tyr	gtt Val	ttt Phe	gat Asp 250	gtg Val	gaa Glu	atc Ile	aaa Lys	tcc Ser 255	ctc Leu	825
cgg Arg	tca Ser	gat Asp	aga Arg 260	Lys	ctc Leu	ctt Leu	ctc Leu	ttt Phe 265	Lys	atg Met	acc Thr	gac Asp	tat Tyr 270	agc Ser	tct Ser	873
tcc Ser	ttc Phe	cta Leu 275	Phe	aaa Lys	aaa Lys	ttc Phe	tct Ser 280	Asn	aat Asn	tct Ser	tct Ser	gac Asp 285	gaa Glu	gcc Ala	cta Leu	921
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caa Gln	gag Glu	gtc Val	aaa Lys	aaa Lys 325	Glu	ccc Pro	cgc Arc	cgg Arg	gac Asp 330	Leu	gct Ala	aag Lys	gaa Glu	ggg Gly 335	GIU	
aag Lys	agg Arg	gtg Val	gaa Glu 340	ı Lev	cat His	: Ala	His	Thi	acc Thr	. Met	g agt : Sei	cag Gln	atg Met 350	: Asp	ggt Gly	1113
ttg Lev	g gtg 1 Val	ccg Pro	Ala	aag Lys	gat s As <u>r</u>	tto Lev	gto 1 Val 360	Lys	g caa s Glr	a gca n Ala	gco A Ala	gct Ala 365	. Phe	gac Asp	caa Gln	1161
ccg	g gct Ala 370	a Ile	gco Ala	ato a Ile	e act	gat Asy 375	Hi	gct S Ala	gta a Val	a gto L Val	caa L Gli 380	ı Ser	tto Phe	c cca e Pro	gag Glu	1209
gco Ala	cat a His	tat	gct Ala	ggo a Gly	tta y Le	a gad 1 Asj	act	gg; c Gl	t gtt y Val	t aaa l Lys	a at	t ctt e Lev	tac ı Tyr	ggt Gly	gtg Val	1257

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acc Thr	gga Gly	cta Leu 435	tcg Ser	gct Ala	cgt Arg	tat Tyr	gac Asp 440	caa Gln	atc Ile	att Ile	gaa Glu	ttg Leu 445	gcc Ala	gct Ala	gtg Val	140	1
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									acc Thr							149	7
									gaa Glu 490							154	5
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gct Ala	tac Tyr 610	aag Lys	cag Gln	gcc Ala	cgg Arg	cca Pro 615	acc Thr	cat His	gcc Ala	agt Ser	att Ile 620	ttg Leu	gtc Val	aag Lys	aat Asn	1929	•

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cag Gln	gac Asp 690	tat Tyr	gat Asp	tat Tyr	att Ile	gaa Glu 695	gtt Val	atg Met	ccc Pro	aag Lys	tca Ser 700	gcc Ala	tat Tyr	att Ile	gac Asp	. 2169
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ctc	tat Tyr	gcc Ala	ccg Pro 820	Lys	atg Met	gaa Glu	GJÀ	tcg Ser 825	Asp	caa Gln	gag Glu	ata Ile	cgt Arg 830	Gln	atg Met	2553
agt Ser	tac Tyr	aag Lys 835	Gln	gcc Ala	aag Lys	gct Ala	ctc Leu 840	Tyr	Gly	gac Asp	ccc Pro	ttg Leu 845	Pro	agt Ser	att	2601

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gaa gat ggc tat ttg gtt ggt tcc agg ggg tcg gtt ggg tca agc ttt 2745 Glu Asp Gly Tyr Leu Val Gly Ser Arg Gly Ser Val Gly Ser Ser Phe 890

2793 gtg gcc acc atg acc ggg atc aca gaa gtc aac cca cta ccg ccc cac Val Ala Thr Met Thr Gly Ile Thr Glu Val Asn Pro Leu Pro Pro His 900 905 910

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2889 gtg ggg tcc ggc ttt gac tta gag gcc aaa aaa tgt ccg gaa tgt caa Val Gly Ser Gly Phe Asp Leu Glu Ala Lys Lys Cys Pro Glu Cys Gln 930 935

age cta atg gaa tea gae gge cae gae att eee tte gaa ace tte ett 2937 Ser Leu Met Glu Ser Asp Gly His Asp Ile Pro Phe Glu Thr Phe Leu 950

2985 ggt ttt aat ggg gac aag gtg cca gat atc gat ttg aac ttc tca ggt Gly Phe Asn Gly Asp Lys Val Pro Asp Ile Asp Leu Asn Phe Ser Gly 965 970

gaa tac cag gcc aag gcc cac aac tat acc aag gtt ttg ttt gga gaa 3033 Glu Tyr Gln Ala Lys Ala His Asn Tyr Thr Lys Val Leu Phe Gly Glu 985

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gcc ttt ggt ttt gtc aag ggt tat gaa agg gac aag cag ata aac tac 3129 Ala Phe Gly Phe Val Lys Gly Tyr Glu Arg Asp Lys Gln Ile Asn Tyr 1010 1015 1020

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cgg tca acc ggc cag cac cca gga ggg att atc gtc ata ccg gat gac 3225 Arg Ser Thr Gly Gln His Pro Gly Gly Ile Ile Val Ile Pro Asp Asp 1045

atg gat gtg ttt gat ttc acc ccc atc cag tac ccg gct gac gac cag 3273 Met Asp Val Phe Asp Phe Thr Pro Ile Gln Tyr Pro Ala Asp Asp Gln 1065

acg get gag tgg caa act acc cac ttt gac ttc cac tec atc gac gaa 3321

Thr	· Ala	Glu 107	Trp 5	Gln	Thr	Thr	His 108		Asp	Phe	His	Ser 108		Asp	Glu	
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cga Arg 110	Lys	ctc Leu	cag Gln	gac Asp	ttg Leu 111	Ser	Gly	ttt Phe	gac Asp	cct Pro 111	Gln	gaa Glu	ata Ile	ccg Pro	gta Val 1120	3417
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ctt cgt gct cca ttt agg Leu Arg Ala Pro Phe Arg 1380	g gca gtc cct to g Ala Val Pro Se 1385	cc ttg ggg tcc agt er Leu Gly Ser Ser 1390	Ala Ala			
.cag gct gtc att gat gcc Gln Ala Val Ile Asp Ala 1395	c agg gag gac ag Arg Glu Asp Se 1400	gc gac ttc ttg tcc er Asp Phe Leu Ser 1405	aag gaa 4281 Lys Glu			
gac cta tca aaa cgg ggc Asp Leu Ser Lys Arg Gly 1410	aag ttg tcg aa Lys Leu Ser Ly 1415	aa acg gtc atg gac ys Thr Val Met Asp 1420	tac ctg 4329 Tyr Leu			
gac aat aac cac gtt tta Asp Asn Asn His Val Let 1425 143	ı Asp His Leu Pı	eg gac gaa aac caa ro Asp Glu Asn Gln 1435	ctt tcc 4377 Leu Ser 1440			
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Leu Gln Phe Pro Gln Ile Leu Pro Phe Lys Asp Phe Gln Val Leu Glu

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55 60 50 Ser Ala Leu Leu Gln His Ile Pro Glu Val Asn Gln Ile His Leu Arg Val Asp Ala Gln Asp Asp Ser Phe Asp Gln Asp Leu Leu Gln Asp Tyr 85 90 Trp Pro Lys Ala Val Lys Phe Ser Gly Val Asp Ser Pro Leu Cys Asn Asp Leu Leu Asp Lys Thr Leu Pro Tyr Leu Asp Gly Lys Gln Val Tyr Phe Asp Leu Asp His Glu Val Thr Arg Asp Lys Phe Asp His Asp Phe 130 135 Leu Pro Arg Ile Gln Ala Gly Tyr Gln Gln Val Gly Phe Pro Asn His 145 155 Phe Lys Ile Lys Ala Arg Val Asp Ala Gln Lys Asn Ser Asp Gln Ile Ala Ala Phe Arg Lys Glu Lys Glu Glu Lys Asp Gln Ala Leu Ser Gln 180 Glu Leu Thr Asn Gln Phe Ile Lys Ala Ser Gln Lys Lys Glu Glu Gly 200 Gly Ser Lys Ala Lys Ser Glu Ala Leu Lys Met Gly Arg Ala Ile Pro Asp His Glu Thr Ile Thr Gln Met Val Asp Val Glu Glu Glu Glu Ser 225 Arg Leu Thr Phe Glu Gly Tyr Val Phe Asp Val Glu Ile Lys Ser Leu 245 Arg Ser Asp Arg Lys Leu Leu Phe Lys Met Thr Asp Tyr Ser Ser 260 265 Ser Phe Leu Phe Lys Lys Phe Ser Asn Asn Ser Ser Asp Glu Ala Leu 280

275

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Phe	Asp	Gln	Val	Gln	Glu	Gly	Met	$\mathtt{Trp}$	Leu	Lys	Val	Arg	Gly	Ser	Val
	290					295					300				

- Gln Glu Asp Thr Phe Val Lys Asp Leu Val Val Met Ala Gln Asp Ile 310 315 305
- Gln Glu Val Lys Lys Glu Pro Arg Arg Asp Leu Ala Lys Glu Gly Glu 330 325
- Lys Arg Val Glu Leu His Ala His Thr Thr Met Ser Gln Met Asp Gly 340 345
- Leu Val Pro Ala Lys Asp Leu Val Lys Gln Ala Ala Ala Phe Asp Gln 360
- Pro Ala Ile Ala Ile Thr Asp His Ala Val Val Gln Ser Phe Pro Glu 375
- Ala His Tyr Ala Gly Leu Asp Thr Gly Val Lys Ile Leu Tyr Gly Val 395
- Glu Ala Asn Leu Val Ser Asp Gly Glu Leu Val Ala Tyr Asn Pro Ala 405 410
- Asp Ile Lys Leu Glu Glu Ala Thr Tyr Val Val Phe Asp Val Glu Thr 420
- Thr Gly Leu Ser Ala Arg Tyr Asp Gln Ile Ile Glu Leu Ala Ala Val
- Lys Met Glu Asn Gly Glu Ile Val Ser Glu Phe Gln Glu Phe Ile Asp 450
- Pro Gly Gln Pro Leu Ser Glu Thr Thr Thr Asn Leu Thr Gly Ile Thr 475 470
- Asp Asp Met Val Gln Gly Ser Lys Ser Glu Asp Glu Val Leu His Ala . 490
- Phe Gln Ala Phe Ser Glu Gly Thr Val Leu Val Ala His Asn Ala Ser 505

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Phe Asp Met Gly Phe Ile Asn Thr Ala Tyr Gln Arg His Gly Leu Gly 520 515 Gln Ala Asp Gln Pro Val Ile Asp Thr Leu Glu Leu Ser Arg Met Leu 535 His Pro Asn Leu Lys Ser His Arg Leu Asn Thr Leu Ala Lys Arg Tyr 550 Asp Val Ala Leu Glu His His His Arg Ala Ile Tyr Asp Ser Glu Ser 565 Thr Ala Lys Leu Leu Trp Ile Phe Leu Lys Glu Ala Lys Asp Gln Tyr 580 585 Asp Met Thr Ser His Gln Asp Leu Asn Ser Gln Val Gly Glu Gly Glu 600 595 Ala Tyr Lys Gln Ala Arg Pro Thr His Ala Ser Ile Leu Val Lys Asn Gln Lys Gly Leu Lys Asn Leu Phe Lys Ile Val Ser His Ala His Val 625 630 Asn Tyr Phe Tyr Arg Val Pro Arg Ile Pro Lys Ser Ile Leu Ser Lys 650 Tyr Arg Glu Gly Leu Leu Val Gly Ser Gly Cys Gly Gln Gly Glu Leu 665 Phe Glu Ala Ile Met Gln Lys Gly Tyr Asp Glu Ala Leu Ala Val Ala 675 Gln Asp Tyr Asp Tyr Ile Glu Val Met Pro Lys Ser Ala Tyr Ile Asp 690 695 Leu Leu Asp Arg Asp Leu Ile Lys Asp Glu Ala Thr Leu Glu Glu Met 710 705 Ile Glu Asn Leu Val Lys Ile Gly His Glu Leu Asp Ile Pro Val Val

730

725

Ala Thr Gly Asn Val His Tyr Leu Asn Pro Glu Asp Ala Val Leu Arg
740 745 750

Asp Ile Leu Leu Glu Thr Ala Lys Lys Gly Ala Phe Ser Lys Ala Arg 755 760 765

Asn Pro Glu Val His Phe Arg Thr Thr Asp Glu Met Leu Glu Glu Phe 770 775 780

Ser Phe Leu Gly Gln Asp Gln Ala Tyr Glu Ile Val Val Thr Asn Thr 785 790 795 800

Gln Lys Ile Ala Asp Ser Ile Glu Ser Ile Ser Pro Val Lys Glu Gly 805 810 815

Leu Tyr Ala Pro Lys Met Glu Gly Ser Asp Gln Glu Ile Arg Gln Met 820 825 830

Ser Tyr Lys Gln Ala Lys Ala Leu Tyr Gly Asp Pro Leu Pro Ser Ile 835 840 845

Val Glu Glu Arg Leu Glu Lys Glu Leu Lys Ser Ile Ile Asp Asn Asn 850 855 860

Phe Ser Val Ile Tyr Leu Ile Ser Gln Lys Leu Val Lys Lys Ser Val 865 870 875 880

Glu Asp Gly Tyr Leu Val Gly Ser Arg Gly Ser Val Gly Ser Ser Phe 885 890 895

Val Ala Thr Met Thr Gly Ile Thr Glu Val Asn Pro Leu Pro Pro His 900 905 910

Tyr Arg Cys Pro Asn Cys Gln His Thr Glu Phe Phe Thr Asn Gly Glu 915 920 925

Val Gly Ser Gly Phe Asp Leu Glu Ala Lys Lys Cys Pro Glu Cys Gln 930 935 940

Ser Leu Met Glu Ser Asp Gly His Asp Ile Pro Phe Glu Thr Phe Leu 945 950 955 960

Gly Phe Asn Gly Asp Lys Val Pro Asp Ile Asp Leu Asn Phe Ser Gly

Glu Tyr Gln Ala Lys Ala His Asn Tyr Thr Lys Val Leu Phe Gly Glu

Asp His Val Tyr Arg Ala Gly Thr Ile Thr Thr Ile Ala Asp Lys Thr 

Ala Phe Gly Phe Val Lys Gly Tyr Glu Arg Asp Lys Gln Ile Asn Tyr 

Arg Ser Ala Glu Val Asp Arg Leu Ser Asp Gly Leu Thr Gly Val Arg 

Arg Ser Thr Gly Gln His Pro Gly Gly Ile Ile Val Ile Pro Asp Asp 

Met Asp Val Phe Asp Phe Thr Pro Ile Gln Tyr Pro Ala Asp Asp Gln 

Thr Ala Glu Trp Gln Thr Thr His Phe Asp Phe His Ser Ile Asp Glu 

Asn Val Leu Lys Leu Asp Ile Leu Gly His Asp Asp Pro Thr Met Ile 

Arg Lys Leu Gln Asp Leu Ser Gly Phe Asp Pro Gln Glu Ile Pro Val 

Ser Asp Glu Asp Val Met Lys Ile Phe Ser Gly Pro Glu Val Leu Gly . 1130 

Val Thr Pro Glu Gln Ile Phe Ser Asn Thr Gly Thr Leu Gly Val Pro 

Glu Phe Gly Thr Gln Phe Val Arg Glu Met Leu Glu Gln Thr His Pro 

Ser Thr Phe Ala Glu Leu Leu Gln Ile Ser Gly Leu Ser His Gly Thr 

Asp Val Trp Leu Gly Asn Ala Glu Glu Leu Ile Arg Asn His Asn Ile 

Pro Leu Ser Glu Val Ile Gly Cys Arg Asp Asp Ile Met Val Tyr Leu 1205 1210 1215

- Gln His Gln Gly Leu Glu Asp Ser Leu Ala Phe Lys Ile Met Glu Phe 1220 1225 1230
- Val Arg Lys Gly Arg Gly Leu Gln Asp Asp Trp Ile Ala Thr Met Lys 1235 1240 1245
- Glu Asn Asp Val Pro Asp Trp Tyr Ile Glu Ser Cys Lys Lys Ile Lys 1250 1255 1260
- Tyr Met Phe Pro Lys Ala His Ala Ala Ala Tyr Val Leu Met Ala Leu 1265 1270 1275 1280
- Arg Val Ala Tyr Phe Lys Val His Tyr Pro Leu Tyr Tyr Tyr Ala Ala 1285 1290 1295
- Tyr Phe Ser Ile Arg Ala Ser Asp Phe Asp Leu Ile Ala Met Val Lys 1300 1305 1310
- Gly Lys Glu Gly Ile Lys Gly Ala Met Lys Glu Ile Arg Asp Lys Glu 1315 1320 1325
- Arg Glu Lys Thr Ala Thr Ala Lys Asp Lys Ala Leu Leu Thr Val Leu 1330 1335 1340
- Glu Val Ala Asn Glu Met Val Glu Arg Gly Phe Asp Phe Lys Met Val 1345 1350 1355 1360
- Asp Ile Asn Lys Ser Gln Ala Lys Asp Phe Val Ile Glu Asp Asn Gly 1365 1370 1375
- Leu Arg Ala Pro Phe Arg Ala Val Pro Ser Leu Gly Ser Ser Ala Ala 1380 1385 1390
- Gln Ala Val Ile Asp Ala Arg Glu Asp Ser Asp Phe Leu Ser Lys Glu 1395 1400 1405
- Asp Leu Ser Lys Arg Gly Lys Leu Ser Lys Thr Val Met Asp Tyr Leu 1410 1415 1420

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Asp Asn Asn His Val Leu Asp His Leu Pro Asp Glu Asn Gln Leu Ser 1425 1430 1435 1440

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aag tat gag ccc aat Lys Tyr Glu Pro Asn 30	ggt gga cca Gly Gly Pro 35	gca ggc ggc Ala Gly Ģly	gac ggt ggc Asp Gly Gly 40	agt ggc 147 Ser Gly									
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ttc cgc tac aac ccc Phe Arg Tyr Asn Pro 60	cat ttt aag His Phe Lys 65	gca gat agt Ala Asp Ser 70	ggc caa aat Gly Gln Asn	ggt atg 243 Gly Met 75									
ccc aag ggg atg aat Pro Lys Gly Met Asn 80	ggt aag aag Gly Lys Lys	gca gag gac Ala Glu Asp 85	ttg att atc Leu Ile Île	agt gtc 291 Ser Val 90									
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tcc att gcc gaa aac Ser Ile Ala Glu Asr	ggc gag ccg Gly Glu Pro	ggc caa gag Gly Gln Glu	cgg gat gtc Arg Asp Val	gaa ttg 483 Glu Leu									

140	145	. 150	155
gaa tta aaa gtc atg Glu Leu Lys Val Met 160			
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gga gcc tat cac ttt Gly Ala Tyr His Phe 190	act aca ctt go Thr Thr Leu Al 195	cc cct aat tta ggt q la Pro Asn Leu Gly v 200	gta gtg aat 627 Val Val Asn
gca gtg gac ggc aag Ala Val Asp Gly Lys 205	gaa ttt gtc tt Glu Phe Val Le 210	tg gcg gat att cct ( eu Ala Asp Ile Pro ( 215	ggc tta att 675 Gly Leu Ile
gaa ggg gct tca gaa Glu Gly Ala Ser Glu 220	ggg gtt ggt tt Gly Val Gly Le 225	tg ggg att gac ttc o eu Gly Ile Asp Phe i 230	ctc aag cat 723 Leu Lys His 235
	Ile Leu Leu Hi	at gta ctt gat atg : is Val Leu Asp Met : 245	
	Ile Asp Asp Ph	tt gac cag att aac he Asp Gln Ile Asn ( 60	
		ac cgc aag cag gtc sp Arg Lys Gln Val 280	
		gg gat aat tta atc rg Asp Asn Leu Ile 295	
gcc gag tta gac ago Ala Glu Leu Asp Ser 300	cgg gac ctt ga Arg Asp Leu As 305	ac tat gaa atc ttt sp Tyr Glu Ile Phe 310	gaa gtg tca 963 Glu Val Ser 315
gct gcc acc cag gct Ala Ala Thr Gln Ala 320	Gly Ile Gln As	ac cta gtc atc cga sp Leu Val Ile Arg 325	cta gcc gac 1011 Leu Ala Asp 330
tta gtc gac caa ctg Leu Val Asp Gln Leu 335	Asp Gln Ala Pi	ca agt tta gac cag ro Ser Leu Asp Gln 40	gaa gaa act 1059 Glu Glu Thr 345
		ac aag ttt caa gct Yr Lys Phe Gln Ala 360	
aaa ttt gac ctt gac Lys Phe Asp Leu Asp 365	cgc gac cct go Arg Asp Pro G 370	aa ggg gta tgg ttg lu Gly Val Trp Leu 375	gtt tct ggt 1155 Val Ser Gly

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gcc Ala	att Ile	atg Met	cgg Arg	ttt Phe 400	tct Ser	cgc Arg	cag Gln	cta Leu	aga Arg 405	Gly ggg	atg Met	gga Gly	gta Val	gac Asp 410	caa Gln	1251	
gcc Ala	tta Leu	aga Arg	gac Asp 415	aag Lys	Gly ggg	gct Ala	cag Gln	tct Ser 420	ggt Gly	gac Asp	ctc Leu	gtc Val	caa Gln 425	gtt Val	gaa Glu	1299	
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Gly	Gln	. Glu 115		Leu	Ala	Ala	Gln 120		Gly	' Arg	Gly	Gly 125		Gly	Asn		

Lys Arg Phe Ala Thr His Lys Asn Pro Ala Pro Ser Ile Ala Glu Asn

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135 140 130 Gly Glu Pro Gly Gln Glu Arg Asp Val Glu Leu Glu Leu Lys Val Met Ala Asp Val Gly Leu Val Gly Tyr Pro Ser Val Gly Lys Ser Thr Leu 170 Leu Ser Val Val Ser Gly Ala Lys Pro Lys Ile Gly Ala Tyr His Phe Thr Thr Leu Ala Pro Asn Leu Gly Val Val Asn Ala Val Asp Gly Lys 200 Glu Phe Val Leu Ala Asp Ile Pro Gly Leu Ile Glu Gly Ala Ser Glu 215 210 Gly Val Gly Leu Gly Ile Asp Phe Leu Lys His Ile Glu Arg Thr Arg 235 225 Ile Leu Leu His Val Leu Asp Met Ser Gly Met Glu Gly Arg His Pro 250 Ile Asp Asp Phe Asp Gln Ile Asn Gln Glu Leu Lys Asp Tyr Asn Glu 260 Lys Leu Leu Asp Arg Lys Gln Val Ile Val Ala Asn Lys Met Asp Leu 280 Pro Gln Ser Arg Asp Asn Leu Ile Glu Phe Lys Ala Glu Leu Asp Ser 300 295 Arg Asp Leu Asp Tyr Glu Ile Phe Glu Val Ser Ala Ala Thr Gln Ala 310 305 Gly Ile Gln Asp Leu Val Ile Arg Leu Ala Asp Leu Val Asp Gln Leu 325 Asp Gln Ala Pro Ser Leu Asp Gln Glu Glu Thr Ser Glu Ala Asp Gln 345 340 Arg Val Val Tyr Lys Phe Gln Ala Asp Gln Asp Lys Phe Asp Leu Asp 360 355

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Arg Asp Pro Glu Gly Val Trp Leu Val Ser Gly Pro Lys Val Glu Arg 370 . 375 Leu Tyr Ala Met Thr Asn Phe Asp His Glu Glu Ala Ile Met Arg Phe 395 390 Ser Arg Gln Leu Arg Gly Met Gly Val Asp Gln Ala Leu Arg Asp Lys 405 Gly Ala Gln Ser Gly Asp Leu Val Gln Val Glu Asp Phe Val Phe Glu 425 Phe Met Asp 435 <210> 73 <211> 1338 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (25)..(1338) <223> aagagaaaga aagaaggtgt actg atg gct aat cct tta gta gcc ata atc Met Ala Asn Pro Leu Val Ala Ile Ile ggc cgg cct aat gtc ggc aag tca act att ttc aac cgg att att gga 99 Gly Arg Pro Asn Val Gly Lys Ser Thr Ile Phe Asn Arg Ile Ile Gly gac cgc tta gcc att gtc cag gat gaa ccc ggg gtc acc cgg gac cgt 147 Asp Arg Leu Ala Ile Val Gln Asp Glu Pro Gly Val Thr Arg Asp Arg 35 195 att tat gcc gat gct gaa tgg ttg ggc aaa gac ttt tct gtt ata gat Ile Tyr Ala Asp Ala Glu Trp Leu Gly Lys Asp Phe Ser Val Ile Asp 50 243 acg gga gga atc act ttt gat gat ttg ccc ttg cat gaa gaa ata aaa Thr Gly Gly Ile Thr Phe Asp Asp Leu Pro Leu His Glu Glu Ile Lys 65 gtc caa gct gaa att gcc att gat gaa gca gat gtc atc gtc atg gta 291 Val Gln Ala Glu Ile Ala Ile Asp Glu Ala Asp Val Ile Val Met Val 75 80

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att Ile	ttg Leu	cag Gln	cag Gln	tcc Ser 110	aac Asn	aaa Lys	ccc Pro	gtg Val	gtc Val 115	ctt Leu	gct Ala	gtt Val	aat Asn	aaa Lys 120	aca Thr	387
gat Asp	aat Asn	cct Pro	gag Glu 125	ctt Leu	aga Arg	aat Asn	gaa Glu	ata Ile 130	tat Tyr	gag Glu	ttt Phe	tac Tyr	ggg Gly 135	tta Leu	ggc Gly	435
ttg Leu	ggt Gly	gac Asp 140	ccc Pro	ctt Leu	ccg Pro	gta Val	tcc Ser 145	Gly ggg	tct Ser	cac His	ggc Gly	cta Leu 150	ggc	ttt Phe	GJÀ āāā	483
gac Asp	ctc Leu 155	tta Leu	gac Asp	gca Ala	gtg Val	gtg Val 160	gcc Ala	aac Asn	ttt Phe	cct Pro	aat Asn 165	gag Glu	gcc Ala	aat Asn	atg Met	531
gct Ala 170	tat Tyr	gac Asp	caa Gln	gat Asp	acc Thr 175	att Ile	aag Lys	ttc Phe	tgc Cys	ttg Leu 180	att Ile	ggt Gly	cgt Arg	ecc Pro	aat Asn 185	579
gtt Val	ggc Gly	aag Lys	tct Ser	agc Ser 190	cta Leu	gtt Val	aat Asn	gct Ala	att Ile 195	att Ile	Gly ggg	gaa Glu	gac Asp	cgg Arg 200	gtt Val	627
ata Ile	gtc Val	tct Ser	gaa Glu 205	cta Leu	gaa Glu	Gly aaa	acc Thr	acc Thr 210	Arg	gat Asp	gca Ala	att Ile	gac Asp 215	act Thr	ccc Pro	675
ttt Phe	atg Met	acc Thr 220	Gln	gat Asp	Gly	cag Gln	gac Asp 225	tat Tyr	gtt Val	atg Met	atc	gat Asp 230	Thr	gct Ala	Gl <sup>à</sup> aaa	723
atc Ile	cgg Arg 235	Arg	cgg Arg	Gly	aag Lys	gtc Val 240	Tyr	gaa Glu	aaa Lys	act Thr	gaa Glu 245	Lys	tat Tyr	tct Ser	gtt Val	771
atg Met 250	Arg	gca	cag Gln	cga Arg	gct Ala 255	Ile	gac Asp	cgg Arg	tct Ser	gat Asp 260	gtg Val	gtc Val	ttg Leu	tgt Cys	gtc Val 265	819
ctg Leu	gat Asp	gct	gaa Glu	aca Thr 270	Gly	att Ile	aga Arg	gac Asp	caa Gln 275	Asp	aag Lys	aag Lys	gtt Val	Phe 280	GTA	867
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gac Asp	acg Thr	att Ile 300	Lys	aaa Lys	gag Glu	act Thr	aac Asr 305	Thr	atg Met	cga Arg	gac J Asp	ttt Phe 310	Glu	ttg Lev	caa Gln	963
att	cgc	gad	caa	a tto	cgo	tac	cto	cac	tat	gci	c cca	ato	ctt	ttc	gtc	1011

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										165	/235					
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cgg g Arg V	tc al	tat Tyr	tat Tyr	aac Asn 350	cgc Arg	aat Asn	caa Gln	cgg Arg	gtc Val 355	aag Lys	tcc Ser	tcc Ser	ctc Leu	tta Leu 360	aat Asn	1107
gat g Asp V	jtg Val	ctg Leu	agt Ser 365	gat Asp	gca Ala	cta Leu	gcc Ala	agc Ser 370	aat Asn	cct Pro	gca Ala	cct Pro	agt Ser 375	aag Lys	tca Ser	1155
ggg a	ag Lys	cga Arg 380	ctc Leu	aag Lys	gtc Val	ttt Phe	tat Tyr 385	gcg Ala	acc Thr	cag Gln	gta Val	gcc Ala 390	act Thr	aat Asn	cca Pro	1203
cct a Pro I	act Thr 395	ttt Phe	gtg Val	gtt Val	ttt Phe	gtc Val 400	aat Asn	gat Asp	cct Pro	gac Asp	ctc Leu 405	atg Met	cac His	ttc Phe	tcc Ser	1251
tat g Tyr 0 410	gag Slu	cgc Arg	ttt Phe	tta Leu	gaa Glu 415	aat Asn	cga Arg	ttc Phe	cgc Arg	gaa Glu 420	agc Ser	ttt Phe	gac Asp	ttc Phe	tat Tyr 425	1299
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Ser'	Thr	Ile	Phe 20	. Asr	Arg	Ile	: Ile	Gly 25	Asp	Arg	Leu	a Ala	11e 30	e Val	Gln	
Asp	Glu	Pro 35	Gly	va]	. Thr	Arg	Asp 40	Arg	; Ile	• Туг	Ala	Asp 45	Ala	ı Glı	ı Trp	
	Gly 50	Lys	Asp	Phe	e Ser	val 55	. Ile	e As <u>ı</u>	Thi	Gly	/ Gl <sub>3</sub> 60	/ Ile	e Thi	r Phe	e Asp	
Asp 65	Leu	Pro	) Lev	ı His	5 Glu 70	ı Glı	ı Ile	e Lys	s Val	L Glr 75	n Ala	a Gli	ı Ile	e Ala	a Ile 80	

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Asp Glu Ala Asp Val Ile Val Met Val Thr Ser Val Lys Glu Gly Ile 85 90 95

Thr Asp Leu Asp Asp Gln Val Ala Leu Ile Leu Gln Gln Ser Asn Lys
100 105 110

Pro Val Val Leu Ala Val Asn Lys Thr Asp Asn Pro Glu Leu Arg Asn 115 120 125

Glu Ile Tyr Glu Phe Tyr Gly Leu Gly Leu Gly Asp Pro Leu Pro Val 130 135 140

Ser Gly Ser His Gly Leu Gly Phe Gly Asp Leu Leu Asp Ala Val 145 150 155 160

Ala Asn Phe Pro Asn Glu Ala Asn Met Ala Tyr Asp Gln Asp Thr Ile 165 170 175

Lys Phe Cys Leu Ile Gly Arg Pro Asn Val Gly Lys Ser Ser Leu Val 180 185 190

Asn Ala Ile Ile Gly Glu Asp Arg Val Ile Val Ser Glu Leu Glu Gly 195 200 205

Thr Thr Arg Asp Ala Ile Asp Thr Pro Phe Met Thr Gln Asp Gly Gln 210 215 220

Asp Tyr Val Met Ile Asp Thr Ala Gly Ile Arg Arg Arg Gly Lys Val 225 230 235

Tyr Glu Lys Thr Glu Lys Tyr Ser Val Met Arg Ala Gln Arg Ala Ile 245 250 255

Asp Arg Ser Asp Val Val Leu Cys Val Leu Asp Ala Glu Thr Gly Ile 260 265 270

Arg Asp Gln Asp Lys Lys Val Phe Gly Tyr Ala His Gln Ala Gly Lys 275 280 285

Gly Ile Ile Ile Leu Val Asn Lys Trp Asp Thr Ile Lys Lys Glu Thr 290 295 300

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Asn Thr Met Arg Asp Phe Glu Leu Gln Ile Arg Asp Gln Phe Arg Tyr 305 310 315 320	
Leu His Tyr Ala Pro Ile Leu Phe Val Ser Ala Lys Thr Lys Gln Arg 325 330 335	
Leu Glu Val Ile Pro Glu Leu Val Asp Arg Val Tyr Tyr Asn Arg Asn 340 345 350	
Gln Arg Val Lys Ser Ser Leu Leu Asn Asp Val Leu Ser Asp Ala Leu 355 360 365	
Ala Ser Asn Pro Ala Pro Ser Lys Ser Gly Lys Arg Leu Lys Val Phe 370 375 380	
Tyr Ala Thr Gln Val Ala Thr Asn Pro Pro Thr Phe Val Val Phe Val 385 390 395 400	
Asn Asp Pro Asp Leu Met His Phe Ser Tyr Glu Arg Phe Leu Glu Asn 405 410 415	
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atc caa ttg ccc ctc ctg atg gac cgc ctg aag gag ctt ggc atg gag Ile Gln Leu Pro Leu Leu Met Asp Arg Leu Lys Glu Leu Gly Met Glu	147

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	•																
				35					40					45			
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gct ga Ala As 80	sp 1	cta Leu	gac Asp	gaa Glu	gga Gly	ata Ile 85	acc Thr	gtc Val	acc Thr	ctc Leu	ctg Leu 90	gct Ala	aaa Lys	aac Asn	aag Lys	291	
gct gg Ala G 95	gc ly '	tac Tyr	cag Gln	gct Ala	ctc Leu 100	tta Leu	gcc Ala	tta Leu	tcg Ser	act Thr 105	gac Asp	ctt Leu	caa Gln	gtt Val	aac Asn 110	339	
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tat ac	ca hr	ata Ile	ttc Phe 130	cca Pro	agc Ser	tct Ser	gac Asp	cca Pro 135	aaa Lys	gtg Val	aaa Lys	gca Ala	gac Asp 140	ctc Leu	tta Leu	435	
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acc Thr 415	Phe	gcg Ala	gct Ala	aag Lys	tcc Ser 420	tcc Ser	atc Ile	agg Arg	gaa Glu	att Ile 425	Met	cgg Arg	acc Thr	ttg Leu	ggt Gly 430	1299
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tca Ser	gaa Glu	. atg . Met	gac Asp 690	Gln	gga Gly	cgg	gaa Glu	aaa Lys 695	Phe	ata Ile	aga Arg	gga Gly	gcc Ala 700	ttg Leu	gac Asp	2115
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tac Tyr	aag Lys	tat Tyr	gcc Ala 770	cat His	gaa Glu	gtc Val	cgg Arg	gct Ala 775	aga Arg	aaa Lys	att Ile	aaa Lys	cta Leu 780	cta Leu	aaa Lys	2355
cca Pro	gac Asp	atc Ile 785	aac Asn	caa Gln	agc Ser	ctt Leu	gga Gly 790	tct Ser	ttt Phe	acg Thr	gtt Val	cgg Arg 795	cag Gln	aat Asn	Gly ggc	2403
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gaa Gli 895	ı Ası	c cto o Lev	g gaa 1 Glu	a gaç ı Glı	ı Phe	ago Ser	Pro	Lys	Asp	cto Lev 905	ı Ile	caa Glr	tat Tyr	gaa Glu	gaa Glu 910	2739
ga: Gl:	a tta u Lei	a aco	c ggt r Gl	ttty Pho	e Ty	tto Phe	tco Sei	ago Sei	cac His	Pro	ttg Lei	g ago ı Sei	c cgg	tat Typ 925	gac Asp	2787
tc Se:	c cto	g cga u Ara	a caq g Gli 93	n As	c tta p Le	ı Lys	a acç s Thi	tco Sei 935	r Phe	ata e Ile	a gci	t gat a Ası	t tta Dei 940	ı Glı	gag ı Glu	2835
gg Gl:	c caa	a tc n Se	t tg	c ca s Gl:	a gt n Va	t tta l Le	a ggt	t cag	g cto n.Lev	g gt ı Va	t car l Gl:	a gto n Vai	c cgg	g aaa	a act 5 Thr	2883

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Thr Gly Gln Ile Ser	ctg gtg gtc ttt ccg aat gta tac cgc gaa tgc Leu Val Val Phe Pro Asn Val Tyr Arg Glu Cys 980 985 990	2979
cta cct tac ctc aaa Leu Pro Tyr Leu Lys 995	gaa gga gtg gtc ctg gtc gtc tca ggc aag gta Glu Gly Val Val Leu Val Val Ser Gly Lys Val 1000 1005	3027
gaa gtt agg aag gga Glu Val Arg Lys Gly 1010	gaa atc cag cta aaa gtc cag acc atg aaa gag Glu Ile Gln Leu Lys Val Gln Thr Met Lys Glu 1015 1020	3075
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gcc cga cat ccc ggc Ala Arg His Pro Gly 1055	cag aag cga gtg att gtt tac gac cag gcc agc Gln Lys Arg Val Ile Val Tyr Asp Gln Ala Ser 1060 1065 1070	3219
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gat acc cta aac cag Asp Thr Leu Asn Gln 1090	ctc cag gac ctc cta ggc cag gat tct tgt atc Leu Gln Asp Leu Leu Gly Gln Asp Ser Cys Ile 1095 1100	3315
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Leu Pro Leu Leu Met Asp Arg Leu Lys Glu Leu Gly Met Glu Ala Val

45 35 40 Ala Leu Thr Asp His Asn Val Met His Gly Ala Val Glu Phe Tyr Gln Glu Ala Lys Lys His Gly Ile Lys Pro Ile Met Gly Leu Arg Ala Asp Leu Asp Glu Gly Ile Thr Val Thr Leu Leu Ala Lys Asn Lys Ala Gly 90 Tyr Gln Ala Leu Leu Ala Leu Ser Thr Asp Leu Gln Val Asn Lys Gln Ala Ile Thr Leu Asp Gln Val Arg Ser Val Ala Gln Asp Leu Tyr Thr 120 115 Ile Phe Pro Ser Ser Asp Pro Lys Val Lys Ala Asp Leu Leu Asp Lys 135 Gln Ala Ser Asn Leu Thr Ala Met Thr Gln Asn Leu Pro His Ser Tyr 1.55 Leu Gly Leu Val Pro Asp Gln Asp Gln Lys Ile Tyr Gln Leu Ala Arg Thr Leu Ser Asp Ser Gly Gly Leu Lys Val Leu Ala Leu Ser Asp Val Arg Cys Leu Glu Glu Ser Gln Val Ser Thr Leu Glu Ile Leu Ser His 200 195 Ile Lys Ala Asn Gln Lys Ile Gln Phe Asp Thr Gln Ala Arg Glu Asn 220 210 Tyr Ala Leu Arg Ser Pro Gln Glu Met Glu Ser Phe Phe Asn Gln Val Gly Leu Gly Gln Ala Leu Lys Asn Thr Lys Asp Val Ala Gln Ser Val 245 Asp Trp Ser Leu Asp Leu Gly Gln Ala Lys Leu Pro Ala Phe Asp Leu 265

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Pro Glu Gly Glu Thr Lys Asp Ser Tyr Leu Gly Lys Leu Ala Gln Lys 280 275

Gly Leu Gln Glu Arg Val Pro Gly Tyr Gly Gln Asp Tyr Gln Asp Arg 295 290

Leu Asp Lys Glu Leu Ala Val Ile Ser Ser Met Gly Phe Ser Asp Tyr 315 305

Phe Leu Ile Val Trp Asp Leu Met Gln Phe Ala Arg Gln Glu Lys Ile 330

Glu Thr Gly Phe Gly Arg Gly Ser Ala Ala Ala Ser Leu Val Ser Tyr 345 340

Ala Leu Tyr Ile Thr Gly Val Asp Pro Ile His Tyr Asp Leu Leu Phe 360

Glu Arg Phe Leu Asn Lys Asp Arg Phe Thr Met Pro Asp Ile Asp Leu 375

Asp Phe Pro Asp Asn Lys Arg Gln Val Ile Leu Asp Tyr Val Tyr Arg 395 390 385

Lys Tyr Gly Pro Asp His Val Ala Gln Ile Leu Thr Phe Gly Thr Phe 405 410

Ala Ala Lys Ser Ser Ile Arg Glu Ile Met Arg Thr Leu Gly Tyr Lys

Asn Glu Asp Met Lys Thr Trp Ser Gln Ala Ile Pro Asp Thr Val Asn 440

Ile Ser Leu Ser Lys Ala Tyr Asp Glu Ser Lys Asp Leu Gln Lys Leu 455

Val Gln Gln Ser His Glu Asn Glu Arg Ile Phe Ala Met Ala Gln Asp 470 475

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Gly	Lys	Val 515	Pro	Asn	Thr	Gln	Phe 520	Thr	Met	Glu	Asp	Val 525	Glu	Ala	Val
Gly	Leu 530	Leu	Lys	Met	Asp	Phe 535	Leu	Ser	Leu	Lys	Asn 540	Leu	Thr	Ile	Leu
Ala 545	Asp	Cys	Leu	Asn	Phe 550	Ser	Gln	Tyr	Glu	Gly 555	Gln	Gly	Gly	Gly	Ile 560
Ser	Lys	Gln	Asp	11e 565	Pro	Ile	Asp	Asp	Pro 570	Lys	Thr	Leu	Asp	Leu 575	Phe
Ala	Arg	Gly	Asp 580	Thr	Asn	Gly	Val	Phe 585	Gln	Phe	Glu	Lys	Glu 590	Gly	Ile
Lys	Lys	Val 595		Arg	Gln	Leu	Gln 600	Pro	Thr	Ser	Phe	Glu 605	Asp	Ile	Val
Ala	Thr 610		Ala	Leu	Tyr	Arg 615	Pro	Gly	Pro	Met	Gly 620	Gln	Ile	Glu	Asn
Tyr 625	Ile	Asn	Arg	Lys	His 630		Gln	Glu	Lys	Ile 635	Ile	Tyr	Pro	His	Glu 640
Asp	Leu	Lys	: Asp	Ile 645		Glu	. Val	Thr	Tyr 650	Gly	·Ile	Ile	· Val	Tyr 655	Gln
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700

695

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- Ala Asn Tyr Gly Phe Asn Arg Ala His Ala Val Ala Tyr Ser Met Leu 725 730 735
- Ala Tyr His Met Ala Tyr Phe Lys Val His Gln Pro Lys Ser Phe Phe 740 745 750
- Ala Ala Val Met Lys Ala Asp Trp Gly Asn Lys Ala Lys Ile Tyr Lys 755 760 765
- Tyr Ala His Glu Val Arg Ala Arg Lys Ile Lys Leu Leu Lys Pro Asp 770 775 780
- Ile Asn Gln Ser Leu Gly Ser Phe Thr Val Arg Gln Asn Gly Ile Gln 785 790 795 800
- Val Gly Leu Lys Met Val Lys Gly Val Ala Ser Pro Phe Val Asn His 805 810 815
- Ile Leu Glu Ile Arg Lys Glu Lys Gly Ala Phe Thr Ser Leu Arg Asp 820 825 830
- Phe Cys Glu Lys Ile Asp Ser Gln Phe Leu Ser Gln Asp Pro Ile Glu 835 840 845
- Ala Leu Ile Leu Val Gly Ala Phe Asp Gln Met Gly Pro Asn Arg Arg 850 855 860
- Thr Met Leu Ala Gly Leu Glu Ala Thr Ile Glu Phe Val Ala Lys Ser 865 870 875 880
- Ser Gly Asn Ile Thr Leu Phe Asp Thr Leu Lys Pro Arg Gln Glu Asp 885 890 895
- Leu Glu Glu Phe Ser Pro Lys Asp Leu Ile Gln Tyr Glu Glu Glu Leu 900 905 910
- Thr Gly Phe Tyr Phe Ser Ser His Pro Leu Ser Arg Tyr Asp Ser Leu 915 920 925
- Arg Gln Asp Leu Lys Thr Ser Phe Ile Ala Asp Leu Glu Glu Gly Gln 930 935 940
- Ser Cys Gln Val Leu Gly Gln Leu Val Gln Val Arg Lys Thr Gln Thr

945 950 955 960

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Tyr Leu Lys Glu Gly Val Val Leu Val Val Ser Gly Lys Val Glu Val 995 1000 1005

Arg Lys Gly Glu Ile Gln Leu Lys Val Gln Thr Met Lys Glu Ala Ser 1010 1015 1020

Gln Val Gln Lys Glu Thr Lys Gln Leu Tyr Leu Lys Phe Ala Asp Leu 1025 1030 1035 1040

Asn Gln Asp Lys Glu Ser Phe Arg Gln Val Gln Lys Ile Leu Ala Arg 1045 1050 1055

His Pro Gly Gln Lys Arg Val Ile Val Tyr Asp Gln Ala Ser Gln Gln 1060 1065 1070

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act Thr	ttt Phe	gaa Glu 65	gaa Glu	ggt Gly	cag Gln	ccc Pro	gaa Glu 70	tcg Ser	gtc Val	agt Ser	cac His	att Ile 75	gat Asp	ttg Leu	acg Thr	240
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Gly 95	Lys	ctg Leu	Ala	Asn	Gly 100	Ile	Gly	Asp	Asp	Phe 105	Val	Ser	Thr	Ala	Leu 110	336
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Met	Tyr	gaa Glu	Asn 130	Pro	Ala	Leu	Lys	Lys 135	Asn	Lys	Ala	Phe	Leu 140	Ile	Glu	432
Gln	Gly	cat His 145	Tyr	Trp	Met	Glu	Pro 150	Asp	Ile	Gly	Phe	Leu 155	Ala	Glu	Gly	480
Tyr	Glu 160		Leu	Gly	Arg	Phe 165	Pro	Asp	Leu	qzA	Arg 170	Ile	Met	Ala	Glu	528
Phe 175	Asn	cat His	Phe	Ile	11e 180	Ala	Arg	Asn	Pro	Gly 185	Ile	Leu	Ser	Gly	Lys 190	576
Lys	Val	ctc Leu	Val	Thr 195	Ala	Gly	Gly	Thr	Val 200	Glu	Arg	Ile	Asp	Pro 205	Val	624
cgg Arg	tat Tyr	att	tcc Ser 210	aat Asn	gat Asp	tct Ser	tct Ser	ggt Gly 215	Lys	atg Met	ggc	cac His	caa Gln 220	ctt Leu	gct Ala	672

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gac ttg ccg acc agt ccc ttt att gac cgc ttt cag gtg gag tcc acc

720

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ctc Leu	atg Met	atg Met	gcc Ala	gca Ala 275	gcg Ala	gtg Val	tct Ser	gac Asp	tac Tyr 280	cgg Arg	cca Pro	gtc Val	aac Asn	cgg Arg 285	tca Ser	864
gac Asp	aaa Lys	aag Lys	atg Met 290	aaa Lys	aag Lys	caa Gln	gat Asp	aat Asn 295	tta Leu	acc Thr	att Ile	gaa Glu	ctg Leu 300	gaa Glu	aaa Lys	912
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aat Asn	gtc Val 320	Gly	ttt Phe	gca Ala	gca Ala	gaa Glu 325	acc Thr	cat His	aac Asn	ctt Leu	gaa Glu 330	gaa Glu	tat Tyr	gcc Ala	caa Gln	1008
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ggc Gly	cġg Arg	gga Gly	gac Asp	cgg Arg 355	Gly	ttt Phe	aat Asn	gcg Ala	gat Asp 360	Glu	aat Asn	gcg Ala	gcc Ala	ctt Leu 365	gtt Val	1104
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Va!	l Tyr	r Ly:	s Sei 20	r Le	а Туг	r Lei	ı Leı	1 Arg 25	g Glu	ı Ile	e Ile	e Ly:	s Glr 30	ı Gly	, Gly	

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Glu Val Arg Val Ala Met Thr Gln Ala Ala Cys Gln Phe Val Asn Pro 40 Leu Ser Phe Gln Val Leu Ser Gln Lys Lys Val Gln Ile Asp Thr Phe Glu Glu Gly Gln Pro Glu Ser Val Ser His Ile Asp Leu Thr Asp Trp Ala Asp Tyr Ser Ile Val Ala Pro Ala Thr Ala Asn Ile Ile Gly Lys 90 85 Leu Ala Asn Gly Ile Gly Asp Asp Phe Val Ser Thr Ala Leu Leu Ala Thr Asp His Pro Ile Phe Leu Val Pro Ala Met Asn Thr Lys Met Tyr 115 120 Glu Asn Pro Ala Leu Lys Lys Asn Lys Ala Phe Leu Ile Glu Gln Gly 135 130 His Tyr Trp Met Glu Pro Asp Ile Gly Phe Leu Ala Glu Gly Tyr Glu 1.55 Gly Leu Gly Arg Phe Pro Asp Leu Asp Arg Ile Met Ala Glu Phe Asn His Phe Ile Ile Ala Arg Asn Pro Gly Ile Leu Ser Gly Lys Lys Val Leu Val Thr Ala Gly Gly Thr Val Glu Arg Ile Asp Pro Val Arg Tyr 200 195 Ile Ser Asn Asp Ser Ser Gly Lys Met Gly His Gln Leu Ala Gln Ala 210 Ala Tyr Glu Ala Gly Ala Gln Val Ser Leu Val Thr Ala Ser Asp Leu

Pro Thr Ser Pro Phe Ile Asp Arg Phe Gln Val Glu Ser Thr Leu Asp

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181/235 Leu Tyr Gln Thr Val Ser Asp Leu Tyr Asp His His Asp Ile Leu Met 265 Met Ala Ala Ala Val Ser Asp Tyr Arg Pro Val Asn Arg Ser Asp Lys 275 280 Lys Met Lys Lys Gln Asp Asn Leu Thr Ile Glu Leu Glu Lys Asn Pro 295 Asp Ile Leu Ala Glu Met Gly Arg Arg Lys Asp Gln Gln Ile Asn Val 315 310 305 Gly Phe Ala Ala Glu Thr His Asn Leu Glu Glu Tyr Ala Gln Lys Lys 325 Leu Ala Ser Lys Gln Ala Asp Leu Ile Val Ala Asn Glu Val Gly Arg 345 Gly Asp Arg Gly Phe Asn Ala Asp Glu Asn Ala Ala Leu Val Phe Ser 360 Ser Asp Gln Asp Pro Leu Glu Leu Pro Leu Gln Ser Lys Lys Asp Met 380 375 Ala Lys Lys Ile Ile Glu Val Val Ala Ser Lys Leu Pro Ala Ser Pro 395 390 385 Lys <210> 79 <211> 1053 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (22)..(1053) <223> <400> 79 aagaagaagg gaggaagact g atg aaa att gaa gac caa ctc aaa aaa att 51 Met Lys Ile Glu Asp Gln Leu Lys Lys Ile

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15

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tac Tyr	ttg Leu	tta Leu	gac Asp 30	cag Gln	gtt Val	aaa Lys	aaa Lys	agt Ser 35	ttg Leu	agc Ser	cag Gln	gcc Ala	ctt Leu 40	ttg Leu	gac Asp	147	
cag Gln	gat Asp	gaa Glu 45	gct Ala	tct Ser	atg Met	aat Asn	ttt Phe 50	ggt Gly	caa Gln	ttt Phe	aat Asn	atg Met 55	atg Met	gct Ala	gat Asp	195	
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aag Lys	cgg Arg	aaa Lys	aca Thr	gat Asp 95	ctg Leu	gac Asp	cat His	gac Asp	ttg Leu 100	gat Asp	cgc Arg	ttg Leu	ctg Leu	gct Ala 105	tac Tyr	339	
ctc Leu	caa Gln	aac Asn	cca Pro 110	gcc Ala	gac Asp	ttt Phe	act Thr	gtt Val 115	ctc Leu	gtc Val	ttc Phe	ttt Phe	gcc Ala 120	PIO	tat Tyr	387	
gag Glu	aaa Lys	ctg Leu 125	Asp	aag Lys	cgg	aag Lys	aag Lys 130	gtc Val	acc Thr	aaa Lys	gcc Ala	cta Leu 135	Leu	cag Gln	gaa Glu	435	
gct Ala	gag Glu 140	Ile	ata :Ile	gat Asp	gcc Ala	agt Ser 145	Ser	cca Pro	gac Asp	caa Gln	aga Arg 150	ASP	cta Lev	aaa Lys	gat Asp	483	
atg Met 155	Val	cag Glr	aaa Lys	aaa Lys	gta Val	. Lys	gct Ala	cga Arg	ggc Gly	tac Tyr 165	GLI	ttt Phe	gac Asi	aaa Lys	gga Gly 170	. 531	
gct Ala	tta Lev	aag Lys	g gcc s Ala	ctg Leu 175	ı Val	gaa Glu	aaa Lys	acc Thr	aat Asr 180	ı Ala	a ac	tta Lei	agt 1 Sex	c cgg c Arg 185	gtc Val	579	
atg Met	caa Glr	ı gaç ı Glı	ttq 1 Lei 190	ı Ası	aag b Lys	g tta s Lev	tto Phe	ttq Lev 195	ı Tyı	c cat	tta s Le	a gat 1 Asi	gad Asj 20	5 гу	atc Ile	627	
ato Ile	acc Thi	gte Vai	l Gl	g tca n Sei	a gti r Val	gad L Asp	c cag Glr 210	ı Va.	gta L Val	a tca l Se:	a cca r Pro	a age o Se: 21:	r ne	g gaa u Gl	a agt ı Ser	675	
aat Asr	gto n Val 220	l Ph	t ag e Se:	t ati	t aad e Asi	gad n Ası 225	y Ty	ati	t tta e Le	a ago u Se:	c gg r Gl 23	A GT	a ag n Se	c cag r Gli	g gct n Ala	723	
gct Ala 23	a Il	a cg e Ar	g gc	c tt a Ph	t aa e As 24	n Asj	c tta p Le	a at	t ca e Gl:	a ca n Gl 24	n Ly	g ga s Gl	a ga u Gl	g cc u Pr	a att o Ile 250	•	

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Lys Ile Ile Ala I	tt atg atg aac le Met Met Asn 55	caa ttc cgt tta tta ttg cag gtt Gln Phe Arg Leu Leu Leu Gln Val 260 265	819
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tca gat cat ttg a Ser Asp His Leu I 315	att aaa acg ggc le Lys Thr Gly 320	aag gtg acc tcg caa ttg caa ttt Lys Val Thr Ser Gln Leu Gln Phe 325 330	1011
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Asn Phe Gly Gln I 50	Phe Asn Met Met 55	Ala Asp Ser Leu Asp Met Ala Leu 60	ı
Ser Asp Ala Glu S 65	Ser Tyr Pro Phe 70	Phe Gly Asp Lys Arg Leu Val Tyo 75 80	c
	Phe Phe Leu Thr 85	Gly Glu Lys Arg Lys Thr Asp Let 90 95	1
Asp His Asp Leu 1	Asp Arg Leu Leu	Ala Tyr Leu Gln Asn Pro Ala As 105 110	·

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Phe	Thr	Val 115	Leu	Val	Phe	Phe	Ala 120	Pro	Tyr	Glu	Lys	Leu 125	Asp	Lys	Arg
Lys	Lys 130	Val	Thr	Lys	Ala	Leu 135	Leu	Gln	Glu	Ala	Glu 140	Ile	Ile	Asp	Ala
Ser 145	Ser	Pro	Aṣp	Gln	Arg 150	Asp	Leu	Lys	Asp	Met 155	Val	Gln	Lys	Lys	Val 160
ŗÀz	Ala	Arg	Gly	Tyr 165	Gln	Phe	Asp	Lys	Gly 170	Ala	Leu	Lys	Ala	Leu 175	Val
Glu	Lys	Thr	Asn 180		Asn	Leu	Ser	Arg 185	Val	Met	Gln	Glu	Leu 190	Asp	Lys
Leu	Phe	Leu 195		His	Leu	Asp	Asp 200	Lys	Ile	Ile	Thr	Val 205	Gln	Ser	Val
Asp	Gln 210		Val	Ser	Pro	Ser 215	Leu	Glu	Ser	Asn	Val 220	Phe	Ser	Ile	Asn
Asp 225		·Ile	. Leu	Ser	Gly 230		Ser	Gln	Ala	Ala 235	Ile	Arg	Ala	Phe	Asn 240
Asp	Leu	ı Ile	e Glr	Gln 245		Glu	Glu	Pro	1le 250	Lys	Ile	Ile	Ala	Ile 255	Met
Met	. Ası	ı Glr	n Phe 260		, Leu	. Leu	. Leu	Glr 265	Val	. Lys	lle	Leu	Arg 270	Thr	Lys
Gly	туз	Glr 275		ı Gly	r Glu	Ile	280		: Ile	e Lev	Lys	Val 285	His	Pro	Tyr
Arç	y Va. 290		s Lei	ı Ala	a Ile	e Glu 295	ı Lys	Glr	ı Glu	ı Ile	Phe 300	e Ser	· Lys	Gln	Ser
Let 305		r Thi	r Ala	а Туз	arg 310		c Lev	ı Ile	∋ Glv	315	Asp	His	: Lev	ılle	Lys 320
Thi	c Gl	y Ly	s Va	1 Th:		Glı	ı Lev	ı Glı	n Phe 330	e Glu	ı Lev	ı Phe	e Ala	335	ı Gln

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Phe Lys Asp Ser Val Met Asn 340

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cac His	ctg Leu	gat Asp	atc Ile 20	att Ile	aaa Lys	agg Arg	gcc Ala	agc Ser 25	cac His	tta Leu	ttc Phe	gat Asp	gaa Glu 30	gtc Val	atc Ile	96
gtt Val	gca Ala	gtt Val 35	gct Ala	aat Asn	aat Asn	aca Thr	tcg Ser 40	aaa Lys	aat Asn	agt Ser	atg Met	ttg Leu 45	aac Asn	ttt Phe	gac Asp	144
caa Gln	aaa Lys 50	ttg Leu	aac Asn	ctg Leu	gtt Val	gaa Glu 55	caa Gln	tca Ser	att Ile	gct Ala	agc Ser 60	cag Gln	ggt Gly	cta Leu	gct Ala	192
aat Asn 65	gtt Val	caa Gln	gcc Ala	aag Lys	aca Thr 70	tta Leu	gag Glu	tca Ser	ggc Gly	ttg Leu 75	att Ile	gtt Val	gac Asp	ttt Phe	gct Ala 80	240
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gac Asp	ttt Phe	gaa Glu	tat Tyr 100	Glu	att Ile	gcc Ala	att Ile	gag Glu 105	Asp	tta Leu	aat Asn	aag Lys	gtc Val 110	. 6111	gac Asp	336
cca Pro	gct Ala	att Ile 115	Glu	aca Thr	gtt Val	tac Tyr	cta Leu 120	Val	tcg Ser	tct Ser	tcc Ser	aaa Lys 125	тұт	cgg Arg	tcc Ser	384
att Ile	tct Ser	Ser	tct Ser	att	gtt Val	cgg Arg 135	Glu	att Ile	att	aaç Lys	ttt Phe 140	ASI	ggc Gly	c cgg / Arg	g ctt g Leu	432
gat Asp 145	Asp	cta Le	a gta ı Val	a cct L Pro	gac Asp 150	ccc Pro	gto Val	gto Val	gaa Glu	tat 1 Tyr 155	Pne	aaa Lys	a aaa s Lys	a taa	<b>a</b>	477

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Gln Lys Leu Asn Leu Val Glu Gln Ser Ile Ala Ser Gln Gly Leu Ala 50

Asn Val Gln Ala Lys Thr Leu Glu Ser Gly Leu Ile Val Asp Phe Ala

Lys Asp Gln Gly Ala Ser Ser Leu Val Arg Gly Leu Arg Ser Val Lys 90 95

Asp Phe Glu Tyr Glu Ile Ala Ile Glu Asp Leu Asn Lys Val Gln Asp 105 100

Pro Ala Ile Glu Thr Val Tyr Leu Val Ser Ser Lys Tyr Arg Ser 125 115

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2	aat Asn	gcc Ala	gtt Val	aac Asn	cgg Arg 30	atg Met	att Ile	gaa Glu	gat Asp	gga Gly 35	gtc Val	gaa Glu	ggc Gly	gtt Val	gaa Glu 40	ttt Phe	150
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	gcc Ala	aaa Lys	gaa Glu	caa Gln 125	Gly	gct Ala	ttg Leu	act Thr	gta Val 130	Gly	gtt Val	att Ile	acc Thr	cgg Arg 135	ccg Pro	ttc Phe	438
	act Thr	ttt Phe	gaa Glu 140	. Gly	cca Pro	aaa Lys	cgt Arg	ggg Gly 145	Arg	ttt Phe	gca Ala	gcc Ala	gaa Glu 150	GJÀ āāā	att Ile	gcc Ala	486
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acc gga aac caa gac tta acc ctc ttt gaa gct caa gat gct tct gat Thr Gly Asn Gln Asp Leu Thr Leu Phe Glu Ala Gln Asp Ala Ser Asp 270 275 280	870
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tcc atc aat gaa gac ctg gaa gat gag gtc atc gtt acc gtt att gca Ser Ile Asn Glu Asp Leu Glu Asp Glu Val Ile Val Thr Val Ile Ala 300 305 310	966
act ggt atc act ggt aaa gac atg ggc gag aaa tct tct aaa tcc tca Thr Gly Ile Thr Gly Lys Asp Met Gly Glu Lys Ser Ser Lys Ser Ser 315 320 325	1014
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Gln Ala Leu Asp Ala Asn Arg Ala Glu Thr Lys Ile Gln Leu Gly Glu
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Lys Leu Thr Arg Gly Leu Gly Ala Gly Ala Asn Pro Glu Val Gly Arg 65 70 75 80

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Ala Asp Met Val Phe Val Thr Ala Gly Met Gly Gly Gly Thr Gly Thr 100 105 110

Gly Gly Ala Gly Ile Ile Ala Arg Ile Ala Lys Glu Gln Gly Ala Leu 115 120 125

Thr Val Gly Val Ile Thr Arg Pro Phe Thr Phe Glu Gly Pro Lys Arg 130 135 140

Gly Arg Phe Ala Ala Glu Gly Ile Ala Gln Met Arg Glu His Val Asp 145 150 155 160

Thr Leu Val Thr Ile Ser Asn Asn Arg Leu Leu Glu Ile Val Asp Lys 165 170 175

Lys Thr Pro Met Met Glu Ala Phe Arg Glu Ala Asp Asn Val Leu Arg 180 185 190

Gln Gly Val Gln Gly Ile Ser Asp Leu Ile Thr Asn Pro Gly Tyr Val 195 200 205

Asn Leu Asp Phe Ala Asp Val Lys Thr Val Met Ala Asn Gln Gly Ser 210 215 220

Ala Leu Met Gly Ile Gly Ser Ala Ser Gly Glu Asn Arg Thr Ala Glu 225 230 235 240

Ala Thr Lys Lys Ala Ile Ser Ser Pro Leu Leu Glu Val Ser Leu Asn 245 250 255

Gly Ala Glu Asn Val Leu Leu Asn Ile Thr Gly Asn Gln Asp Leu Thr 260 265 270

Leu Phe Glu Ala Gln Asp Ala Ser Asp Ile Val Gly Ala Ala Ala Ser 275 280 285

Gly Asp Val Asn Ile Ile Phe Gly Thr Ser Ile Asn Glu Asp Leu Glu 290 295 300

Asp Glu Val Ile Val Thr Val Ile Ala Thr Gly Ile Thr Gly Lys Asp 305 310 315

Met Gly Glu Lys Ser Ser Lys Ser Ser Asn Arg Ser Gln Gly Pro Ser 325 330 335

Gln Lys Ser Gln Ala Arg Ser Ala Ser Glu Ser Ser Phe Ser Ser Trp 340 345 350

Gln Asn Gln Ser Asn Glu Arg Pro Gly Glu Asp Gln Asp Arg Pro Ser 355 360 365

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caa acg acc cta tca g Gln Thr Thr Leu Ser 2 225	Ala Ile His A	at gag caa gtg aag tat gcc aat sp Glu Gln Val Lys Tyr Ala Asn 30 235.	723
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atc aac acc tca cag Ile Asn Thr Ser Gln 255	caa aat gca g Gln Asn Ala G 260	gaa aag ctc aaa aga gaa gtt ggg Slu Lys Leu Lys Arg Glu Val Gly 265	819
Ala Ile Lys Ser Gln	tct gat tca a Ser Asp Ser 1 275	act gtt caa gta gat gtt gta ggt Thr Val Gln Val Asp Val Val Gly 280 285	
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and and the sac	Gln Ile Phe (	gaa aaa gtg aag gct gac ctt gac Glu Lys Val Lys Ala Asp Leu Asp 310 315	963
cca att aac gcc ttc Pro Ile Asn Ala Phe 320	caa ttg cca g Gln Leu Pro G 325	ggt ggt gcc gtt att tcc ggc gg Gly Gly Ala Val Ile Ser Gly Gly 330	1011
tca gct gcc ata cca Ser Ala Ala Ile Pro 335	ggt att gac Gly Ile Asp 340	agc ttg gct gaa gac atc ttc aa; Ser Leu Ala Glu Asp Ile Phe Ly; 345	g 1059 s
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gcc ttc act gtg gca Ala Phe Thr Val Ala 370	gtc ggc ttg Val Gly Leu	acc ctc tac caa gcc cag act tc Thr Leu Tyr Gln Ala Gln Thr Se 375	t 1155 r
gat att gag cgg gcc Asp Ile Glu Arg Ala 385	atc aac cag Ile Asn Gln	tcc atc ttg caa aat atc ggt at Ser Ile Leu Gln Asn Ile Gly Il 390 395	t 1203 e
Asn Pro Asp Ser Gln 400	Pro Ala Asn 405	cgg ata gtt gac cag gat gat to Arg Ile Val Asp Gln Asp Asp Se 410	
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<213> Alloiococcus otitidis

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Lys Gly Leu Lys Arg Gly Met Val Val Asp Ile Asp Ala Thr Val Gln 50 55

Ala Ile His Thr Ala Val Lys Gln Ala Ala Asp Lys Thr Gly Val Met 65 70 75 80

Ile Asn Gln Leu Ile Val Gly Val Pro Ala Asn Gly Val Ser Ile Glu 85 90 95

Pro Cys His Gly Val Ile Thr Val Asp Asp Arg Ser Lys Glu Ile Asp 100 105 110

Ser Gln Glu Val Asn Arg Val Val Asn Gln Ser Ile Ala Asn Ile Val 115 120 125

Pro Pro Asp Arg Asp Leu Leu Ser Val Ser Leu Glu Glu Phe Ile Val 130 135 140

Asp Gly Phe Asp Glu Ile His Asp Pro Arg Gly Met Val Gly Gln Arg 145 150 155 160

Leu Glu Leu Tyr Gly Thr Ala Ile Ser Val Pro Lys Thr Ile Leu His 165 170 175

Asn Ile Arg Cys Val Glu Lys Ala Gly Tyr Gln Ile Ala Ala Leu 180 185 190

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- Ile Leu Gln Pro Gln Ala Met Ala Lys Val Ala Leu Ser Glu Asp Glu 195 200 205
- Arg Asn Phe Gly Thr Val Met Val Asp Ile Gly Gly Gln Thr Thr 210 215 220
- Leu Ser Ala Ile His Asp Glu Gln Val Lys Tyr Ala Asn Val Val Gln 225 230 235
- Glu Ala Gly Glu Tyr Ile Thr Lys Asp Ile Ser Ile Val Ile Asn Thr 245 250 255
- Ser Gln Gln Asn Ala Glu Lys Leu Lys Arg Glu Val Gly Ala Ile Lys 260 265 270
- Ser Gln Ser Asp Ser Thr Val Gln Val Asp Val Val Gly Gln Asn Glu 275 280 285
- Pro Val Lys Ile Lys Glu Ser Tyr Val Gly Glu Ile Ile Glu Ala Arg 290 295 300
- Val Ser Gln Ile Phe Glu Lys Val Lys Ala Asp Leu Asp Pro Ile Asn 305 310 315 320
- Ala Phe Gln Leu Pro Gly Gly Ala Val Ile Ser Gly Gly Ser Ala Ala 325 330 335
- Ile Pro Gly Ile Asp Ser Leu Ala Glu Asp Ile Phe Lys Val Arg Ser 340 345 350
- Glu Leu Tyr Ile Pro Asp Tyr Met Gly Ile Arg Thr Pro Ala Phe Thr 355 360 365
- Val Ala Val Gly Leu Thr Leu Tyr Gln Ala Gln Thr Ser Asp Ile Glu 370 375 380
- Arg Ala Ile Asn Gln Ser Ile Leu Gln Asn Ile Gly Ile Asn Pro Asp 385 390 395 400
- Ser Gln Pro Ala Asn Arg Ile Val Asp Gln Asp Asp Ser Val Gln Ser 405 410 415

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ggg aca ggt ggt tat gtt tgt gcc cct gtc ata tac cag gcg acc aag Gly Thr Gly Gly Tyr Val Cys Ala Pro Val Ile Tyr Gln Ala Thr Lys

tta ggc att cca agt ctc att cac gaa caa aat agt gtc gcc ggg gtg

435

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cag Gln	gaa Glu	gct Ala	gaa Glu 160	aaa Lys	tcc Ser	ttt Phe	gcc Ala	aag Lys 165	tat Tyr	aag Lys	gat Asp	aag Lys	ctg Leu 170	gtt Val	ttg Leu	531
act Thr	ggt Gly	aat Asn 175	cca Pro	aga Arg	gga Gly	cag Gln	gaa Glu 180	gtc Val	agc Ser	caa Gln	gtc Val	aag Lys 185	ggt Gly	ggc Gly	ctt Leu	579
agc Ser	ctc Leu 190	cac His	aag Lys	tat Tyr	Gly ggc	atg Met 195	gac Asp	atg Met	tcc Ser	caa Gln	cct Pro 200	tca Ser	gta Val	att Ile	att Ile	627
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		agt Ser	caa Gln	ctg Leu 225	Ala	gag Glu	agg Arg	gac Asp	tac Tyr 230	GIN	gtc Val	ttg Leu	ttt Phe	gtg Val 235	ccg Pro	723
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GJ7 āāc	cag Glr	cat His	: Lys	ccg Pro	tca Ser	aac Asn	att 11e	? Phe	att	gaa Glu	tco Ser	tat Tyr 265	TIE	gat Asp	aac Asn	819
ato Mei	g ccc E Pro	Gli	a gtt n Val	ttt Phe	aag Lys	gct Ala 275	ı Ile	gac Asp	ttg Lev	gtg ı Val	g gtt L Val 280	Cys	cgt Arg	agt Sei	Gly ggg	867
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- 4-		a ag o Se	t cco	c aat o Asi 30!	n Val	a acq	g gct r Ala	t gad a Ası	c cac p Hi: 310	S GII	a aco	c aaar r Ly	a aat s Asi	gc Ala 31	t atg a Met 5	963
ag Se	t tt r Le	g gt u Va	t aa 1 As: 32	n Gl	a caa n Gli	a gci n Ala	t gg a Gl	c tta y Le 32	u Me	g at	t aa e Ly	g ga s Gl	a aat u Asi 330	1 AS	t cta p Leu	1011
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gc Al	a aa .a Ly	a ag	a aa g As	c aa n Ly	g at s Me	g gc t Al	с са a Gl	a ca n Gl	a gc n Al	g aa a Ly	a ga s Gl	a at u Me	g gg t Gl	c ca y Gl	a ccc n Pro	1107

PCT/US02/36122 WO 03/104391 197/235 360 355 350 caa gct tca gac aag ttg atc gct ctc atc ttg tcc atg gtt aag gaa 1155 Gln Ala Ser Asp Lys Leu Ile Ala Leu Ile Leu Ser Met Val Lys Glu 375 370 1179 gat att aac tca gac atc gat taa Asp Ile Asn Ser Asp Ile Asp 385 <210> 88 <211> 387 <212> PRT <213> Alloiococcus otitidis <400> 88 Met Glu Thr Lys Lys Gln Ala Leu Lys Val Leu Leu Ser Gly Gly Thr Gly Gly His Ile Tyr Pro Ala Leu Ala Leu Ala Lys His Leu Ala 20 Ser Leu His Ser Asp Val Glu Phe Leu Tyr Val Gly Thr Gln Arg Gly 40 Leu Glu Asn Lys Leu Val Pro Gln Ala Gly Leu Asp Phe Ile Pro Ile Lys Val Glu Gly Phe Ser Arg Lys Phe Asn Phe Lys Ser Ile Lys Tyr 65 Asn Thr Lys Ser Leu Ile Tyr Phe Leu Lys Ala Leu Ser Lys Ser Lys 90 Gln Ile Ile Lys Asp Phe Gln Pro Asp Val Val Ile Gly Thr Gly Gly 105 Tyr Val Cys Ala Pro Val Ile Tyr Gln Ala Thr Lys Leu Gly Ile Pro 115

Ser Leu Ile His Glu Gln Asn Ser Val Ala Gly Val Thr Asn Lys Phe

Leu Ala Arg Tyr Val Asp Lys Ile Ala Leu Ser Phe Gln Glu Ala Glu

155

150

Lys Ser Phe Ala Lys Tyr Lys Asp Lys Leu Val Leu Thr Gly Asn Pro 165 170 175

Arg Gly Gln Glu Val Ser Gln Val Lys Gly Gly Leu Ser Leu His Lys 180 185 190

Tyr Gly Met Asp Met Ser Gln Pro Ser Val Ile Ile Phe Gly Gly Ser 195 200 ·205

Arg Gly Ala Tyr Ala Ile Asn Lys Ala Phe Val Glu Ala Tyr Ser Gln 210 215 220

Leu Ala Glu Arg Asp Tyr Gln Val Leu Phe Val Pro Gly Ser Ala Asn 225 230 235

Phe Ser Arg Ile Lys Gln Glu Ile Asp Asn Arg Tyr Gly Gln His Lys 245 250 255

Pro Ser Asn Ile Phe Ile Glu Ser Tyr Ile Asp Asn Met Pro Gln Val 260 265 270

Phe Lys Ala Ile Asp Leu Val Val Cys Arg Ser Gly Ala Thr Thr Leu 275 280 285

Ala Glu Ile Met Ser Leu Gly Leu Ala Ser Ile Leu Ile Pro Ser Pro 290 295 300

Asn Val Thr Ala Asp His Gln Thr Lys Asn Ala Met Ser Leu Val Asn 305 310 315 320

Gln Gln Ala Gly Leu Met Ile Lys Glu Asn Asp Leu Asn Gly Gln Ser 325 330 335

Leu Leu Asn Cys Leu Asp Asp Leu Met His Asp Asp Ala Lys Arg Asn 340 345 350

Lys Met Ala Gln Gln Ala Lys Glu Met Gly Gln Pro Gln Ala Ser Asp 355 360 365

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Asp Ile Asp

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ttg tta aaa aaa cta ggg gcc aag gtc atc gtc aat gac aag ttg gcc Leu Leu Lys Lys Leu Gly Ala Lys Val Ile Val Asn Asp Lys Leu Ala 30 35 40	147
cta gaa aat aat acg gaa gcc cag gtc tta att gaa gag ggc ttc caa Leu Glu Asn Asn Thr Glu Ala Gln Val Leu Ile Glu Glu Gly Phe Gln 45 50 55	195
gtt atc acc ggc tac cac cca gag gat tta ctt gat gca agc ttt gac Val Ile Thr Gly Tyr His Pro Glu Asp Leu Leu Asp Ala Ser Phe Asp 60 65 70	243
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cag gct gaa aaa ctg gct att ccc att tta act gaa gtg gac gtg gca Gln Ala Glu Lys Leu Ala Ile Pro Ile Leu Thr Glu Val Asp Val Ala 90 95 100	339
gga agc atc tta aaa gcc aag ccc atc gct gtt acc ggg acc aat ggc Gly Ser Ile Leu Lys Ala Lys Pro Ile Ala Val Thr Gly Thr Asn Gly 110 115	387
aag aca act acc gta tct tta att tat gat att tta gcc caa gat caa Lys Thr Thr Thr Val Ser Leu Ile Tyr Asp Ile Leu Ala Gln Asp Gln 125 130	435
gcg gaa agc cct gaa cct aaa cca gtc tac aag cta ggc aat att ggc Ala Glu Ser Pro Glu Pro Lys Pro Val Tyr Lys Leu Gly Asn Ile Gly 140 145 150	483
caa ccg gtt agt gac ttg gcc tta gaa att aaa gct gaa tct aac ctg Gln Pro Val Ser Asp Leu Ala Leu Glu Ile Lys Ala Glu Ser Asn Leu 155 160 165	531

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cct Pro	cat His	ata Ile	gca Ala	gtc Val 190	att Ile	acc Thr	aat Asn	att Ile	tat Tyr 195	tcc Ser	gcc Ala	cac His	ctt Leu	gac Asp 200	tac Tyr	627
cat His	aag Lys	agt Ser	cgg Arg 205	gag Glu	gaa Glu	tat Tyr	gtt Val	agg Arg 210	gct Ala	aag Lys	cta Leu	agg Arg	att Ile 215	acc Thr	cag Gln	675
gct Ala	caa Gln	ggt Gly 220	ccg Pro	gat Asp	gac Asp	tac Tyr	cta Leu 225	gtc Val	tac Tyr	tac Tyr	cag Gln	ggt Gly 230	cag Gln	gaa Glu	gaa Glu	723
ttg Leu	gct Ala 235	agc Ser	ctg Leu	gtc Val	aaa Lys	aaa Lys 240	tac Tyr	tct Ser	aaa Lys	gcc Ala	cag Gln 245	ctg Leu	gtc Val	ccc Pro	tat Tyr	771
act Thr 250	gac Asp	aag Lys	ggt Gly	caa Gln	ctg Leu 255	aac Asn	caa Gln	gga Gly	gcc Ala	tat Tyr 260	Ile	aag Lys	gat Asp	gac Asp	tat Tyr 265	819
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aaa Lys	ata Ile	aag Lys 300	Gly	ctc Leu	tct Ser	aac Asn	caa Gln 305	Thr	att	gcc Ala	caa Gln	gct Ala 310	. vaı	aac Asn	cac	963
tto Phe	aaa Lys 315	Gly	gtt Val	gcc Ala	cac His	cgc Arg 320	Ser	cag Gln	gtg Val	gtt Val	ggg Gly 325	Arg	tat Tyr	gag Glu	gac Asp	1011
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cta Lei	a gad ı Ası	c cgc	caa g Glr 365	n Ası	gat Ası	ttt Phe	tco Ser	aag Lys	: Le	gad 1 Asi	cat His	gct Ala	cta Lev 375	ı Ası	agg n Arg	1155
gt! Va	t aaq l Ly:	g ggg s Gly 380	y Vai	gtt l Val	t tgt l Cy:	t ttt s Phe	ggc Gly 385	7 Gli	aco n Thi	c aaa r Lys	a gat s Asj	t aag p Ly: 390	s re	agco aAla	cgg a Arg	1203

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cct gaa gca gtt gat Pro Glu Ala Val Asp 410	ttg gct tac g Leu Ala Tyr : 415	gac ttg agt gag cca gga caa gtc Asp Leu Ser Glu Pro Gly Gln Val 420 425	1299
att tta ttt tct cct Ile Leu Phe Ser Pro 430	Ala Cys Ala	agt tgg gac caa tat gct aac ttt Ser Trp Asp Gln Tyr Ala Asn Phe 435 440	1347
gaa gag aga gga caa Glu Glu Arg Gly Gln 445	gat tat gtt Asp Tyr Val	gat gca atc cag cag ctg gtt gaa Asp Ala Ile Gln Gln Leu Val Glu 450 455	1395
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Gln Val Leu Ile Gl 50	ú Glu Gly Ph∈ 55	e Gln Val Ile Thr Gly Tyr His Pro 60	
Glu Asp Leu Leu As 65	sp Ala Ser Phe 70	e Asp Phe Val Val Lys Asn Pro Gly 75 80	
Ile Pro Tyr Thr A:		l Gly Gln Ala Glu Lys Leu Ala Ile 90 95	
Pro Ile Leu Thr G 100	lu Val Asp Va	l Ala Gly Ser Ile Leu Lys Ala Lys 105 110	
Pro Ile Ala Val T 115	hr Gly Thr As 12	on Gly Lys Thr Thr Thr Val Ser Leu 20 125	

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- Ile Tyr Asp Ile Leu Ala Gln Asp Gln Ala Glu Ser Pro Glu Pro Lys
- Pro Val Tyr Lys Leu Gly Asn Ile Gly Gln Pro Val Ser Asp Leu Ala
- Leu Glu Ile Lys Ala Glu Ser Asn Leu Val Val Glu Leu Ser Ser Phe 165
- Gln Leu Gln Ser Leu Thr Tyr Phe Thr Pro His Ile Ala Val Ile Thr 185
- Asn Ile Tyr Ser Ala His Leu Asp Tyr His Lys Ser Arg Glu Glu Tyr 205 200
- Val Arg Ala Lys Leu Arg Ile Thr Gln Ala Gln Gly Pro Asp Asp Tyr 210
- Leu Val Tyr Tyr Gln Gly Gln Glu Leu Ala Ser Leu Val Lys Lys 230
- Tyr Ser Lys Ala Gln Leu Val Pro Tyr Thr Asp Lys Gly Gln Leu Asn 250
- Gln Gly Ala Tyr Ile Lys Asp Asp Tyr Leu Ile Tyr Asn Gln Glu Pro 265 260
- Val Met Ala Leu Asp Arg Val Gln Val Ser Gly Ser His Asn Leu Gln 275
- Asn Ile Leu Ala Ala Val Cys Val Ala Lys Ile Lys Gly Leu Ser Asn 295
- Gln Thr Ile Ala Gln Ala Val Asn His Phe Lys Gly Val Ala His Arg 315
- Ser Gln Val Val Gly Arg Tyr Glu Asp Arg Leu Phe Val Asn Asp Ser 330 335
- Lys Ala Thr Asn Ser Leu Ala Thr Gln Lys Ala Leu Glu Ala Tyr Asp 345 340

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Gln Asp Thr Ile Leu Leu Val Gly Gly Leu Asp Arg Gln Asp Asp Phe 355 Ser Lys Leu Asp His Ala Leu Asn Arg Val Lys Gly Val Val Cys Phe 375 Gly Gln Thr Lys Asp Lys Leu Ala Arg Tyr Phe Lys Asp Arg His Ile 395 Glu Gly Val Glu Leu Ala Gln Thr Val Pro Glu Ala Val Asp Leu Ala 405 Tyr Asp Leu Ser Glu Pro Gly Gln Val Ile Leu Phe Ser Pro Ala Cys 420 Ala Ser Trp Asp Gln Tyr Ala Asn Phe Glu Glu Arg Gly Gln Asp Tyr 445 440 Val Asp Ala Ile Gln Gln Leu Val Glu Arg Leu Glu Gln Arg Ser Lys 455 450 Tyr Gly Asn 465 <210> 91 <211> 651 <212> DNA <213> Alloiococcus otitidis <220> <221> CDS <222> (7)..(651) <223> <400> 91 actagt atg aag caa aaa act caa gcg aca gcg gtc aac cag acc caa 48 Met Lys Gln Lys Thr Gln Ala Thr Ala Val Asn Gln Thr Gln 5 1 aca gag gca gaa gaa aga caa gaa acc cgt cgg aaa att ggc ctc atg 96 Thr Glu Ala Glu Glu Arg Gln Glu Thr Arg Arg Lys Ile Gly Leu Met 20 15 ggg ggg acc ttt aat ccg ccc cat ctg ggt cat tta ctg gta gct gaa 144 Gly Gly Thr Phe Asn Pro Pro His Leu Gly His Leu Leu Val Ala Glu 40 caa gtt tat gag gcc ttg gac ttg gat aat att cac ttt atg ccc act 192

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cgg Arg	gtt Val 80	gat Asp	atg Met	gtg Val	gat Asp	tat Tyr 85	gcc Ala	atc Ile	gaa Glu	gat Asp	aac Asn 90	ccc Pro	cac His	ttt Phe	tct Ser	288
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acc Thr	att Ile	aaa Lys	gaa Glu	ttg Leu 115	aaa Lys	gag Glu	gct Ala	agc Ser	ccg Pro 120	aat Asn	aca Thr	gat Asp	tat Tyr	tac Tyr 125	ttc Phe	384
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gaa Glu	caa Gln	tta Leu 145	Leu	gat Asp	tta Leu	gtt Val	caa Gln 150	ttt Phe	gtt Val	ggt Gly	gtg Val	aag Lys 155	Arg	cca Pro	Gly	480
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Tyr Glu Ala Leu Asp Leu Asp Asn Ile His Phe Met Pro Thr Ala Lys 50 55

Pro Gly His Ala Ala Gly Lys Glu Thr Ile Asp Ala Ser Tyr Arg Val 65 70 75 80

Asp Met Val Asp Tyr Ala Ile Glu Asp Asn Pro His Phe Ser Leu Asn 85 90 95

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Lys Glu Leu Lys Glu Ala Ser Pro Asn Thr Asp Tyr Tyr Phe Ile Ile 115 120 125

Gly Glu Asp Ser Val Met Asp Leu Ala Gln Trp Lys Asn Ile Glu Gln 130 135 140

Leu Leu Asp Leu Val Gln Phe Val Gly Val Lys Arg Pro Gly Tyr Gln 145 150 155 160

Ala Asp Val Asp Phe Pro Ile Ile Trp Val Asp Thr Pro Glu Leu Asp 165 170 175

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Val Val Asp Ala Asp Arg Val Ala Arg Gln Val Val Glu Pro Gly Ser 55

Pro Gly Leu Asp Gln Leu Val Asp Tyr Phe Gly Gln Glu Ile Leu Thr 70 65

Gln Asp Gly Gly Leu Asp Arg Lys Tyr Leu Gly Asp Leu Ile Phe Arg 85

Asn Ser Gln Ala Lys Glu Ala Val Asn Arg Ile Leu His Pro Leu Ile 105

Arg Gln Ser Ile Gln Asn Gln Ile Lys Thr Ala Ile Gly Gln Asp Leu

Asp Leu Leu Val Leu Asp Ile Pro Leu Leu Tyr Glu Thr Gly Gln Ala 135

Asp Asp Tyr Gln Ala Val Met Val Val Ser Leu Pro Tyr Gln Asp Gln 150 145

Val Ser Arg Leu Met Asp Arg Asp Gly Ile Asp Arg Asp Gln Ala Leu 170 165

Arg Lys Ile Gln Ala Gln Met Ser Leu Glu Glu Lys Val Lys Leu Ala 190 185

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caa gga tct gat Gln Gly Ser Asp 35	gtc aaa aag Val Lys Lys	tat ttc to Tyr Phe Pl 40	tt acc caa aaa a he Thr Gln Lys S 4	gc tta gaa 144 Ger Leu Glu 25
gaa aaa aat ata Glu Lys Asn Ile 50	aac att tta Asn Ile Lev	a gaa ttt g a Glu Phe A 55	ac cct gat aac a sp Pro Asp Asn 1 60	itc aaa cca 192 Ile Lys Pro
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gag aag gtg gtc ggg gac act act att atc gat gac tat gct cac cac

Glu Lys Val Val Gly Asp Thr Thr Ile Ile Asp Asp Tyr Ala His His

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	Il∈	e Gly	y As <u>ı</u>	) Let		e Glu	ı His	Phe	Thr 105		: Ile	e Ala	a Ile	Th:	Gly	y Ser	•		
	His	s Gly	y Lys 11:	_	r Sei	Thi	Thi	Gly 120		ı Met	: Ala	a Hi:	s Val	L Phe	e Se:	r Gly	-		

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230 225

Arg Glu Gly Ser Ala Phe Asp Leu Tyr Ile Lys Gly Glu Phe Tyr Lys 245

His Phe Thr Ile Pro Thr Tyr Gly Asn His Asn Ile Gln Asn Ala Leu 265

Ala Val Ile Ala Val Ala Tyr Tyr Glu Gly Leu Asp Gln Asp Leu Val 280

Ala Gln Arg Leu Ala Asn Phe Ala Gly Val Lys Arg Arg Phe Thr Glu

Lys Val Val Gly Asp Thr Thr Ile Ile Asp Asp Tyr Ala His His Pro 310 305

Ala Glu Ile Arg Ala Thr Ile Asp Ala Ala Arg Gln Lys Tyr Pro Asp 325 330

Lys Asp Ile Val Thr Val Phe Gln Pro His Thr Phe Thr Arg Thr Val

Ala Leu Leu Asp Glu Phe Ala Gln Ala Leu Asp Leu Ala Asp Gln Val · 365 · 360 355

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80

85

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Lys Lys Ser Phe Tyr Met Tyr Ala Ala Ser Ser Asn Lys Lys Arg Asn 260 265 270

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gag Glu	gcc Ala	atg Met	gac Asp 190	ttt Phe	gac Asp	gac Asp	ctg Leu	att Ile 195	atg Met	caa Gln	acc Thr	gtc Val	cgt Arg 200	ctc Leu	ttc Phe		627
aag Lys	gaa Glu	aag Lys 205	Pro	gat Asp	acc Thr	ctg Leu	tct Ser 210	Tyr	tac Tyr	cag Gln	gcc Ala	aag Lys 215	ttc Phe	cag Gln	tat Tyr		675
atc Ile	cat His	gtt Val	gac Asp	gaa Glu	tac Tyr	cag Gln	gat Asp	acc Thr	aac Asn	caa Gln	gcc Ala	caa Gln	tac Tyr	caa Gln	ctg Leu		723

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gct Ala	gac Asp	cag Gln	tct Ser	att Ile 255	tat Tyr	ggt Gly	tgg Trp	cgg Arg	ggg Gly 260	gct Ala	gat Asp	atg Met	gga Gly	aat Asn 265	att Ile	819
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caa Gln	aat Asn	tac Tyr 285	cgg Arg	tca Ser	acc Thr	aag Lys	tct Ser 290	ata Ile	atc Ile	agg Arg	gca Ala	gcc Ala 295	aat Asn	gat Asp	gtt Val	915
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gac Asp	aag Lys	tta Leu	cgc Arg 430	cag Gln	gct Ala	ggc Gly	cag Gln	gag Glu 435	atg Met	ggt Gly	tgg Trp	tcg Ser	ctt Leu 440	tac Tyr	gaa Glu	1347
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aga Arg	cta Leu 460	tta Leu	gac Asp	ttc Phe	agt Ser	caa Gln 465	atg Met	att Ile	gaa Glu	aat Asn	ttc Phe 470	agg Arg	aaa Lys	atg Met	acg Thr	1443
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									tcc Ser							1587
									tta Leu							1635
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		WO 0	3/104	391						219/2	235					PCT/US02/36122
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aaa Lys	aaa Lys 700	tgg Trp	ggc	caa Gln	gga Gly	acc Thr 705	att Ile	att Ile	gag Glu	att Ile	aaa Lys 710	ggt Gly	tct Ser	ggc Gly	tcg Ser	2163
gac Asp 715	ctc Leu	cag Gln	ctc Leu	aac Asn	att Ile 720	gcc Ala	ttt Phe	cca Pro	Asp	gaa Glu 725	GJA āāā	atc Ile	aag Lys	ccc Pro	ttg Leu 730	2211
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Gln	Gly	· Gly	Ser	Gly 85	Val	Trp	Val	Ser	Thr 90	Phe	His	Ser	Met	Cys 95	Val	
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Thr Ile Ala Asp Pro Ser Glu Gln Lys Ser Leu Met Lys Gln Val Leu 115 120 125

Lys Asp Leu Asm Ile Asp Pro Lys Arg Tyr Asm Pro Lys Ala Ile Leu

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Ala Glu Ile Ser Asn Ala Lys Asn Asp Leu Leu Asp Glu Gln Thr Tyr 150 Arg Lys Gln Ala Asp Asp Tyr Phe Lys Glu Val Val Ala Asp Cys Tyr 175 165 170 Asp Ala Tyr Gln Arg Gln Leu Arg Gln Ser Glu Ala Met Asp Phe Asp 185 180 Asp Leu Ile Met Gln Thr Val Arg Leu Phe Lys Glu Lys Pro Asp Thr Leu Ser Tyr Tyr Gln Ala Lys Phe Gln Tyr Ile His Val Asp Glu Tyr 215 210 220 Gln Asp Thr Asn Gln Ala Gln Tyr Gln Leu Val Gln Leu Leu Ala Gln Arg Phe Lys Asn Val Cys Val Val Gly Asp Ala Asp Gln Ser Ile Tyr 250 Gly Trp Arg Gly Ala Asp Met Gly Asn Ile Leu Asn Phe Glu Lys Asp 260 265 . 270 Tyr Pro Glu Ala Gln Thr Ile Phe Leu Glu Gln Asn Tyr Arg Ser Thr 275 280 285 Lys Ser Ile Ile Arg Ala Ala Asn Asp Val Ile Gln Asn Asn Ile Asn Arg Arg Asp Lys Asn Leu Trp Thr Ala Asn Asp Glu Gly Asp Lys Val 310 Ser Leu Tyr Ala Ala Arg Ser Glu Gln Asp Glu Ala Gln Phe Ile Val Gly Thr Ile His Asp Leu Thr Glu Gly Lys Lys Ala Gly Tyr Gly Asp 345 Ile Ala Ile Leu Tyr Arg Thr Asn Ala Met Ser Arg Val Ile Glu Glu

360

365

355

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Thr Phe Ile Lys Ser Asn Ile Pro Tyr Lys Ile Val Gly Gly Thr Gly 370 375 380

Phe Tyr Gln Arg Lys Glu Ile Arg Asp Leu Ile Ala Tyr Leu Thr Leu 385 390 395 400

Val Ala Asn Pro Ala Asp Asp Leu Ser Phe Ser Arg Ile Val Asn Glu 405 410 415

Pro Lys Arg Gly Ile Gly Pro Gly Thr Leu Asp Lys Leu Arg Gln Ala 420 425 430

Gly Gln Glu Met Gly Trp Ser Leu Tyr Glu Thr Ala Leu Asn Ala Asp 435 440 445

Ala Thr Asn Leu Pro Ser Arg Ala Val Asn Arg Leu Leu Asp Phe Ser 450 450 460

Gln Met Ile Glu Asn Phe Arg Lys Met Thr Glu Tyr Leu Pro Ile Thr 465 470 475 480

Asp Leu Thr Glu Lys Ile Leu Glu Asp Thr Gly Tyr Gln Lys Ala Leu 485 490 495

Glu Lys Asp Arg Thr Leu Glu Ser Gln Ala Arg Leu Glu Asn Leu Gln 500 505 510

Glu Phe Tyr Ser Val Thr Glu Glu Phe Asp Glu Glu Asp Asp Asn 515 520 525

Lys Ser Leu Leu Ala Phe Leu Thr Asp Leu Ser Leu Leu Ser Pro Ala 530 540

Asp Asp Val Glu Glu Gly Arg Gly Gln Val Thr Met Met Thr Leu His 545 550 550 555

Ala Ala Lys Gly Leu Glu Phe Pro Tyr Val Phe Ile Ala Gly Met Glu
565 570 575

Glu Gly Ile Phe Pro Leu Ser Arg Ala Ala Glu Asp Pro Glu Ser Leu 580 585 590

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Glu Glu Glu Arg Arg Leu Ala Tyr Val Gly Ile Thr Arg Ala Glu Gln
595 600 605

Ala Leu Tyr Leu Thr Arg Ala Met Met Arg Gln Leu Tyr Gly Arg Thr 610 620

Gln Ala Asn Pro Lys Ser Arg Phe Leu Ser Glu Ile Ser Ser Asp Leu 625 630 635 640

Val Gln Asp Leu Gly Ala Thr Thr Gly Ser Leu Ser Gln Thr Gly Gly
645 650 655

Lys Val Ser Pro Arg Leu Gly Gly Arg Lys Ala Ser Gly Tyr Lys Ala 660 665 670

Asn Ala Trp Ser Gln Gln Ser Val Gly Ala Thr Gly Ala Glu Lys Glu 675 680 685

Asp Trp Glu Val Gly Asp Lys Val His His Lys Lys Trp Gly Gln Gly 690 695 700

Thr Ile Ile Glu Ile Lys Gly Ser Gly Ser Asp Leu Gln Leu Asn Ile 705 · 710 715 720

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aca gtc aag gta gaa ggg gct aag aat gct gcc ctt cct atc ctg gct
Thr Val Lys Val Glu Gly Ala Lys Asn Ala Ala Leu Pro Ile Leu Ala

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					Asp								gat Asp		240
													aag Lys		288
													ggt Gly 110		336
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_													gaa Glu		432
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_								_					att Ile		528
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													GJÅ aaa		672
			-	_							_	-	 ata Ile		720
													gtc Val		768

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agt g Ser G															864
atg g Met G	ly I					_	_		_						912
cct g Pro G 3	-				_	_	_		-	-		-	-		960
cta g Leu A 320	_			_	_		_	-	-	_	_		-	-	1008
ttc a Phe M									-						1056
gat g Asp G															1104
aga g Arg V	al (					-		_		-		_			1152
ggt t Gly L 3	_	_	-	_		_			_						1200
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ctc a Leu L					taa										1314

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<400> 102

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Gly Thr Phe Met Val Ala Ala Gly Val Thr Gln Gly Asn Val Leu Ile 245 250 255

Glu Asp Cys Ile Val Glu His Asn Arg Pro Leu Ile Ser Lys Leu Ser 260 265 270

Glu Met Gly Val Gln Phe Glu Glu Glu Lys Thr Gly Leu Arg Val Met 275 280 285

Gly Pro Glu Thr Leu Gln Ala Thr Asp Val Lys Thr Leu Pro Tyr Pro 290 295 300

Gly Phe Pro Thr Asp Met Gln Ser Pro Met Thr Val Ala Gln Thr Leu 305 310 315 320

Ala Glu Gly Arg Ser Ile Met Arg Glu Thr Val Phe Glu Asn Arg Phe 325 330 335

Met His Met Glu Glu Leu Arg Lys Met Asp Ala Gln Phe Thr Val Asp 340 345 350

Gly Gln Ser Leu Ile Ile Glu Gly Gly Lys Lys Leu Gln Gly Ala Arg 355 360 365

Val Gln Ser Ser Asp Leu Arg Ala Ser Ala Ser Leu Ile Ile Ala Gly 370 375 380

Leu Val Ala Asp Gly Val Thr Lys Val Thr Asn Leu Asn His Leu Asp 385 390 395 400

Arg Gly Tyr Tyr Lys Phe His Glu Lys Leu Gln Gln Leu Gly Ala Ser 405 410 415

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Lys Lys Gly Glu 435

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tcg Ser	att Ile	agg Arg 90	gta Val	gac Asp	cgg Arg	gta Val	cga Arg 95	cag Gln	gtc Val	aag Lys	gat Asp	gcc Ala 100	cta Leu	agc Ser	aag Lys	342
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ccg Pro	gca Ala	GJA GGG	Asp	Val	act Thr	att Ile	Phe	ttg Leu	Leu	Val	act Thr	agc Ser	Arg	caa Gln 150	aac Asn	486
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	aag Lys															678
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	tta Leu 265															870
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	gcc Ala															1014
_	ata Ile									•						1026
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Arg Ile Leu Ala Asn His Ala Leu Lys His Ala Tyr Leu Phe Glu Gly 20 25 30

Leu Ala Gly Ser Gly Lys Leu Glu Met Ser Arg Tyr Ile Ala Lys Arg 35 40 45

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Leu Phe Cys Pro Asn Gln Asp Gln Gly Gln Ala Cys Gln Val Cys Pro Thr Cys Leu Arg Ile Asp Gln Gly Gln His Pro Asp Val Val Glu Ile Ala Pro Glu Gly Lys Gly Arg Ser Ile Arg Val Asp Arg Val Arg Gln 90 Val Lys Asp Ala Leu Ser Lys Ser Gly Val Glu Ser Gln Lys Lys Met Ile Ile Leu Asn Gln Ala Asp Lys Met Thr Pro Ser Ala Ala Asn Ser 120 Leu Leu Lys Phe Leu Glu Glu Pro Ala Gly Asp Val Thr Ile Phe Leu 135 130 Leu Val Thr Ser Arg Gln Asn Leu Leu Pro Thr Ile Val Ser Arg Cys 150 155 145 Gln Val Ile Gln Phe Ala Lys Gln Asp Leu Lys Thr Arg Ile Glu Asp 170 Leu Val Glu Ala Gly Leu Ser Gln Glu Glu Ala His Leu Ala Ser His Leu Ser Gln Asp Leu Asp Leu Ala Lys Ser Leu Ile Glu Glu Asp 200 Leu Leu Ala Val Ser Gln Lys Ile Trp Gln Trp Phe Ser Tyr Leu Met 215 Asn Gln Asp Asp Leu Ala Phe Ile Leu Val Gln Arg Asp Leu Met Ala 225 230 235 Phe Ile Gln Asp Arg Asp Asp Cys Gln Met Val Cys Asp Leu Ile Leu 245 Tyr Leu Phe Gln Asp Leu Leu His Leu His Tyr His Leu Asp Ser Pro 260

Ala Cys Phe Ala Gly His Glu Ser Asp Leu Arg Tyr Phe Met Asp Leu 275 280 285	
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agt ttt gct gat gta tcc ggc cag cat gtg gtc acc aag acc cta aag Ser Phe Ala Asp Val Ser Gly Gln His Val Val Thr Lys Thr Leu Lys 15 20 25  aat gcc att aaa aat gat aat acc agt cat gcc tac ctg ttt act gga Asn Ala Ile Lys Asn Asp Asn Thr Ser His Ala Tyr Leu Phe Thr Gly 30 35 40 45  ccc cgg ggg acg ggc aag acc agt gtg gca aaa ata ttt gcc aag gcc Pro Arg Gly Thr Gly Lys Thr Ser Val Ala Lys Ile Phe Ala Lys Ala	147
agt ttt gct gat gta tcc ggc cag cat gtg gtc acc aag acc cta aag Ser Phe Ala Asp Val Ser Gly Gln His Val Val Thr Lys Thr Leu Lys 15 20 25  aat gcc att aaa aat gat aat acc agt cat gcc tac ctg ttt act gga Asn Ala Ile Lys Asn Asp Asn Thr Ser His Ala Tyr Leu Phe Thr Gly 30 45  ccc cgg ggg acg ggc aag acc agt gtg gca aaa ata ttt gcc aag gcc Pro Arg Gly Thr Gly Lys Thr Ser Val Ala Lys Ile Phe Ala Lys Ala 50 55 60  att aat tgc ccc tac tcg gat gat ggg gag cct tgt aat gaa tgt cag Ile Asn Cys Pro Tyr Ser Asp Asp Gly Glu Pro Cys Asn Glu Cys Gln	147 195
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								cct Pro								867
		_			-		_	ctt Leu				_			-	915
								tta Leu 310								963
								gcc Ala								1011
								cta Leu								1059

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caa Gln	aac Asn	ttg Leu 400	caa Gln	gct Ala	gga Gly	gcc Ala	aaa Lys 405	caa Gln	Gly ggg	cct Pro	aag Lys	caa Gln 410	aga Arg	gct Ala	aag Lys	1251
tca Ser	aaa Lys 415	gct Ala	ggc Gly	ccc Pro	aag Lys	caa Gln 420	tct Ser	ggc	cct Pro	GJA GGC	aag Lys 425	tct Ser	aga Arg	agc Ser	cac His	1299
cgt Arg 430	cac His	cag Gln	caa Gln	ggc	ttc Phe 435	aag Lys	gtt Val	aac Asn	cgg Arg	aaa Lys 440	gcc Ala	gtt Val	tac Tyr	tct Ser	atc Ile 445	1347
ttg Leu	gac Asp	cag Gln	gcg Ala	acc Thr 450	cgt Arg	aaa Lys	gac Asp	ctg Leu	gac Asp 455	gac Asp	ctc Leu	caa Gln	gac Asp	ctc Leu 460	tgg Trp	1395
cca Pro	gac Asp	ttg Leu	atc Ile 465	Asn	gtc Val	ttg Leu	acc Thr	atc Ile 470	Ser	caa Gln	aag Lys	gct Ala	atc Ile 475	tta Leu	aac Asn	1443
aat Asn	tcc Ser	aaa Lys 480	Pro	gtt Val	gct Ala	gct Ala	agt Ser 485	Pro	gag Glu	ggt Gly	ttg Leu	gtg Val 490	gtg Val	acc Thr	ttt Phe	1491
gaa Glu	tat Tyr 495	Asp	att Ile	cta Leu	tgt Cys	gag Glu 500	Arg	gca Ala	gag Glu	tct Ser	gac Asp 505	gag Glu	acc Thr	ttg Leu	caa Gln	1539
acg Thr 510	Ala	atc Ile	ggc Gly	aat Asn	tac Tyr 515	Ile	gaa Glu	aaa Lys	att Ile	atc Ile 520	Gly	cgc	cgt Arg	cca Pro	aga Arg 525	1587
ctg Leu	gtc Val	tgt Cys	gtg Val	r cct Pro 530	Glu	gac Asp	aag Lys	tgg Trp	ccg Pro 535	Thr	atc Ile	cgc	cgc	gat Asp 540	ttt Phe	1635
atc Ile	aag Lys	cag Gln	atg Met 545	Lys	aaa Lys	gaa Glu	gat Asp	ggc Gly 550	Ser	act Thr	aaa Lys	gct	ggc Gly 555	Glr	gca Ala	1683
agt Ser	gac Asp	ggo Gly 560	/ Lys	tcg Ser	gat Asp	gat Asp	gac Asp 565	Pro	ggt Gly	caa Gln	gaa Glu	gac Asp 570	Asn	cag Glr	gcc Ala	1731

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ctt aac aag gct gtg gag ctt ttc ggt aaa gac aat att aca atc aaa 1779 Leu Asn Lys Ala Val Glu Leu Phe Gly Lys Asp Asn Ile Thr Ile Lys

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gat taa 1785

Asp 590

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<211> 590

<212> PRT

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<400> 106

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Asp Val Ser Gly Gln His Val Val Thr Lys Thr Leu Lys Asn Ala Ile  $20 \hspace{1cm} 25 \hspace{1cm} 30$ 

Lys Asn Asp Asn Thr Ser His Ala Tyr Leu Phe Thr Gly Pro Arg Gly 35 40 45

Thr Gly Lys Thr Ser Val Ala Lys Ile Phe Ala Lys Ala Ile Asn Cys 50 55

Pro Tyr Ser Asp Asp Gly Glu Pro Cys Asn Glu Cys Gln Ile Cys Gln 65 70 75 80

Glu Ile Thr Gln Gly Ser Leu Gly Asp Val Ile Glu Ile Asp Ala Ala 85 90 95

Ser Asn Asn Gly Val Glu Glu Ile Arg Asp Ile Arg Glu Lys Ala Asn 100 105 110

Tyr Ala Pro Thr Ser Ala Val Tyr Lys Val Tyr Ile Ile Asp Glu Val 115 120 125

His Met Leu Ser Ser Gly Ala Phe Asn Ala Leu Leu Lys Thr Leu Glu 130 135 140

Glu Pro Pro Ala Asn Val Val Phe Ile Leu Ala Thr Thr Glu Pro His 145 150 155 160

Lys Ile Pro Ala Thr Ile Ile Ser Arg Thr Gln Arg Phe Asp Phe Lys
165 170 175

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Arg	Ile	Asp	Asn 180	Gln	Asp	Ile	Ile	Asp 185	Arg	Leu	Ile	Tyr	Ile 190	Leu	Glu
Glu	Asp	Gln 195	Val	Pro	Tyr	Ser	Lys 200	Glu	Ala	Val	Leu	Ser 205	Leu	Ala	Asn
Ala	Ala 210	Glu	Gly	Gly	Met	Arg 215	Asp	Ala	Leu	Ser	Met 220	Leu	Asp	Gln	Ala
Leu 225		Phe	Met	Thr	Asp 230	Glu	Leu	Thr	Glu	Glu 235	Val	Ala	Leu	Gln	Ile 240
Thr	Gly	Ser	Ile	Thr 245	Gln	Ser	Leu	Leu	Leu 250	Glu	Tyr	Leu	Gln	Val 255	Ile
Ser	Gln	Gly	Gln 260	Thr	Glu	Glu	Gly	Leu 265	Lys	Leu	Leu	Gln	Glu 270	Val	Leu
Gly	Glu	Gly 275	Lys	Asp	Pro	Ser	Arg 280	Phe	Val	Glu	Asp	Ala 285	Ile	Met	Met
Thr	Arg 290	Asp	Leu	Leu	Leu	Tyr 295	Gln	Thr	Ser	Gln	Gly 300	Asp	Asn	Phe	Val
Pro 305	Lys	Leu	Ala	Arg	Leu 310	Asp	Asp	Gln	Phe	Glu 315	Asp	Leu	Ala	Lys	Asp 320
Leu	Asp	Lys	Glu	Met 325	Ala	Tyr	His	Ile	Ile 330	Asp	Val	Leu	Asn	Gln 335	Thr
Gln	Asp	Asp	Leu 340	Arg	Leu	Ser	Asn	His 345	Gly	Glu	Val	Tyr	Leu 350	Glu	Ile
Ala	Thr	Val 355	_	Leu ′	Ser		Pro 360		Ser	Ala	Val	Gln 365		Ile	Gln
Ala	Ser 370	Gln	Val	Asn	Met	Val 375	Asp	Gln	Asp	Asn	Lys 380	Glu	Glu	Ile	Ala
Gln 385	Leu	Gln	Asn	Gln	Val 390	Lys	Ser	Leu	Gln	Gln 395	Ser	Ile	Gln	Asn	Leu 400

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Gln Ala Gly Ala Lys Gln Gly Pro Lys Gln Arg Ala Lys Ser Lys Ala

Gly Pro Lys Gln Ser Gly Pro Gly Lys Ser Arg Ser His Arg His Gln

Gln Gly Phe Lys Val Asn Arg Lys Ala Val Tyr Ser Ile Leu Asp Gln

Ala Thr Arg Lys Asp Leu Asp Asp Leu Gln Asp Leu Trp Pro Asp Leu

Ile Asn Val Leu Thr Ile Ser Gln Lys Ala Ile Leu Asn Asn Ser Lys 470

Pro Val Ala Ala Ser Pro Glu Gly Leu Val Val Thr Phe Glu Tyr Asp 485

Ile Leu Cys Glu Arg Ala Glu Ser Asp Glu Thr Leu Gln Thr Ala Ile

Gly Asn Tyr Ile Glu Lys Ile Ile Gly Arg Arg Pro Arg Leu Val Cys

Val Pro Glu Asp Lys Trp Pro Thr Ile Arg Arg Asp Phe Ile Lys Gln 535

Met Lys Lys Glu Asp Gly Ser Thr Lys Ala Gly Gln Ala Ser Asp Gly 545 555

Lys Ser Asp Asp Pro Gly Gln Glu Asp Asn Gln Ala Leu Asn Lys 565

Ala Val Glu Leu Phe Gly Lys Asp Asn Ile Thr Ile Lys Asp 585